Review

New insights in gastrointestinal "pediatric" neoplasms in adult patients: pancreatoblastoma, hepatoblastoma and embryonal sarcoma of the liver. A practical approach by GIPPI-GIPAD Groups

Vassilena Tsvetkova¹, Gaetano Magro², Giuseppe Broggi², Claudio Luchini¹, Filippo Cappello³, Chiara Caporalini⁴, Anna Maria Buccoliero⁴, Luisa Santoro³

¹ Department of Diagnostics and Public Health, Section of Pathology, Verona University and Hospital Trust, Verona, Italy; ² Department of Medical and Surgical Sciences and Advanced Technologies "G. F. Ingrassia", Anatomic Pathology, University of Catania, Catania, Italy; ³ Department of Pathology, Azienda Ospedaliera Universitaria di Padova, Padova, Italy; ⁴ Pathology Unit, Meyer Children Hospital, Florence, Italy

Summary

Pediatric solid neoplasms are rare and very different from those observed in adults. The majority of them are referred to as embryonal because they arise as a result of alterations in the processes of organogenesis or normal growth and are characterized by proliferation of primitive cells, reproducing the corresponding tissue at various stages of embryonic development. This review will focus on embryonal gastrointestinal pediatric neoplasms in adult patients, including pancreatoblastoma, hepatoblastoma, and embryonal sarcoma of the liver. Although they are classically considered pediatric neoplasms, they may (rarely) occur in adult patients. Hepatoblastoma represents the most frequent liver neoplasm in the pediatric population, followed by hepatocellular carcinoma and embryonal sarcoma of the liver; while pancreatoblastoma is the most common malignant pancreatic tumor in childhood. Both in children and adults, the mainstay of treatment is complete surgical resection, either up front or following neoadjuvant chemotherapy. Unresectable and/or metastatic neoplasms may be amenable to complete delayed surgery after neoadjuvant chemotherapy. However, these neoplasms display a more aggressive behavior and overall poorer prognosis in adults than in children, probably because they are diagnosed in later stages of diseases.

Key words: hepatic embryonal sarcoma, hepatoblastoma, pancreatoblastoma, pediatric tumors, gastro-intestinal tumors

Introduction

Pediatric neoplasms represent an important diagnostic challenge for pathologists, even in specialized institutions, both for their rarity and for their peculiarity when compared to adult neoplasms. The majority of them are referred to as embryonal because they arise as a result of alterations in the process of organogenesis or normal growth and are characterized by a proliferation of primitive cells, reproducing the corresponding tissue at various stages of embryonic development. Considering these peculiarities, these tumors are the borderland between embryology and pathology, as defined by Willis in 1950. This review will focus on embryonal gastrointestinal pediatric neoplasms in adults, which comprise pancreatoblastoma, hepatoblastoma and embryonal

Received and accepted: November 9, 2022

Correspondence

Gaetano Magro

Department of Medical and Surgical Sciences and Advanced Technologies "G. F. Ingrassia", Anatomic Pathology, University of Catania, 95123 Catania, Italy E-mail: g.magro@unict.it

Conflict of interest

The Authors declare no conflict of interest.

How to cite this article: Tsvetkova V, Magro G, Broggi G, et al. New insights in gastro-Intestinal "pediatric" neoplasms in adult patients: pancreatoblastoma, hepatoblastoma and embryonal sarcoma of the liver. A practical approach by GIPPI-GIPAD Groups. Pathologica 2022;114:64-78. https://doi. org/10.32074/1591-951X-559

© Copyright by Società Italiana di Anatomia Patologica e Citopatologia Diagnostica, Divisione Italiana della International Academy of Pathology



This is an open access journal distributed in accordance with the CC-BY-NC-ND (Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International) license: the work can be used by mentioning the author and the license, but only for non-commercial purposes and only in the original version. For further information: https://creativecommons. org/licenses/by-nc-nd/4.0/deed.en sarcoma of the liver. Although they are classically considered pediatric neoplasms, they may (rarely) occur in adult patients with an overall poorer prognosis than in children. The pathologist plays a crucial role in the initial diagnosis with important implications in patient management and therapeutic choice. Moreover, the introduction of pediatric therapeutic collaborative protocols has achieved the correct identification of histologic subtypes through systematic central histopathological review, and to collect frozen material for the integration of new biological parameters for future tailored treatments.

Pancreatoblastoma

Pancreatoblastoma (PB) is a malignant epithelial neoplasm characterized by solid architecture, acinar differentiation and the presence of squamoid nests. This neoplasm is typical of young patients (median age at diagnosis: 4-5 years) and represents almost 25% of all pancreatic lesions during childhood ¹⁻⁵. However, PB can also be diagnosed in adults (second age peak: 40 years old), although in this age range it is considered a very rare malignancy, representing < 1% of all pancreatic tumors ¹⁻³. No sex predominance is established. The etiology of this entity is unknown and the vast majority of cases are sporadic. However, emerging clinical data suggest an association between PB and two different genetic syndromes, namely Beckwith-Wiedemann syndrome 1,6,7 and familial adenomatous polyposis (FAP) 8.

CLINICAL FEATURES

The most common signs/symptoms of PB include abdominal pain, nausea, vomiting, weight loss and diarrhea; a palpable mass can be found in pediatric patients. Notably, a non-negligible proportion of cases is incidentally discovered.

Rarely, PB can secrete various hormones provoking the corresponding symptoms. In the pediatric population, elevated serum levels of alpha-fetoprotein (AFP) are a common finding, which can be used as a biomarker for follow-up after surgical resection. This occurrence, however, is uncommon in adults ^{9,10}. A small fraction of patients may present with a concomitant para-neoplastic Cushing syndrome, due to an inappropriate secretion of ACTH ^{1,11-13}.

65

MACROSCOPIC FEATURES

PBs are usually large masses, with diameters ranging from 1.5 to 20 cm (mean 10 cm) ¹⁴. They are usually well circumscribed or incapsulated, solid, neoplasms. PBs can occur in the head, the body or the tail of the pancreas without showing any specific anatomical predilection. The cut surface has tan to yellowish color and consists of various lobules separated by fibrous bands forming nests of different sizes. Some PBs can present hemorrhagic changes and/or cystic degeneration, which are more often associated with genetic syndromes ¹⁵.

MICROSCOPIC FEATURES AND CYTOLOGY

Histologically, the tumor is composed of highly cellular lobules separated by fibrous bands and admixed with unequally distributed squamous nests (Fig. 1) ^{1,2}. The number, cellularity and width of the fibrous bands on the tumor sample can vary, also depending on the patient's age. In adult patients for example, PBs are similar to those of childhood, but stromal bands are usually less abundant and less cellular ¹. PBs can exhibit solid, acinar and/or trabecular growth patterns. The neoplastic elements are monomorphic and roundish-to-polygonal, with small nuclei including an evident nucleolus; mild to modest nuclear atypia is generally observed. The histological hallmark of PBs, which is a necessary element to differentiate them from acinar cell carcinomas, is represented by squamous nests (Fig. 1) ^{1,2}. Even though squamoid nests, can be easily documented in histological sections, as pale and round areas, sometimes with a whorled aspect, their identification in cytological samples may be challenging. Interestingly, Reid and colleagues ¹⁶ distinguished two different subtypes of cells composing squamoid nests of PBs on cytological samples. The first subtype is represented by well defined, large and ovoid cells with clear cytoplasm, without significant atypia. The second subgroup is composed of ill-defined, loosely cohesive cells that may present nuclear clearing due to the accumulation of biotin ¹⁷. Although squamoid nests are a crucial key for PB diagnosis, their distribution is uneven throughout the neoplasm and between different PBs. This indicates that, in case of pancreatic tumors with acinar differentiation, an accurate and exhaustive tumor sampling should be performed in order to document the presence of squamoid nests, which can be focal. Lastly, calcifications are sometimes present.

IMMUNOHISTOCHEMICAL AND MOLECULAR FEATURES

The predominant acinar component of PB at immunohistochemistry (IHC) shows a diffuse and strong pos-



Figure 1. Figure summarizing the most important microscopic features of pancreatoblastoma. (A) Pancreatoblastomas are usually well-circumscribed neoplasms (hematoxylin-eosin, magnification: 2X); (B) the stromal component may be well represented (hematoxylin-eosin, magnification: 4X); (C) the classic histological appearance is of a hypercellular neoplasm with pale and roundish areas representing the so-called squamoid nests (hematoxylin-eosin, magnification: 10X); (D) detail at a higher magnification, with immunostaining for Bcl-10: this marker shows a very strong positivity in pancreatoblastomas, except for squamoid nests, which are negative (magnification: 20X).

itivity for Bcl-10 and trypsin, while squamoid nests remain negative (Fig. 1). Squamous nests, on the other hand, may express immunostaining for β -catenin (aberrant nuclear accumulation), CD200 and EMA^{1,2,18}. PBs can also show positive staining for chromogranin and synaptophysin, but, if present, positivity is usually focal or very focal. IHC may be required to exclude a neuroendocrine tumor, but in this kind of differential diagnosis the morphology is usually sufficient ^{1,5}. Diagnostic criteria for the most important differential diagnosis of PBs are summarized in Table I.

The molecular profile of PB has not been completely deciphered. One of the most common genetic alterations in PB is represented by the loss of the short arm of chromosome 11 (loss of 11p), also seen in patients with Beckwith-Wiedemann syndrome ⁶. This alteration leads to the dysregulation of IGF2¹⁶. A genetic hallmark of PB is represented by the presence of recurrent upregulation of the WNT pathway ^{1,8,18-21}; this can depend on activating *CTNNB1* mutations but also due to inactivating *APC* mutations, the latter present in FAP syndrome ²². Of note, no cases of PBs with microsatellite instability high-tumor mutational burden have been so far reported.

PROGNOSIS

In adults, PB may be successfully treated, in the case of limited extension, by radical resection; unfortunately, they can present with a significant degree of local invasion and distant metastasis in up to 1/3 of patients at the time of diagnosis. The most common site of distant metastases is represented by the liver. The overall survival rate of PB patients is 50%, which increases to up to 2/3 of patients in patients with surgically resected tumors 1,2,21. Differently from adults, children display a more favorable prognosis. One possible reason is that children are diagnosed in earlier stages, with small tumor masses and without local or distant metastasis ^{1,2,21}. Computed tomography and magnetic resonance are of help in radiological diagnosis; such analyses are recommended also for the follow-up after resection, integrating radiologic data with AFP dosage ^{23,24}.

Tumor type/ subtype	Most common architectural patterns	Squamoid nests	Necrosis	Stroma	Evident nucleoli	Acinar cell IHC markers expression	Neuroendocrine marker expression	Most important IHC markers for diagnosis
Pancreatoblastoma	Acinar, trabogular, solid	Yes	Possible	Fibrous, often	Yes	Yes	No or focal	Bcl10, trypsin;
Acinar cell carcinoma	Acipar	No	Frequent	Fibrous	Voc	Voc	No or focal	Bolio trypsin
	glandular, trabecular, solid	NO	Fiequein	occasional	ies	165	NO OF IOCAI	Berro, trypsin
Pancreatic NET	Nesting,	No	Very rare	Highly	No (salt-	No	Yes, diffuse and	Chromogranin,
	trabecular, glandular, solid			vascular, hyalinized	pepper chromatin)		strong	Synaptophysin, Ki67 for grading

Table I. Most important features helpful in the main differential diagnosis of pancreatoblastoma.

Abbreviations: IHC: immunohistochemical; SN: squamoid nests, NET: neuroendocrine tumors.

Hepatoblastoma

Hepatoblastoma (HB) is a malignant hepatic embryonal tumor arising from a hepatocyte precursor cell that recapitulates the various stages of liver development, consisting of a combination of either epithelial or epithelial and mesenchymal elements. HB is the most frequent liver tumor in children, accounting for 1% of all pediatric malignancies, with a worldwide estimated incidence of 1-1.5 cases per million children ^{25,26}. A rising incidence of HB has been reported, probably related to a higher number of premature birth and low-birth-weight survivors ^{26,27}. The majority of HBs are sporadic, but a subset can occur in the context of several congenital abnormalities and constitutional genetic syndromes, such as Beckwith-Wiedemann syndrome, Simpson-Golabi-Behmel syndrome, trisomy 18 (Edward syndrome), Sotos syndrome and familial adenomatous polyposis coli ²⁸⁻³¹.

CLINICAL FEATURES

HB mostly affects infants and young children between 6 months and 5 years (80-90%), with a median age of 18 months ^{32,33}, but cases have also been reported in neonates, adolescents and (rarely) adults up to the age of 80 years ³⁴. There is a slight male predominance ³². HB usually presents as an enlarging solitary mass in 80-85% of cases, involving the right lobe (55-60%), the left lobe (15-20%), or both lobes in the remaining cases. Multifocal masses at presentation do occur, and metastases at diagnosis are present in 5% of cases, usually to the lungs. The most common presentation symptoms are abdominal pain, anorexia, weight loss, nausea, and vomiting; jaundice is present in less than 5%. Liver enzymes are generally normal; thrombocytosis with platelet counts above 450,000/ µL is frequent. Alpha-fetoprotein (AFP) is highly elevated (in the thousands and even millions ng/mL), and is useful for monitoring recurrence of disease and chemotherapy response. Imaging studies are necessary for PRETEXT (PRE-Treatment EXTent of tumor) staging of the tumor, and are important for disease assessment and treatment selection. Enhanced computerized tomography (CT) and magnetic resonance imaging (MRI) are recommended for this purpose ^{35,36}. HB presents as well-delineated hypodense (rarely isodense) mass on CT. Foci of calcification, ossification, and hemorrhage may be present. By MRI, most tumors are T2 hyperintense, T1 hypointense, and hypointense with gadolinium in the epatobiliary phase.

MACROSCOPIC FEATURES

HB is usually a single, well-delineated and lobulated mass, within a normal liver, with a variegated cut surface, and an irregular thin pseudocapsule. Post-therapy tumors usually show cystic change, necrosis, hemorrhage, and a more accentuated pseudocapsule (Fig. 2). Gross examination is crucial either in upfront surgery (primary resection) or in post-chemotherapy surgery. A mapping liver tumor resection specimens with a complete sampling of at least 1 cross-section of tumor (similar to Wilms tumor or osteosarcoma) is recommended, in order to assess the percent of



Figure 2. Gross resection of post-therapy tumor. The cut surface shows cystic change, necrosis, and hemorrhage.

post-chemotherapy necrosis and the morphology of the residual tumor. Accurate sampling also permits identification of worrisome/significant areas, which require further sampling. Additional sections from normal parenchyma should be performed ³⁷.

MICROSCOPIC AND IMMUNOHISTOCHEMICAL FEATURES

According to the International Pediatric Liver Tumor Consensus Classification (PLTCC), HB is histologically classified on the basis of the components present as either epithelial HB or mixed HB when both epithelial and mesenchymal components are present ^{38,39}.

Epithelial HB

Well-Differentiated Fetal (WDF) HB (or pure fetal HB with low mitotic activity) (Fig. 3) is composed of thin plates, cords or nests of uniformly small-medium, polygonal cells with a well-defined outline and abundant eosinophilic to clear vacuolated cytoplasm, resulting in a characteristic dark-and-clear cell pattern. The nuclei are central and round, without nucleoli. Mitoses are rare (< 2/10 HPF); necrosis and pleomorphism are absent. Extramedullary hematopoiesis is typical. Immunohistochemically, the neoplastic cells show strong cytoplasmic positivity for glutamine synthetase (GS); weak, but diffuse staining of glypican 3 (GPC3) in a fine stippled pericanalicular cytoplasmic pattern; and membranous, even cytoplasmic, with rare, never strong diffuse, nuclear positivity for β -catenin. WDF HB cannot be diagnosed on biopsies or postchemotherapy specimens, but only on primary resection specimens. According to the Pediatric Hepatic International Tumor Trial (PHITT) protocol, the diagnosis

Figure 3. Well differentiated fetal hepatoblastoma. (A) Uniform small-medium and polygonal cells with well-defined outline and characteristic dark-and-clear cell pattern. (B) Cytoplasmic and membranous β -catenin staining with weak positive and

negative nuclei. (C) Cytoplasmic GPC3 staining in a fine stippled pattern. (D) Strong cytoplasmic GS staining.





Figure 4. Crowded fetal hepatoblastoma. (A) The neoplastic cells show slightly increased N/C, round nuclei with frequent nucleoli and dense eosinophilic cytoplasm; extramedullary hematopoiesis is frequent (inset). (B) Strong nuclear β -catenin staining. (C) Diffuse and coarse cytoplasmic GPC3 staining. (D) Diffuse and strong cytoplasmic GS staining.

of pure fetal HB identifies the group of very-low-risk patients, treated with upfront surgery only, without the need for further therapy if completely resected ⁴⁰.

Crowded fetal (CF) HB (also known mitotically active fetal HB) (Fig. 4) is a fetal HB with closely packed ("crowded") cells and mitotic activity $\ge 2/10$ HPF. The neoplastic cells show higher nuclear-cytoplasmic (N/C) ratio, round nuclei with frequent nucleoli and a dense eosinophilic cytoplasm due to minor cytoplasmic glycogen storage. Extramedullary hematopoiesis is frequent. Nuclear pleomorphism and atypical mitoses are absent (and should suggest a pleomorphic fetal HB when seen). Immunohistochemically, the neoplastic cells show diffuse and strong cytoplasmic positivity for GS, and a diffuse and coarse cytoplasmic staining of GPC3. Many nuclei are positive for b-catenin. These tumors will require chemotherapy. Embryonal HB (Fig. 5) recapitulates the embryonic stage of liver developmental and is composed of poorly cohesive cells with scant and poorly outlined cytoplasm, high N/C, and a large, angulated to oval nucleus with a prominent nucleolus. Mitoses are frequent and necrosis may be seen. The neoplastic cells are arranged in solid sheets or plates of variable thickness, incomplete/complete tubulo-glandular structures and rosette-like configurations. The vascular network is well developed encompassing a fine capillary network and large vascular channels. Extramedullary hematopoiesis is very rarely observed. Immunohistochemically, the neoplastic cells show uniform nuclear β -catenin positivity, variable GS staining from patchy single cell positivity to negativity, and variable GPC3 from absent to strong, coarse, diffuse cytoplasmic staining. Embryonal HB almost always occurs in



Figure 5. Embryonal hepatoblastoma. (A) The neoplastic cells are densely arranged in solid sheets/plates, incomplete/ complete tubulo-glandular structures, and rosette-like configurations, and show scant cytoplasm, high N/C, angulated/oval nucleus and numerous mitoses (inset). (B) Uniform nuclear β -catenin staining. (C) Strong, coarse, granular GPC3 staining. (D) Variable GS staining.

combination, often intermixed without demarcation, with a fetal component. A zonation is observed with embryonal cells in the centre, surrounded by CF cells, rimmed by varying proportion of WDF cells.

Small cell undifferentiated HB (SCU HB) was originally termed "anaplastic" and described as a lesion having small cells resembling those of neuroblastoma. Subsequently, the term "anaplasia" had been replaced by "small-cell undifferentiated (SCU)", based on the evidence of small undifferentiated round and spindle cells. However, the definition of SCU HB in recent years has dramatically changed. In fact, this entity in the past was uniformly assigned a worse prognosis both in cases with diffuse and in cases with minimal small cell morphology. Subsequently, the evidence of integrase interactor 1 (INI-1) loss of expression in tumors with a diffuse SCU morphology, has contributed in their reclassification as as hepatic rhabdoid tumors. HBs with a minimal SCU component show sheets and nests of small round to ovoid cells with scant cytoplasm, relatively fine nuclear chromatin with variable mitoses, intimately intermixed with embryonal HB areas. INI-1 is usually preserved. Whether a small component of SCU, usually less than 5% of tumor, with INI-1 preserved, in an otherwise typical epithelial HB is associated with an aggressive clinical behavior, remains object of debate.

Macrotrabecular (MT) HB (Fig. 6A) is a provisional category, representing a growth pattern rather than a histotype. Its morphologic overlap with pediatric hepatocellular carcinoma (HCC) and hepatocellular neoplasm NOS (HCN-NOS) (see below) may be a di-



Figure 6. Variants of hepatoblastoma. (A) Macrotrabecular pattern with fetal morphology and focal pleomorphism. (B-C) Post-chemotherapy lung metastasis of same patient showed in A. The neoplasm is more pleomorphic, with abnormal mitoses, conspicuous nucleoli, and strong nuclear β -catenin staining (inset C). (D) Mixed HB with osteoid showing strong nuclear β -catenin staining (inset).

agnostic challenge, especially on biopsy. MT pattern could be found pure (rarely) or in combination with other patterns, and is characterized by trabeculae greater than 5 cells and less than 10-20 cells in thickness. The cells within these macrotrabeculae show crowded fetal or embryonal morphology, reproducing the immunophenotype of these components, with strong nuclear β -catenin staining. The presence of trabeculae less than 10-20 cells in thickness, the coexistence of otherwise typical areas of HB, and the strong nuclear β -catenin staining help in distinguishing this pattern from pediatric HCC. At present, there is no evidence of a prognostic significance of macrotrabecular pattern.

Pleomorphic epithelial HB is an uncommon pattern of HB, more often seen in post-chemotherapy resec-

tion specimens and in metastases following chemotherapy. Nuclear features are more pleomorphic when compared with WDF or CF HB, with irregular shape, abnormal mitoses, and large, conspicuous nucleoli. When these pleomorphic cells assume a macrotrabecular growth pattern, the tumor may simulate HCC or may overlap with HCN-NOS (Fig. 6B, C). Strong nuclear β -catenin, strong GS, and variable GPC3 staining help in distinguishing pleomorphic MT HB from pediatric HCC. Immunostains are less useful in the distinction from HCN-NOS.

Cholangioblastic HB exhibits prominent cholangioblastic features and forms small ducts. The cells tend to be cuboidal rather than columnar, and the nuclei are usually round with coarse chromatin. This variant needs to be differentiated from acinar structures in areas of fetal HB and from ductular reaction at the periphery of the tumor, especially after chemotherapy. The choloangioblastic component shows nuclear β -catenin positivity unlike ductular reaction, and GS and GPC3 negativity unlike the acinar structures of fetal HB.

Mixed HB

Mixed HB is characterized by a complex mixture of epithelial and mesenchymal elements. The neoplastic mesenchymal elements are integral part of the tumor, showing nuclear β -catenin positivity, and do not represent the result of the chemotherapy or a "metaplastic" change (Fig. 6D). The mesenchymal elements most often consist of mature/immature fibrous tissue, osteoid, and cartilage. A small percentage of mixed HBs displays teratoid features (*i.e. teratoid HB or HB with heterologous elements*) characterized by a mixture of heterologous elements, such as endoderm, neural elements, and neuroectodermal derivates.

Hepatocellular neoplasm - NOS

Hepatocellular neoplasm - NOS (HCN-NOS) is a new provisional category, previously designated as "transitional cell liver tumors" (TCLT) by Prokurat ⁴¹, including hybrid lesions with overlapping HB and HCC features. HCN-NOS are highly aggressive and typically occur in older children (over the age of 8 years) with high or very high serum AFP levels, and an overall unfavourable outcome. The neoplasm may show macrotrabecular growth pattern and HCC-like features, especially in post-therapy specimens. At the molecular level, HCN-NOS carry β -catenin (*CTNNB1*) mutations as well as other mutations seen in HCC, such as TERT promoter mutations ^{42,43}.

MOLECULAR FEATURES

HBs are neoplasms with relatively stable genomes, with a limited number of structural and numerical abnormalities ^{44,45}. The vast majority (up to 90%) of HBs harbor activation of the canonical Wnt-signaling pathway, through somatic mutations of *CTNNB1* in over 80%, or more rarely, other Wnt-signaling genes; germline alterations, including *APC* mutations ⁴⁴⁻⁴⁹ may also be observed. *NFE2L2* (also known as *NRF2*) is the second most mutated gene, found in 5-10% of HB, and followed by mutations in *TERT* promoter, both associated with poor prognosis. Other pathways involved in HB pathogenesis include Notch, Sonic Hedgehog, PI3K/AKT, EGFR and Hippo pathway

(YAP) ^{42,46,50-53}. Integrated genomic studies reported 3 distinct risk-stratifying molecular HB subtypes associated with low, intermediate, and high risk ^{42,53}. High risk tumors are characterized by high *NFE2L2* activity; high LIN28B, HMGA2, SALL4, and AFP expression; low let-7 expression; and HNF1A activity; and high coordinated expression of oncofetal proteins and stem cell markers ^{49,51}. Moreover, HB epigenomic profiling revealed genome-wide dysregulation of RNA editing in HB demonstrating additional epigenomic clusters, including an aggressive subtype characterized by progenitor-like phenotype, methylation features, strong 14q32 locus expression, and *CTNNB1* and *NFE2L2* mutations ⁵⁴.

PROGNOSIS

Tumor stage

The PRETEXT system is used for staging and risk stratification for HB. The PRETEXT system comprises the PRETEXT group, and the annotation factors. The PRETEXT groups (PRETEXT I, II, III, or IV) reflect hepatic parenchymal tumor involvement; while the annotation factors describe the extension of tumor beyond the hepatic parenchyma and include hepatic venous/ inferior vena cava involvement (V), portal venous involvement (P), extrahepatic disease (E), multifocality (F), tumor rupture (R), and metastatic disease (M). CHIC (Children's Hepatic International Collaboration) has created a new risk-stratified staging system in children with HB, the Children's Hepatic International Collaboration - Hepatoblastoma Stratification (CHIC-HS) ⁴⁰. This system was established with risk factors including PRETEXT groups, metastatic disease, age at diagnosis (< 3 years, 3-7 years, and \geq 8 years), AFP concentration (\leq 100 µg/L and 101-1000 µg/L), PRETEXT annotation factors, and tumor resectbility at diagnosis. The primary and most important factor for risk stratification is the PRETEXT group, followed by metastatic disease. All patients with metastatic disease were defined as high risk. Age \geq 8 years in PRE-TEXT I/II/III group and age ≥ 3 years in PRETEXT IV group were high-risk factor. Younger patients with AFP level \leq 100 ng/mL are stratified as high-risk group.

Embryonal sarcoma of the liver

Embryonal sarcoma of the liver (ESL), also known as "*undifferentiated embryonal sarcoma*", is a malignant mesenchymal neoplasm of the liver. The terms "*embryonal and undifferentiated*" refer to the fact that the tumor is histologically composed of mesenchymal cells with no evidence of differentiation ^{1,55,56}. ESL,



Figure 7. Ultrasound examination of ESL showing a solid hyperechoic tumor, with hypoechoic/anechoic, cystic portions (A). ESL typically presents on coronal and axial MRI as hyperintense on T2 and hypointense on T1 mass with distinct borders (B, C).

first described by Stocker and Ishak in 1978 57, typically occurs in children with an age ranging from 5 to 20 years, with a peak of incidence between 6 and 10 years ^{1,54-60}. It accounts approximately for 6-13% of all hepatic pediatric malignancies, representing the third most frequent malignant tumor of the liver after hepatoblastoma and hepatocellular carcinoma in this age group ^{1,54-60}. Although ESL is classically considered a pediatric neoplasm, some cases have also been reported in adult patients with a predilection for women ^{58,61,62}. The etiology of ESL is largely unknown; based on the evidence that some cases of ESL may contain areas with mesenchymal hamartoma (MH)like histology and/or share with this benign tumor the same chromosome translocation t(11;19) (g13;g13.4), a molecular continuum between these two entities has been suggested^{63,64}.

CLINICAL FEATURES

Most patients present with a palpable mass associated with abdominal distension, pain, fever, anorexia, vomiting, irregular alvus, respiratory distress and weight loss ^{1,54-60,65}. Hemoperitoneum due to the rupture of the liver is a rare complication ^{1,54-60}. Although there are no specific laboratory findings, leukocytosis and increased serum alkaline phosphatase are frequently found 55,56; liver neoplastic serum markers are usually normal. On ultrasonography, ESL usually presents as large solid-cystic mass (Fig. 7A) 55,56,66,67. If the cystic component is prominent, the tumor may be misinterpreted as a benign neoplasm, causing diagnostic and therapeutic delays for the patient 55,56,66,67. Computed tomography usually reveals a single, predominantly hypodense, cystic mass, with internal septations 66,67. Magnetic resonance imaging, showing a T2-hyperintense and T1-hypointense mass (Fig. 7B, C), is also useful for planning the surgical approach, due to its high accuracy in the detection of vascular invasion, biliary obstruction and hilar lymphadenopathy ^{55,56,66,67}. The diagnosis is histologically-based on liver needle/wedge biopsy.

MACROSCOPIC FEATURES

ESL occurs more frequently in the right hepatic lobe ^{55,56}. On gross examination, the tumor presents as a single, large-sized, well-demarcated and unencapsulated lesion, measuring 10-30 cm in its greatest diameter (Fig. 8A) ^{55,56}. The well-defined margins of the mass are due to the presence of a fibrous pseudocapsule resulting from the adjacent compressed liver parenchyma ^{55,56}. The cut surface shows a solid mass, gray-whitish in color, with alternating myxoid and cystic areas (Fig. 8B) ⁵⁵. Necrotic and hemorrhagic areas are commonly seen ⁵⁶.

HISTOPATHOLOGIC FEATURES

Histologically, ESL is predominantly composed of variably-sized spindle, oval to stellate cells, compactly or loosely set in a variably fibro-myxoid stroma (Fig. 9A-D) ^{1,54,55,66}. The most striking feature of ELS is the presence of pleomorphic, often multinucleated, cells with hyperchromatic nuclei (Fig. 9D) ^{1,54,55,68}. In the more collagenized areas, neoplastic cells adopt a spindled morphology and are arranged in a fascicular or storiform growth pattern ^{1,54,55,68}. Notably, neoplastic cells exhibit eosinophilic granular cytoplasm, inconspicuous nucleoli and indistinct cell borders. The presence of multiple, PAS-positive, diastase-resistant, eosinophilic, cytoplasmic and extracellular hyaline globules is a characteristic finding of ESL (Fig. 9C) ^{1,54,55,68}. Mitotic activity is usually high and atypical mitoses and



Figure 8. Gross examination of partial hepatectomy after neoadjuvant chemotherapy showing a large-sized and oval-shaped mass with well-demarcated margins (A). On cut surface, the tumor is yellow to whitish in color and often exhibits alternating solid and cystic areas. Necrosis and extensive fibrosis, due to the effects of the neoadjuvant chemotherapy are seen (B).



Figure 9. Wedge biopsy showing a moderately cellular tumor completely replacing liver parenchyma (H&E; original magnification 25x) (A). Higher magnification showing mitotically-active, rounded- and stellate-shaped neoplastic cells with ill-defined borders, set in a fibrous stroma; apoptotic bodies are also seen. (H&E; original magnification 200x) (B). ESL may also exhibit myxoid areas; notice the typical eosinophilic intra- and extra-cellular hyaline globules (H&E; original magnification 200x) (C). The detection of highly pleomorphic neoplastic cells is an additional characteristic feature (H&E; original magnification 400x) (D).

apoptotic bodies are often seen (Fig. 9B). Intratumoral necrosis and hemorrhage are common. Clusters of entrapped hepatocytes and biliary ducts may be found at the periphery of the tumor, as well as foci of extramedullary hematopoiesis ^{1,55,56}. Uncommon morphological features include tumor areas with MHlike ^{1,55,56,68} or rhabdoid morphology ⁶⁹.

IMMUNOHISTOCHEMICAL AND MOLECULAR FEATURES

ESL exhibits a non-specific immunophenotyped ^{1,55,56}. Neoplastic cells are diffusely stained with vimentin, α -1-antitrypsin and α -1-antichymotrypsin, and focally with glypican-3, α -smooth muscle actin, muscle-specific actin, desmin, CD34, S-100, calponin, cytokeratins, CD68, BCL-2, CD10 and p53 68. A dot-like immunopositivity for cytokeratins and membrane staining with CD56 has been also described⁷⁰. Neoplastic cells are consistently negative for EMA, myogenin, Myo-D1, α -FP and Hep Par-1 ^{1,55,56}. Ki-67 proliferation index is usually high, ranging from 30% to 95%. Hyaline bodies are usually stained with vimentin, α -1-antitrypsin and α-1-antichymotrypsin. The variable co-expression of histiocytic, muscle and epithelial markers suggests a tumor origin from primitive stem cells 55,56. As ESL may exhibit focal or dot-like positivity for cytokeratins AE1/ AE3 and CAM5.2, a misdiagnosis of carcinoma may be rendered. As the half of cases of ESL share the expression of glypican-3 with hepatocellular carcinoma and hepatoblastoma, the use of this immunomarker should be avoided in the differential diagnosis 55,56. The differential diagnosis of ESL in the pediatric age mainly includes embryonal rhabdomyosarcoma, hepatoblastoma and MH; in adult patients, ESL should be distinguished from sarcomatoid hepatocellular carcinoma, malignant melanoma and metastatic gastro-intestinal stromal tumor 55,56. The most relevant criteria for the differential diagnosis are summarized in Table II. The genetic landscape of ELS is still largely unknown. Comparative genomic hybridization data suggested a potential role for chromosomal instability in the pathogenesis of this tumor, showing that copy number alterations are frequently found ^{59,71,72}. In addition, gains in chromosomes 1q, 5p, 6q and losses in chromosome 14, 9p and 11p. are recurrent molecular events of this tumor 59,71,72. As mentioned above, some cases of ESL share with MH the same chromosome translocation t(11;19) (g13;g13.4), suggesting a molecular continuum between these two lesions 63,64. Mutations in the DNA-binding domain of the TP53 gene have also been reported 71,73.

PROGNOSIS

Although ESL is a malignant tumor with an aggressive biological behavior characterized by metastatic spread to the lungs and peritoneum. The prognosis of patients treated with surgical resection (partial hepatectomy) and adjuvant chemotherapy is generally favorable (5-year overall survival > 70%) ^{1,55,56,74-77}. Tu-

Table II. Mai	n differentia	l diagnoses	of embryona	al sarcoma	of the liver.
			•••••••••••••••••••••••••••••••••••••••		•••••••

Tumor type/subtype	Age of presentation	Histopathologic features	Immunohistochemical features
Embrional sarcoma of the liver	6-10 years	Sheets of pleomorphic spindle, oval to stellate cells set in a fibro-myxoid stroma High mitotic activity Atypical mitoses Hyaline globules	Not specific Vimentin, CD68, CD56, BCL2, CD10, a-1-antitrypsin, cytokeratins and Glypican-3
Mesenchymal hamartoma	< 2 years	Myxomatous stroma with branching bile ducts and entrapped hepatocytes arranged into a lobular architecture	Not useful
Hepatoblastoma	Mean age: 19 months	Different subtypes resembling the different stages of liver development	β-catenin (nuclear and membranous staining) Glypican-3 and glutamine synthetase
Embryonal rhabdomyosarcoma of the biliary tract	< 5 years	Small round blue cell tumor	Desmin, Myogenin and Myo-D1
Sarcomatoid hepatocellular carcinoma	Adults	Thickened trabeculae with pleomorphic spindle cells Mallory hyaline bodies	Hep Par-1, Glypican-3, a-FP, arginase, CD10 (canalicular) and pCEA (canalicular)
Metastatic gastro- intestinal stromal tumor	Adults	Spindle cells with eosinophilic cytoplasm	CD117 (c-kit), DOG-1 and CD34
Malignant melanoma	Adults	Highly pleomorphic spindle and/or epithelioid neoplastic cells Melanin pigment High mitotic activity	S100, Melan-A, HMB-45 and SOX-10

mor size > 15 cm and extrahepatic dissemination at the diagnosis are negative prognostic indicators ⁷⁴⁻⁷⁷. Neoadjuvant chemotherapy followed by surgery is reserved to the patients with unresectable tumors, while liver transplantation is a possibility for patients who are resistant to neoadjuvant chemotherapy ⁷⁴⁻⁷⁷.

References

- ¹ WHO Classification of Tumors Editorial Board. WHO Classification of Gastro-Intestinal Tumors, 5th Edition. 2019
- ² Luchini C, Grillo F, Fassan M, et al. Malignant epithelial/exocrine tumors of the pancreas. Pathologica 2020;112:210-226. https:// doi.org/10.32074/1591-951X-167
- ³ Mylonas KS, Nasioudis D, Tsilimigras DI, et al. A population-based analysis of a rare oncologic entity: malignant pancreatic tumors in children. J Pediatr Surg 2018;53:647-652. https://doi.org/10.1016/j. jpedsurg.2017.06.024
- ⁴ Shorter NA, Glick RD, Klimstra D, et al. Malignant pancreatic tumors in childhood and adolescence: the Memorial Sloan-Kettering experience, 1967 to present. J Pediatr Surg 2002;37:887-892. https://doi.org/10.1053/jpsu.2002.32897
- ⁵ Klimstra DS, Wenig BM, Adair CF, et al. Pancreatoblastoma: a clinicopathologic study and review of the literature. Am J Surg Pathol 1995;19:1371-1389. https://doi. org/10.1097/00000478-199512000-00005
- ⁶ Muguerza R, Rodriguez A, Formigo E, et al. Pancreatoblastoma associated with incomplete Beckwith-Wiedemann syndrome: case report and review of the literature. J Pediatr Surg 2005;40:1341-1344. https://doi.org/10.1016/j.jpedsurg.2005.05.025
- ⁷ Weksberg R, Shuman C, Beckwith JB. Beckwith-Wiedemann syndrome. Eur J Hum Genet 2010;18:8-14. https://doi.org/10.1038/ ejhg.2009.106
- ⁸ Abraham SC, Wu TT, Klimstra DS, et al. Distinctive molecular genetic alterations in sporadic and familial adenomatous polyposis-associated pancreatoblastomas: frequent alterations in the APC/beta-catenin pathway and chromosome 11p. Am J Pathol 2001;159:1619-1627. https://doi.org/10.1016/s0002-9440(10)63008-8
- ⁹ Defachelles AS, Martin De Lassale E, Boutard P, et al. Pancreatoblastoma in childhood: clinical course and therapeutic management of seven patients. Med Pediatr Oncol 2001;37:47-52. https:// doi.org/10.1002/mpo.1162
- ¹⁰ Rajpal S, Warren RS, Alexander M, et al. Pancreatoblastoma in adult: case report and review of the literature. J Gastrointest Surg 2006;10829-836. https://doi.org/10.1016/j.gassur.2005.11.011
- ¹¹ Kletter GB, Sweetser DA, Wallace SF, et al. Adrenocorticotropin-secreeting pancreatoblastoma. J Pediatr Endocrinol Metab 2007;20:639-642. https://doi.org/10.1515/jpem.2007.20.5.639
- ¹² Passmore SJ, Berry PJ, Oakhill A. Recurrent pancreatoblastoma with inappropriate adenocorticotrophic hormone secretion. Arch Dis Child 1988;63:1494-1496. https://doi.org/10.1136/ adc.63.12.1494
- ¹³ Quereshi SS, Bhagat M, KurkurePA, et al. Ectopic Cushing syndrome secondary to recurrent pancreatoblastoma in a child: lessons learnt. J Cancer Res Ther 2015;11:1027. https://doi. org/10.4103/0973-1482.151854
- ¹⁴ Ohike N, Morohoshi T. Exocrine pancreatic neoplasms of nonductal origin: acinar cell carcinoma, pancreatoblastoma, and solidpseudopapillary neoplasm. Surg Pathol Clin 2011;4:579-588. https://doi.org/10.1016/j.path.2011.03.001

- ¹⁵ Drut R, Jones MC. Congenital pancreatoblastoma in Beckwith-Wiedemann syndrome: an emerging association. Pediatr Pathol 1988;8:331-339. https://doi.org/10.3109/15513818809042976
- ¹⁶ Reid MD, Bhattarai S, Graham RP, et al. Pancreatoblastoma: cytologic and histologic analysis of 12 adult cases reveals helpful criteria in their diagnosis and distinction from common mimics. Cancer Cytopathol 2019;127:708-719. https://doi.org/10.1002/cncy.22187
- ¹⁷ Tanaka Y, Ijiri R, Yamanaka S, et al. Pancreatoblastoma: optically clear nuclei in squamoid corpuscles are rich in biotin. Mod Pathol 1998;11:945-9.
- ¹⁸ Lawlor RT, Daprà V, Girolami I, et al. CD200 expression is a feature of solid pseudopapillary neoplasms of the pancreas. Virchows Arch 2019;474:105-109. https://doi.org/10.1007/s00428-018-2437-7
- ¹⁹ Isobe T, Seki M, Yoshida K, et al. Integrated molecular characterization of the lethal pediatric cancer pancreatoblastoma. Cancer Res 2018;78:865-876. https://doi.org/10.1158/0008-5472. CAN-17-2581
- ²⁰ Jiao Y, Yonescu R, Offerhaus GJ, et al. Whole-exome sequencing of pancreatic neoplasms with acinar differentiation. J Pathol 2014;232:428-435. https://doi.org/10.1002/path.4310
- ²¹ Dhebri AR, Connor S, Campbell F, et al. Diagnosis, treatment and outcome of pancreatoblastoma. Pancreatology 2004;4:441-51, discussion 452-453. https://doi.org/10.1159/000079823
- ²² Raoul JL, Oziel-Taieb S, Lecomte T, et al. Case report: two cases of metastatic pancreatoblastoma in adults: efficacy of Folfirinox and implication of the Wnt/ -Catenin pathway in genomic analysis. Front Oncol 2021;11:564506. https://doi.org/10.3389/ fonc.2021.564506
- ²³ Yang Z, Gong Y, Ji M, et al. Differential diagnosis of pancreatoblastoma (PB) and solid pseudopapillary neoplasms (SPNs) in children by CT and MR imaging. Eur Radiol 2021;31:2209-2217. https://doi.org/10.1007/s00330-020-07309-3
- ²⁴ Huang Y, Yang W, Hu J, et al. Diagnosis and treatment of pancreatoblastoma in children: a retrospective study in a single pediatric center. Pediatr Surg Int 2019;35:1231-1238. https://doi. org/10.1007/s00383-019-04524-y
- ²⁵ Spector LG, Birch J. The epidemiology of hepatoblastoma. Pediatr Blood Cancer 2012;59:776-779. https://doi.org/10.1002/pbc.24215
- ²⁶ Allan BJ, Parikh PP, Diaz S, et al. Predictors of survival and incidence of hepatoblastoma in the pediatric population. HPB (Oxford) 2013;15:741-746. https://doi.org/10.1111/hpb.12112
- ²⁷ Feusner J. Prematurity and hepatoblastoma: more than just an association? J Pediatr 1998;133:585-586. https://doi.org/10.1016/ s0022-3476(98)70084-8
- ²⁸ Venkatramani R, Spector LG, Georgieff M, et al. Congenital abnormalities and hepatoblastoma: a report from the Children's Oncology Group (COG) and the Utah Population Database (UPDB). Am J Med Genet A. 2014;164A:2250-2255. https://doi.org/10.1002/ ajmg.a.36638
- ²⁹ Clericuzio CL, Chen E, McNeil DE, et al. Serum alpha-fetoprotein screening for hepatoblastoma in children with Beckwith-Wiedemann syndrome or isolated hemihyperplasia. J Pediatr 2003;143:270-272. https://doi.org/10.1067/ S0022-3476(03)00306-8
- ³⁰ Harvey J, Clark S, Hyer W, Hadzie N, et al. Germline APC mutations are not commonly seen in children with sporadic hepatoblastoma. J Pediatr Gastroenterol Nutr 2008; 4:675-677. https:// doi.org/10.1097/MPG.0b013e318174e808
- ³¹ Schraw JM, Desrosiers TA, Nembhard WN, et al. Cancer diagnostic profile in children with structural birth defects: An assessment in 15,000 childhood cancer cases. Cancer 2020;126:3483-3492. https://doi.org/10.1002/cncr.32982

- ³² Schnater JM, Köhler SE, Lamers WH, et al. Where do we stand with hepatoblastoma? A review. Cancer. 2003; 98:668-678. https:// doi.org/10.1002/cncr.11585
- ³³ Litten JB, Tomlinson GE. Liver tumors in children. Oncologist 2008;13:812-820. https://doi.org/10.1634/theoncologist.2008-0011
- ³⁴ Celotti A, D'Amico G, Ceresoli M, et al. Hepatoblastoma of the adult: A systematic review of the literature. Surgical Oncology 2016;25:339-347. https://doi.org/10.1016/j.suronc.2016.07.003
- ³⁵ Towbin AJ, Meyers RL, Woodley H, et al. 2017 PRETEXT: radiologic staging system for primary hepatic malignancies of childhood revised for the Pediatric Hepatic International Tumor Trial (PHITT). Pediatr Radiol 2018;48:536-544. https://doi.org/10.1007/ s00247-018-4078-z
- ³⁶ Schooler GR, Squires JH, Alazraki A, et al. Pediatric Hepatoblastoma, Hepatocellular Carcinoma, and Other Hepatic Neoplasms: Consensus Imaging Recommendations from American College of Radiology Pediatric Liver Reporting and Data System (LI-RADS) Working Group. Radiology 2020;296:493-497. https://doi. org/10.1148/radiol.2020200751
- ³⁷ Finegold MJ, Lopez-Terrada D, Bowen J, et al. Protocol for the examination of specimens from pediatric patients with hepatoblastoma. Arch Pathol Lab Med 2007;131:520-529. https://doi. org/10.5858/2007-131-520-PFTEOS
- ³⁸ Lopez-Terrada D, Alaggio R, de Davila MT, et al. Towards an international pediatric liver tumor consensus classification: proceedings of the Los Angeles COG liver tumors symposium. Mod Pathol. 2014;27:472-491. https://doi.org/10.1038/modpathol.2013.80
- ³⁹ Ranganathan S, Lopez-Terrada D, Alaggio R. Hepatoblastoma and pediatric hepatocellular carcinoma: an update. Pediatr Dev Pathol 2020;23:79-95. https://doi.org/10.1177/1093526619875228
- ⁴⁰ Meyer RL, Maibach R, Hiyama E, et al. Risk-stratified staging in pediatric hepatoblastoma: a unified analysis from the Children's Hepatic tumors International Collaboration. Lancet Oncol 2017;18:122-131. https://doi.org/10.1016/S1470-2045(16)30598-8
- ⁴¹ Prokurat A, Kluge P, Kościesza A, et al. Transitional liver cell tumors (TLCT) in older children and adolescents: a novel group of aggressive hepatic tumors expressing beta-catenin. Med Pediatr Oncol. 2002;39:510-518. https://doi.org/10.1002/mpo.10177
- ⁴² Sumazin P, Chen Y, Treviño LR, et al. Genomic analysis of hepatoblastoma identifies distinct molecular and prognostic subgroups. Hepatology 2017;65:104-121. https://doi.org/10.1002/hep.28888
- ⁴³ Eichenmüller M, Trippel F, Kreuder M, et al. The genomic landscape of hepatoblastoma and their progenies with HCC-like features. J Hepatol 2014;61:1312-1320. https://doi.org/10.1016/j. jhep.2014.08.009
- ⁴⁴ Gröbner SN, Worst BC, Weischenfeldt J, et al. Author Correction: The landscape of genomic alterations across childhood cancers. Nature 2018;559(7714):E10. https://doi.org/10.1038/ s41586-018-0167-2
- ⁴⁵ Tomlinson GE, Douglass EC, Pollock BH, et al. Cytogenetic evaluation of a large series of hepatoblastomas: numerical abnormalities with recurring aberrations involving 1q12-q21. Genes Chromosomes Cancer 2005;44:177-184. https://doi.org/10.1002/ gcc.20227
- ⁴⁶ Luo JH, Ren B, Keryanov S, et al. Transcriptomic and genomic analysis of human hepatocellular carcinomas and hepatoblastomas. Hepatology 2006;44:1012-24. https://doi.org/10.1002/ hep.21328
- ⁴⁷ Armengol C, Cairo S, Fabre M, et al. Wnt signaling and hepatocarcinogenesis: the hepatoblastoma model. Int J Biochem Cell Biol 2011;43:265-270. https://doi.org/10.1016/j.biocel.2009.07.012
- ⁴⁸ Jia D, Dong R, Jing Y, et al. Exome sequencing of hepatoblastoma reveals novel mutations and cancer genes in the Wnt pathway and

ubiquitin ligase complex. Hepatology 2014;60:168-1696. https://doi.org/10.1002/hep.27243.

- ⁴⁹ Bell D, Ranganathan S, Tao J, et al. Novel Advances in Understanding of Molecular Pathogenesis of Hepatoblastoma: A Wnt/β-Catenin Perspective. Gene Expr. 2017; Feb;17(2):141-154. https://doi.org/10.3727/105221616X693639
- ⁵⁰ López-Terrada D, Gunaratne PH, Adesina AM, et al. Histologic subtypes of hepatoblastoma are characterized by differential canonical Wnt and Notch pathway activation in DLK+ precursors. Hum Pathol 2009;40:783-794. https://doi.org/10.1016/j. humpath.2008.07.022
- ⁵¹ Ranganathan S, Ningappa M, Ashokkumar C, et al. Loss of EG-FR-ASAP1 signaling in metastatic and unresectable hepatoblastoma. Sci Rep 2016;6:38347. https://doi.org/10.1038/srep38347
- ⁵² Tao J, Calvisi DF, Ranganathan S, et al. Activation of β-catenin and Yap1 in human hepatoblastoma and induction of hepatocarcinogenesis in mice. Gastroenterology 2014;147:690-701. https:// doi.org/10.1053/j.gastro.2014.05.004
- ⁵³ Cairo S, Armengol C, De Reynie's A, et al. Hepatic stem-like phenotype and interplay of Wnt/beta-catenin and Myc signalling in aggressive childhood liver cancer. Cancer Cell 2008;14:471-484. https://doi.org/10.1016/j.ccr.2008.11.002
- ⁵⁴ Carrillo-Reixach J, Torrens L, Simon-Coma M, et al. Epigenetic footprint enables molecular risk stratification of hepatoblastoma with clinical implications. J Hepatol 2020;73:328-341. https://doi. org/10.1016/j.jhep.2020.03.025.
- ⁵⁵ Putra J, Ornvold K. Undifferentiated embryonal sarcoma of the liver: a concise review. Arch Pathol Lab Med 2015;139:269-273. https://doi.org/10.5858/arpa.2013-0463-RS.
- ⁵⁶ Martins-Filho SN, Putra J. Hepatic mesenchymal hamartoma and undifferentiated embryonal sarcoma of the liver: a pathologic review. Hepat Oncol 2020;7(2):HEP19. https://doi.org/10.2217/ hep-2020-0002
- ⁵⁷ Stocker JT, Ishak KG. Undifferentiated (embryonal) sarcoma of the liver: report of 31 cases. Cancer 1978;42:336-48. https://doi.org/10.1002/1097-0142(197807)42:1<336::aidcncr2820420151>3.0.co;2-v
- ⁵⁸ Shu B, Gong L, Huang X, et al. Undifferentiated embryonal sarcoma of the liver in adults: Retrospective analysis of a case series and systematic review. Oncol Lett 2020;20:102. https://doi. org/10.3892/ol.2020.11963
- ⁵⁹ Setty BA, Jinesh GG, Arnold M, et al. The genomic landscape of undifferentiated embryonal sarcoma of the liver is typified by C19MC structural rearrangement and overexpression combined with TP53 mutation or loss. PLoS Genet 2020;16(4):e1008642. https://doi.org/10.1371/journal.pgen.1008642
- ⁶⁰ Techavichit P, Masand PM, Himes RW, et al. Undifferentiated Embryonal Sarcoma of the Liver (UESL): a single-center experience and review of the literature. J Pediatr Hematol Oncol 2016;38:261-268. https://doi.org/10.1097/MPH.000000000000529
- ⁶¹ Lenze F, Birkfellner T, Lenz P, et al. Undifferentiated embryonal sarcoma of the liver in adults. Cancer 200;112:2274-82. https://doi. org/10.1002/cncr.23431
- ⁶² Li XW, Gong SJ, Song WH, et al. Undifferentiated liver embryonal sarcoma in adults: a report of four cases and literature review. World J Gastroenterol 2010;16:4725-32. https://doi.org/10.3748/ wjg.v16.i37.4725
- ⁶³ Shehata BM, Gupta NA, Katzenstein HM, et al. Undifferentiated embryonal sarcoma of the liver is associated with mesenchymal hamartoma and multiple chromosomal abnormalities: a review of eleven cases. Pediatr Dev Pathol 2011 Mar-;14:111-6. https://doi. org/10.2350/09-07-0681-OA.1
- ⁶⁴ Mathews J, Duncavage EJ, Pfeifer JD. Characterization of translocations in mesenchymal hamartoma and undifferentiated embryo-

nal sarcoma of the liver. Exp Mol Pathol 2013;95:319-324. https://doi.org/10.1016/j.yexmp.2013.09.006

- ⁶⁵ Cao Q, Ye Z, Chen S, et al. Undifferentiated embryonal sarcoma of liver: a multi-institutional experience with 9 cases. Int J Clin Exp Pathol 2014;7:8647-8656.
- ⁶⁶ Gabor F, Franchi-Abella S, Merli L, et al. Imaging features of undifferentiated embryonal sarcoma of the liver: a series of 15 children. Pediatr Radiol 2016;46:1694-1704. https://doi.org/10.1007/ s00247-016-3670-3
- ⁶⁷ Gomes F, Melo D, Esteves C, et al. Undifferentiated embryonal sarcoma of the liver: A rare hepatic tumor and its related characteristic radiological features. Radiol Case Rep. 2021;16:646-650. https://doi.org/10.1016/j.radcr.2020.12.017
- ⁶⁸ Zheng JM, Tao X, Xu AM, et al. Primary and recurrent embryonal sarcoma of the liver: clinicopathological and immunohistochemical analysis. Histopathology 2007;51:195-203. https://doi. org/10.1111/j.1365-2559.2007.02746.x
- ⁶⁹ Papke DJ Jr, Fisch AS, Ranganathan S, et al. Undifferentiated embryonal sarcoma of the liver with rhabdoid morphology mimicking carcinoma: expanding the morphologic spectrum or a distinct variant? Pediatr Dev Pathol 2021:10935266211018930. https:// doi.org/10.1177/10935266211018930
- ⁷⁰ Pérez-Gómez RM, Soria-Céspedes D, de León-Bojorge B, et al. Diffuse membranous immunoreactivity of CD56 and paranuclear dot-like staining pattern of cytokeratins AE1/3, CAM5.2, and OSCAR in undifferentiated (embryonal) sarcoma of the liver. Appl Immunohistochem Mol Morphol 2010;18:195-198. https://doi. org/10.1097/PAI.0b013e3181bb2493
- ⁷¹ Hu X, Chen H, Jin M, et al. Molecular cytogenetic characterization of undifferentiated embryonal sarcoma of the liver: a case

report and literature review. Mol Cytogenet 2012;5:26. https://doi. org/10.1186/1755-8166-5-26

- ⁷² Sowery RD, Jensen C, Morrison KB, et al. Comparative genomic hybridization detects multiple chromosomal amplifications and deletions in undifferentiated embryonal sarcoma of the liver. Cancer Genet Cytogenet. 2001 Apr 15;126:128-133. https://doi. org/10.1016/s0165-4608(00)00404-0
- ⁷³ Lepreux S, Rebouissou S, Le Bail B, et al. Mutation of TP53 gene is involved in carcinogenesis of hepatic undifferentiated (embryonal) sarcoma of the adult, in contrast with Wnt or telomerase pathways: an immunohistochemical study of three cases with genomic relation in two cases. J Hepatol 2005;42:424-429. https:// doi.org/10.1016/j.jhep.2004.10.021
- ⁷⁴ Habibzadeh P, Ansari Asl M, Foroutan HR, et al. Clinicopathological study of hepatic mesenchymal hamartoma and undifferentiated embryonal sarcoma of the liver: a single center study from Iran. Diagn Pathol 2021;16:55. https://doi.org/10.1186/ s13000-021-01117-z
- ⁷⁵ Ismail H, Dembowska-Bagińska B, Broniszczak D, et al. Treatment of undifferentiated embryonal sarcoma of the liver in children--single center experience. J Pediatr Surg 2013;48:2202-2206. https:// doi.org/10.1016/j.jpedsurg.2013.05.020
- ⁷⁶ Pinamonti M, Vittone F, Ghiglione F, et al. Unexpected Liver Embryonal Sarcoma in the Adult: Diagnosis and Treatment. Case Rep Surg 2018;2018:8362012. https://doi.org/10.1155/2018/8362012
- ⁷⁷ Kim DY, Kim KH, Jung SE, et al. Undifferentiated (embryonal) sarcoma of the liver: combination treatment by surgery and chemotherapy. J Pediatr Surg 2002;37:1419-1423. https://doi. org/10.1053/jpsu.2002.35404