#### Check for updates

#### **OPEN ACCESS**

EDITED BY Guangzhi Ning, Tianjin Medical University General Hospital, China

REVIEWED BY Changfeng Fu, The First Hospital of Jilin University, China Qun Xue, Affiliated Hospital of Nantong University, China

\*CORRESPONDENCE Nunzio Vicario Image: Nunzio.vicario@unict.it Rosalba Parenti Image: Nunzio.vicario@unict.it

<sup>†</sup>These authors share first authorship

SPECIALTY SECTION This article was submitted to Multiple Sclerosis and Neuroimmunology, a section of the journal Frontiers in Immunology

RECEIVED 14 November 2022 ACCEPTED 06 December 2022 PUBLISHED 19 December 2022

#### CITATION

Denaro S, D'Aprile S, Alberghina C, Pavone AM, Torrisi F, Giallongo S, Longhitano L, Mannino G, Lo Furno D, Zappalà A, Giuffrida R, Tibullo D, Li Volti G, Vicario N and Parenti R (2022) Neurotrophic and immunomodulatory effects of olfactory ensheathing cells as a strategy for neuroprotection and regeneration. *Front. Immunol.* 13:1098212. doi: 10.3389/fimmu.2022.1098212

#### COPYRIGHT

© 2022 Denaro, D'Aprile, Alberghina, Pavone, Torrisi, Giallongo, Longhitano, Mannino, Lo Furno, Zappalà, Giuffrida, Tibullo, Li Volti, Vicario and Parenti. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

## Neurotrophic and immunomodulatory effects of olfactory ensheathing cells as a strategy for neuroprotection and regeneration

Simona Denaro<sup>1†</sup>, Simona D'Aprile<sup>1†</sup>, Cristiana Alberghina<sup>1</sup>, Anna Maria Pavone<sup>1</sup>, Filippo Torrisi<sup>1</sup>, Sebastiano Giallongo<sup>2</sup>, Lucia Longhitano<sup>2</sup>, Giuliana Mannino<sup>3</sup>, Debora Lo Furno<sup>1</sup>, Agata Zappalà<sup>1</sup>, Rosario Giuffrida<sup>1</sup>, Daniele Tibullo<sup>2</sup>, Giovanni Li Volti<sup>2</sup>, Nunzio Vicario<sup>1\*</sup> and Rosalba Parenti<sup>1\*</sup>

<sup>1</sup>Section of Physiology, Department of Biomedical and Biotechnological Sciences, University of Catania, Catania, Italy, <sup>2</sup>Section of Biochemistry, Department of Biomedical and Biotechnological Sciences, University of Catania, Catania, Italy, <sup>3</sup>Department of Chemical, Biological, Pharmaceutical and Environmental Sciences, University of Messina, Messina, Italy

Accumulating evidence sustains glial cells as critical players during central nervous system (CNS) development, homeostasis and disease. Olfactory ensheathing cells (OECs), a type of specialized glia cells sharing properties with both Schwann cells and astrocytes, are of critical importance in physiological condition during olfactory system development, supporting its regenerative potential throughout the adult life. These characteristics prompted research in the field of cell-based therapy to test OEC grafts in damaged CNS. Neuroprotective mechanisms exerted by OEC grafts are not limited to axonal regeneration and cell differentiation. Indeed, OEC immunomodulatory properties and their phagocytic potential encourage OEC-based approaches for tissue regeneration in case of CNS injury. Herein we reviewed recent advances on the immune role of OECs, their ability to modulate CNS microenvironment *via* bystander effects and the potential of OECs as a cell-based strategy for tissue regeneration.

#### KEYWORDS

OECs, immunomodulation, neurotrophic factors, intercellular communication, neuroregeneration

## **1** Introduction

During the last decades, increasing evidence support the hypothesis that glial cells are important players in crucial aspects of neurogenesis, neuronal functions and diseases (1, 2). Indeed, glial cells guide neuronal migration during development, participate in synaptic formation and plasticity, regulate vasculature and blood-brain barrier (BBB),

modulate neuroimmunity, and support neural regeneration (1-3). The term glia, from the Greek "γλία", meaning "glue", was originally assigned assuming that these cells were responsible to keep neural cells together. In the adult central nervous system (CNS) three main types of glial cells can be distinguished: astrocytes and oligodendrocytes, deriving from neural crest, and microglia, which originate from the myeloid lineage. In the peripheral nervous system (PNS), Schwann cells represent the main class of glia. Olfactory ensheathing cells (OECs) are a type of specialized glia cells, restricted to the olfactory system, which play a crucial role in olfactory development and regeneration (4-6). Indeed, the olfactory system has a unique neurogenic niche where unlike most regions of the nervous system, olfactory sensory neurons retain a lifetime regeneration potential (4, 5). Since the olfactory neuroepithelium is in direct contact with the external environment, it has evolved a remarkable ability to recruit sensory neurons during normal cell turnover or after traumatic olfactory nerve injury (7, 8). This unique feature is now widely attributed to the presence of OECs, able to wrap olfactory axons and support olfactory receptor neurons turnover and axonal regeneration (9-11). OECs perform their axon growth-promoting properties and provide structural support by extending thin processes that envelop group of axons as an insulator (Figure 1) (12, 13). Moreover, when new olfactory sensory neurons are generated from stem cells in the olfactory epithelium, OECs establish functional connections along the olfactory neuroaxis (8, 14).

In contrast to neural crest-derived PNS glia and neural tubederived CNS glia, OECs have generally been thought to originate from the olfactory placode (15). However, several studies show that the olfactory placode arises from ontogenetically heterogeneous sources of cells and OECs derive from neural crest, like Schwann cells (16–18). These cells are located in the lamina propria of the olfactory mucosa, as well as the outer layers of the olfactory bulbs, the inner and outer nerve fiber layers (Figure 1) (19).

OECs share many properties with Schwann cells and astrocytes. They express some typical markers such as the p75 neurotrophic receptor (p75NTR), the polysialylated form of neural cell adhesion molecule (PSA-NCAM), and, like astrocytes, they express the glial fibrillary acid protein (GFAP), and the S100 proteins (20, 21). Furthermore, OECs are able to secret high level of growth factors, such as nerve growth factor (NGF), basic fibroblast growth factors (bFBF), brain derived neurotrophic factor (BDNF), glial derived neurotrophic factor (GDNF), ciliary neurotrophic factor (CNTF), neurotrophins NT4, NT5 and neuregulins, which exhibit important functions as neuronal supporting elements (13, 22–24).

In recent years, significant advances have been made in cellular-based therapies, which focus on the restoration, regrowth or replacement of damaged or dysfunctional cells, tissues and organs, in order to treat neurodegenerative diseases (25) and CNS injuries (26–28). Moreover, cell-based approaches, including OEC grafts, have been reported to induce beneficial effects in spinal cord injury (SCI) models. In addition to neuroprotective mechanisms, axon regeneration and remyelination were observed, leading to significant sensory and locomotor functions amelioration (29–31). Thus, OEC transplantation is proposed as a potential therapeutic strategy for SCI, due to their unique characteristics, such as anti-



course through the lamina propria in the PNS. Olfactory nerves and their associated OECs cross through the cribform plate into the CNS, making connections with the olfactory bulb. OEC, olfactory ensheathing cells; CNS, central nervous system; PNS, peripheral nervous system.

neuroinflammation, growth-promoting factor secretion, and debris clearance activity. However, there is a lack of in-depth studies focusing on the phagocytic function of these cells, particularly the molecular and cellular mechanisms involved in this intricate process and on the synergistic effects with neural and mesenchymal stem cells (MSCs) in improving cell differentiation. Exploring these unique features will lead to a better understanding of the role of OECs in development and regeneration and will identify how the use of OECs can be optimized for neural regeneration therapies. These approaches may benefit from accumulating evidence pointing out a significant role of checkpoint therapy in inducing regeneration upon CNS injury (32). Herein we reviewed the current knowledge about the immunomodulatory and antiinflammatory properties of OECs in neuroinflammation, neurodegeneration and during stem cell differentiation. Owing to the strong pro-regenerative properties of OECs, and their unique ability to promote stem cell differentiation, we explored the potential of OEC transplantation for tissue regeneration.

## 2 Immune role of OECs

The olfactory system is continuously exposed to various pathogens since the primary olfactory neurons are in direct contact with the external environment (7, 8). However, most cases of CNS infections do not occur through the olfactory system. In this scenario OECs play a crucial role in protecting CNS structures. Specifically, they participate in innate immune responses, secrete immunoregulatory molecules and exert their phagocytic activity thus maintaining microenvironmental homeostasis, supporting neuron survival and axonal growth (33–35).

### 2.1 Phagocytic activity of OECs

CNS lesions are characterized by neuronal degeneration and death, and by the persistence of cellular and myelinated debris that create an adverse environment for neural survival, germination of neurites and renewal of neurons (36, 37). Since olfactory receptor neurons renew themselves throughout lifetime, a large amount of apoptotic debris is generated continuously (35). Several studies support phagocytic functions of OECs throughout life (35, 38) especially following injury (38, 39). In fact, by switching from a resting state to a phagocytic phenotype to remove axonal debris and bacteria, they protect the olfactory nerve from microbial infections (35, 40, 41). A combination of morphological and phenotypic changes distinguishes reactive OECs from their resting state, including cytoskeletal hypertrophy and rearrangement (34, 42). However, the identification of specific molecular markers capable of discriminating between quiescent and reactive OECs could

better elucidate the molecular mechanisms underlying their activation.

While Schwann cells participate in debris removal mainly by increasing the secretion of several pro-inflammatory molecules, thus recruiting professional phagocytes, including macrophages and neutrophils (43), OECs operate differently (38, 44). Wright et al. showed that OECs repel macrophages in co-culture, by expressing the macrophage migration inhibitory factor (MIF), which would explain the absence of macrophages in the olfactory nerve bundles (45).

In vitro studies reported that OECs possess several phagocytic-related receptors, including toll-like receptor 4 (TLR4), phosphatidylserine and mannose receptors (34, 46, 47). Particularly, during apoptosis, olfactory neurons display the "eat me" signal phosphatidylserine, recognized by OEC phosphatidylserine receptor, leading to the engulfment of apoptotic and necrotic cell debris (33, 44). Milk fat globule-EGF factor 8 (MFGE-8), which interacts with integrin receptors (48), is a bridging molecule that participates in several cell surface-mediated regulatory events. Li et al. demonstrated in vitro that OECs express MFGE-8 when apoptotic debris is added to the culture (49). Moreover, OECs have been reported to adopt a "microglia-like" phenotype showing high levels of CD11 expression after their transplantation into the X-irradiated spinal cord of female Sprague Dawley rats (50). However, in vitro immunolabelling of OECs has revealed that they do not express this microglial marker in physiological conditions (34). Interestingly, Nazareth et al. reported that OECs produce less pro-inflammatory cytokines, compared to Schwann cells and macrophages when exposed to necrotic bodies (37). Conversely, some anti-inflammatory cytokines, such as interleukin-10 (IL-10) and transforming growth factor beta (TGF- $\beta$ ) promote OEC phagocytic activity (49).

In summary, the phagocytic activity of OECs plays a crucial role in creating a favorable environment to promote neuronal turnover, aiding the overall process of neuronal regeneration. Hence, this peculiar feature of OECs may be particularly useful for neural repair therapies including their transplantation after SCI.

### 2.2 OEC-mediated effects during neuroinflammation

As abovementioned, OECs show several unique properties of inflammatory cells, allowing them to modulate immune responses and neuronal pro-regenerative processes. Overall, inflammation is thought to hinder cell differentiation and regeneration but, although OECs are able to secrete a range of pro-inflammatory cytokines and chemokines after injury or infections, they simultaneously promote nervous regeneration.

Following SCI, resident immune cells, including microglia and astrocytes, are activated by injured-released inflammatory

stimuli (51). Indeed, the microenvironment of lesioned CNS switch towards pro-apoptotic and anti-regenerative milieu. Particularly, inflammatory responses in SCI are mainly mediated by pro-inflammatory cytokines and chemokines secreted by reactive astrocytes and microglia. In this scenario, M1-polarized microglia induces astrocyte activation, resulting in chondroitin sulfate proteoglycan (CSPG) deposits and astrocytic scar formation, which limits the spread of inflammation but at the same time hampers axon regeneration (52). Concomitantly, glial cell activation causes the release of specific chemokines and pro-inflammatory cytokines, including IL-1, IL-6, and TNF. These cytokines, by activating their respective cascades, amplify inflammatory responses, alter the microenvironment and promote cell death, therefore blocking axonal regeneration (Figure 2) (53, 54). As a result, inflammatory response induces secondary tissue damage with detrimental consequences to neural tissue and its functions (55). In general terms, inflammatory response maintains a dynamic balance of proinflammatory and anti-inflammatory cytokine release; therefore, understanding the modulation of the inflammatory response mediated by OECs, could be a successful strategy to improve neuronal functional outcome after CNS injury.

OECs are reported to express chemokines/cytokines and their cognate receptors, such as chemokine (CXC motif) ligand 1 (CXCL1), a neurotrophic chemoattractant, which may have a role during embryogenesis or after OECs transplantation in the injured site, CXCL12, CXCL4, chemokine (CX3C motif) ligand 1 (CX3CL1) (56) that have been proven to play pivotal roles in neuroinflammation, acting as a signaling factor for the recruitment of neutrophilis and various leucocytes (57). The inflammatory monocyte chemotactic protein 1 (MCP-1), and its receptor CCR2 specifically mediates monocytes chemotaxis,

which results in the recruitment of macrophages to the site of injury. Moreover, nuclear factor kappa B (NF-KB)-mediated signaling pathway, responsible for microglia and astrocytes activation after SCI, is activated by TROY, a member of the tumor necrosis factor (TNF) receptor superfamily, which has been detected via in situ hybridization and immunohistochemistry investigations in the olfactory system (58). OECs or OECreleased molecules are able to inhibit NF-KB activation, so exerting a neuroprotective role after CNS injury. OECs also release several signaling molecules, such as TNF and IL-1B, to recruit macrophages, thus modulating inflammation and neurodegeneration (14, 44, 59). In this context, OECs could modulate microglia-astrocyte responses by secreting antiinflammatory cytokines such as IL-4, IL-10, IL-13 and TGF-β, capable to downregulate the pro-inflammatory factors IL-1B, TNF and IL-6 (Figure 2) (60–62). A recent study showed that IL-1 $\alpha$ and IL-1 $\beta$ , which are significantly involved in inflammatory responses, were down-regulated after OEC transplantation at the injury site. This response is probably related to IL-1 receptor antagonist (IL-1Ra) mechanism, which is a competitive inhibitor of IL-1 by binding to its receptor (Figure 2). Therefore, OECs, reaching the site of the lesion, are subjected to pro-inflammatory factors released by the activated microglia, and secrete IL-1Ra in response, thus reducing microglial activation and proinflammatory factor production and limiting microgliamediated pro-inflammatory cytokine release (63).

It is worth noticing that the abovementioned OEC-derived anti-inflammatory factors participate in modulating cell survival, proliferation and migration, thus reducing glial scar and promoting regeneration after SCI (64). IL-4 and TGF- $\beta$  have a direct impact on neural survival given their modulatory effects on acute and chronic immune cell responses and on their



#### FIGURE 2

Schematic overview of the involvement of OECs in inflammation modulation after SCI. Neuronal damage induces pathological increasing of inflammatory responses, which promotes microglia polarization from a resting state to a M1-phenotype and astrocyte activation. OECs are able to modulate these inflammatory events by interacting directly or indirectly with microglia and astrocytes, thus ameliorating the detrimental condition of the altered microenvironment. OEC, olfactory ensheathing cell; SCI, spinal cord injury; TNF, tumor necrosis factor; IL, interleukin; TGF- $\beta$ , transforming growth factor  $\beta$ , IL-1Ra: interleukin-1 receptor antagonist.

expression of detrimental molecules including nitric oxide (NO), reactive oxygen species (ROS), caspase and their secretion of neurotrophins (65).

Taken together these findings suggest that OECs delay the activation of microglia or macrophages and reduce the peak of the immune response, leading to neuroprotection against inflammatory damage.

# 3 OEC bystander effects on cell fate and differentiation

The ability of OECs in regulating neuroprotection is enhanced by the release of several protective factors in the microenvironment as OEC-conditioned medium (OEC-CM) promotes the differentiation of neural stem cells (NSCs) (66). Specifically, using OEC-CM, it has been shown that soluble factors larger than 30 kDa, which are secreted by OECs, promote migration, differentiation and maturation of NSCs within 7 days. By immunocytochemical analysis, it has been shown that NSCs in contact with OEC-CM, exhibited an up-regulation of neurofilament (NF), beta-III-tubulin (TUJ1), GFAP and a downregulation of nestin, suggesting a differentiation of NSCs toward neuronal and astrocytic lineages. In addition, the presence in NSCs of synapsin-1, which is involved in the neurotransmitter release mechanism, has also been demonstrated, supporting the effect of OEC-CM in driving and/or stimulating neuronal differentiation. This study also claims that differentiation of NSCs, promoted by OECs, also occurs through indirect contact (67). OECs also exert their trophic effects directly through the secretion of factors involved in neurogenesis, neural differentiation and response, including both NGF and BDNF, small proteins including neurturin (NTN), CNTF, GDNF (68, 69), and heavier soluble factors including secreted protein acidic and cysteine rich (SPARC), sonic hedgehog protein (SHH), matrix metalloproteinase 2 (MMP 2), fibronectin, and laminin (70–72) (Table 1). Moreover, TGF- $\beta$ 3 secreted by OECs is involved in the regulation of neuronal

differentiation, negatively regulating Yes-associated protein (YAP) (76). The potential of OECs to induce differentiation of NSCs into neurons has also been demonstrated by functional electrophysiological studies that showed that NSC-derived neurons exposed to OEC-CM acquire active electrophysiological properties, expressing sodium and potassium channels suitable for onset of action potentials similar to primary neuronal cells (66). In vivo studies demonstrated that OECs are able to promote NSC differentiation into dopaminergic neurons or cholinergic neurons, pointing out that OECs can induce NSC differentiation toward a specific neuron subtype (77, 78). OEC-induced effects would be exerted by influencing Wnt/beta-catenin signaling pathway, which is important in the proliferation and self-renewal of adult NSCs (79, 80). Indeed, it was shown that CM from Wnt-activated OECs (wOEC-CM) stimulates the proliferation and differentiation of NSCs, by increasing the percentage of Ki67/Sox2 double positive cells, maintaining Nestin expression under differentiation condition, but also stimulating NSC differentiation into Tuj1positive neurons (81).

Many reports have shown that hypoxic preconditioned stem cells survive longer, exhibiting an efficient neuronal differentiation and showing enhanced paracrine effects (82, 83). Wang et al. demonstrated that CM from hyperthermiaconditioned OECs induces NSC neural differentiation more efficiently, thanks to the upregulation of HIF-1 $\alpha$ , leading to synergistic effects that improve differentiation (84). By using OEC-CM under hypoxic condition, olfactory mucosa MSCs (OM-MSCs) are stimulated to differentiate into dopaminergic neurons. Specifically, OEC-CM under hypoxia upregulates transcriptional factors mediated by HIF-1 $\alpha$  and it is involved in the development of dopaminergic neurons from OM-MSCs (85).

MSCs, including adipose tissue-derived MSCs (ASCs), are a type of non-hematopoietic stem cells which under appropriate conditions can give rise to several precursors (86–90). OEC-CM is also implicated in the differentiation ASCs toward a neuronal phenotype (91). ASCs treated with OEC-CM expressed markers

 TABLE 1
 OEC released factors involved in neural differentiation and neurogenesis.

Factors	Molecular weight	Functions	References
Brain Derived Neurotrophic Factor (BDNF)	26.7 kDa	Involved in the promotion of Schwann cell migration	(69)
Ciliary Neurotrophic Factor (CNTF)	22.9 kDa	Support neurogenesis	(67)
Fibronectin	440 kDa	Promote neural progenitor cell migration	(73)
Laminin	400 kDa	Involved in neural progenitor cell differentiation	(74)
Matrix Metalloproteinase 2 (MMP2)	67 kDa	Important for neural cell migration	(75)
Nerve Growth Factor (NGF)	26.7 kDa	Involved in the promotion of Schwann cell migration	(69)
Neurturin (NTN)	23.6 kDa	Support neurogenesis	(67)
Sonic Hedgehog protein (Shh)	67 kDa	Induce NSC differentiation into neurons	(72)
Secreted Protein Acidic and Cysteine Rich (SPARC)	43 kDa	Implicated in neural differentiation and in neurite extension	(70)

of progenitor and mature neurons, including Nestin, protein gene product 9.5 (PGP 9.5), and microtubule-associated protein 2 (MAP2) in a time-dependent manner and exhibited neuron like morphology, while they were negative for GFAP and A2B5, markers of astrocytes and oligodendrocytes, respectively (92). In addition, although a significant increase of Nestin, PGP 9.5, Synapsin I, and GFAP was reported, MAP2 was identified as the most representative, thus suggesting a greater tendency toward the neuronal phenotype (93). This result is confirmed by another study where a neural-like connexin expression was induced in ASCs after OEC-CM treatment (94-98). On the other hand, when ASCs were co-cultured with OECs using 3D collagen scaffolds, they differentiated into cells with OEC-like morphology and were reported to be p75NTR and Nestin positive and GFAP negative. These co-cultured ASCs also expressed various functional markers of mature OECs: BDNF, GDNF and the myelin proteolipid protein (PLP). Thus, these results demonstrate that using specific scaffolds, ASCs might differentiate into OEC-like cells in vitro (99). Altogether, it can be inferred that OECs play a key role in cell differentiation toward a neural type and are able to prompt MSC differentiation towards neural phenotype and even to mature OECs. As such, these intrinsic properties of OECs may be relevant for therapeutic approaches aiming at CNS tissue regeneration.

## 4 OECs for tissue regeneration and transplantation

In recent years, OECs have been investigated for their reparative ability following acute or chronic lesions that involve CNS. As already mentioned, it appears that OECs may play a crucial role in the treatment of SCI (100-103). Usually, SCI severely affects CNS microenvironment, leading to a series of deleterious processes such as inflammation and hypoxia, and progressive cell death (104). OECs exhibit several characteristics that enable them to have beneficial effects in neuro-repairing potential. They are able to reduce the inflammatory response following injury, thereby decreasing the size of the glial scar and promoting angiogenesis. In addition, they promote regrowth, plasticity and remyelination of axons (105-107). OECs can also interact with resident cell populations, particularly astrocytes and meningeal cells, either within the window of the glial scar formation or once the scar has already established (108, 109). Thus, in addition to penetrate glial tissue, OECs also produce extracellular matrix proteases and can reduce astrocytic reactivity. Overall, these properties may reduce glial scar formation and all consequential limitations, which strongly limit axonal regrowth and injury bridging (110). In SCI animal models, grafted OECs exhibit the ability to promote axon

regrowth and propagation (30, 100, 111). In particular, OEC transplantation improves sensorimotor and autonomic nerve recovery, also reducing neuropathic pain due to SCI. OECs secrete a number of neurotrophic factors, which allow the establishment of a favorable microenvironment for the regrowth of damaged axons. Undoubtedly, it is crucial to have functional recovery in transplanted animals in order to consider OEC transplants a successful therapy for the treatment of SCI (107). A study of Ramon-Cueto et al., revealed that adult rats undergoing spinal cord resection and subsequent OEC transplantation, showed both functional and structural recovery. In particular, from 3 to 7 months after surgery, all transplanted animals improved locomotor functions and sensorimotor reflexes (103). To show actual recoveries in transplanted animals, electrophysiological studies were also carried out, demonstrating that animals with transplanted OECs not only exhibited functional recovery, but also showed recovery of action or evoked potentials (112, 113). Studies on OEC transplantation have also been carried out in human clinical in many countries around the world. Completed clinical trials have demonstrated the safety and efficacy of OEC transplantation, but recovery in patients is often highly variable. This variability may be related to a number of factors, such as difficulties in establishing master cell banks and working cell banks, cell purity, and transplantation techniques. Fortyfour eligible trials, involving 1,266 SCI patients, investigated several cell-based treatments to improve functional independent misure (FIM) score. Among them, OEC transplantation proved to ameliorate the FIM score at 6 months, thus improving disease prognosis (114). However, a common consequence in these studies is the poor survival of transplanted cells, with survival rates ranging from 0.3 percent to 3 percent. This issue is probably related to the fact that when OECs are isolated, expanded in vitro, and then transplanted into the injury site, their therapeutic potential is reduced, probably due to bleeding, damaged tissue and anatomical structures and hostile microenvironment present in the lesioned area (115-117). Therefore, in vitro models challenging OECs, by mimicking the injured tissue microenvironment, are needed. For example, it has been found that preconditioned OECs showed increased migratory, phagocytic and immunomodulatory capacities. To improve their efficacy and yield upon graft, cells could be then exposed to a low oxygen level, or they could grow into threedimensional scaffolds before being transplanted into the lesion (116).

Another effective strategy for SCI treatment is the cotransplantation of NSCs and OECs. Indeed, co-grafting of NSCs and OECs ameliorate SCI by inhibiting receptor-interacting protein kinase 3 (RIP3)/mixed lineage kinase domain-like protein (MLKL)mediated necroptosis and stimulating NSC proliferation in the medulla. Evidence reports that OECs are able to increase NSCs

proliferation and differentiation and, importantly, co-grafts significantly support NSCs survival, opening the way for a potential stem cells-based regenerative approach. In this way, neural regeneration could be improved exploiting the synergistic effect of NSCs and OECs (118). In addition, a study by He Y. t al., showed that curcumin-activated OECs (aOECs) effectively improve neuronal differentiation of NSCs even under conditions of inflammation, and co-transplantation of aOECs and NSCs enhaces the neurological recovery of rats after SCI, providing a hopeful strategy for SCI repair by co-transplantation of aOECs and NSCs (119). Besides OEC transplantation, OECs-CM has also been shown to have therapeutic effects for SCI, enhancing functional recovery and axonal regeneration probably because of various factors previously secreted by OECs in their culture medium (120). In this context, exosomes derived from OECs (OEC-Exo) also promote neuronal survival and improve axon condition, facilitating functional recovery following SCI. OEC-Exo can be internalized by microglia/ macrophages and are able to modulate their polarization. The main ability of OEC-Exo consists in an immunomodulatory function that shape immune microenvironment towards a pro-regenerative phenotype, supporting OEC-Exo as neuroprotective and regenerative strategy for CNS diseases (121). Furthermore, OECs secrete, via exosomes, alpha B-crystallin (CryAB), an antiinflammatory protein, leading to an intercellular immune response. Thus, CryAB, together with other OEC-secreted factors, may ameliorate the hostile growth environment created by neurotoxic reactive astrocytes following CNS injury (122).

SCI microenvironment is characterized by a prevalence of M1-like pro-inflammatory macrophages over M2-like. This phenomenon results in a microenvironment that is unfavorable for cell differentiation and regeneration. Therefore, for a better potential regenerative strategy, a fundamental role is played by immune cell modulation (123, 124). Macrophages are a prominent population in SCI microenvironment, also able to alter the activity of transplanted OECs. However, the interaction appears to be reciprocal, as OECs express MIFs and can also lead to reduced macrophage recruitment in vitro (45). To enhance this interaction in favor of OECs by improving the cellular microenvironment at the injury level, a study of vascular endothelial growth factor (VEGF) and platelet-derived growth factor (PDGF) modulation in the SCI microenvironment was carried out. It was shown that CM from macrophages exposed to PDGF or combined VEGF and PDGF, under inflammatory conditions, increased OEC phagocytosis, also modulating the expression of genes related to nerve repair. Specifically, both PDGF and VEGF/PDGF reduced pro-inflammatory cytokines (i.e., TNF) by decreasing NF-KB translocation, promoting phagocytosis of myelin debris. For this reason, administering growth factors before OEC transplantation could improve transplant success and neural recovery (125).

## **5** Conclusions

Our knowledge of the properties, functions, and therapeutic potential of OECs is markedly increasing. OECs can be considered as a good candidate for cell-replacement and have shown remarkable capabilities to exert neuroprotective mechanisms. The uniqueness of OECs appears to collaborate with other recruited cell types to orchestrate the molecular signaling responsible for resolving the inflammatory state and creating a favorable environment for neural regeneration.

However, the development of human OEC transplants for clinical application in SCI still requires an in-depth understanding of the cellular and molecular biological characteristics of OECs. It seems now clear that OECs expanded in vitro and grafted back in vivo show limited therapeutic potential, probably due to the hostile microenvironment at the damaged tissue. In fact, a major issue limiting spinal cord regeneration is also the poor survival of transplanted cells (126). In order to describe the therapeutic potential of OECs it appears critical to characterize OEC gene expression aiming at identifying OEC-specific markers. Indeed, the most used marker to identify OECs, p75NTR, is also expressed in vitro by Schwann cells (10), astrocytes and lamina propria MSCs (127). The lack of a solid method for OECs identification, isolation and purification is among the main factors limiting reproducibility and reliability of transplantation studies. Furthermore, without a unique method for OEC identification, it is possible that their repair capacity is influenced by the presence of the various cells types co-existing alongside OECs.

One of the most effective approaches in transplantation of OECs is co-grafting with NSCs, achieving better therapeutic effects. Indeed OECs, by releasing trophic factors into the microenvironment, also play an important role in promoting the differentiation of NSCs, able to change their morphology, stimulating their differentiation towards mature neurons.

Despite the variability of results reported and limiting factors, OECs should be considered as valuable cell-based approach for SCI and a potential candidate to promote cell differentiation and regeneration. Finally, a deeper understanding of OEC anti-inflammatory properties and their interplay with other cells involved in neuro-repairing is crucial for the development of future therapies, using transplantation of OECs to treat neural injuries.

## Author contributions

Conceptualization: SD, SD'A, NV, and RP; writing—original draft preparation: SD, SD'A, NV, and RP; writing—review and editing: all authors; visualization: SD, SD'A, CA, AP, and FT;

supervision: DLF, AZ, RG, DT, GLV, NV, and RP. All authors contributed to the article and approved the submitted version.

## Funding

SD, CA, and AP were supported by the PhD program in Biotechnologies, Biometec, University of Catania. FT was supported by Fondazione Umberto Veronesi. NV was supported by the PON AIM R&I 2014-2020-E66C18001240007.

## Acknowledgments

The authors thank Servier Medical Art (smart.servier.com, accessed on 13 November 2022)

## References

1. Lobsiger CS, Cleveland DW. Glial cells as intrinsic components of non-cellautonomous neurodegenerative disease. *Nat Neurosci* (2007) 10(11):1355–60. doi: 10.1038/nn1988

2. Barres BA. The mystery and magic of glia: A perspective on their roles in health and disease. *Neuron* (2008) 60(3):430–40. doi: 10.1016/j.neuron.2008.10.013

3. Wang X, Jiang C, Zhang Y, Chen Z, Fan H, Zhang Y, et al. The promoting effects of activated olfactory ensheathing cells on angiogenesis after spinal cord injury through the PI3K/Akt pathway. *Cell Biosci* (2022) 12(1):23. doi: 10.1186/s13578-022-00765-y

4. Graziadei GA, Graziadei PP. Neurogenesis and neuron regeneration in the olfactory system of mammals. II. Degeneration and reconstitution of the olfactory sensory neurons after axotomy. *J Neurocytol* (1979) 8(2):197–213. doi: 10.1007/BF01175561

5. Graziadei PP, Graziadei GA. Neurogenesis and neuron regeneration in the olfactory system of mammals. i. morphological aspects of differentiation and structural organization of the olfactory sensory neurons. *J Neurocytol* (1979) 8 (1):1–18. doi: 10.1007/BF01206454

6. Mackay-Sim A, Kittel P. Cell dynamics in the adult mouse olfactory epithelium: A quantitative autoradiographic study. *J Neurosci* (1991) 11(4):979-84. doi: 10.1523/JNEUROSCI.11-04-00979.1991

7. Schwob JE. Neural regeneration and the peripheral olfactory system. Anat Rec (2002) 269(1):33-49. doi: 10.1002/ar.10047

8. Leung CT, Coulombe PA, Reed RR. Contribution of olfactory neural stem cells to tissue maintenance and regeneration. *Nat Neurosci* (2007) 10(6):720–6. doi: 10.1038/nn1882

9. Doucette R. Glial influences on axonal growth in the primary olfactory system. *Glia* (1990) 3(6):433-49. doi: 10.1002/glia.440030602

10. Chen CR, Kachramanoglou C, Li D, Andrews P, Choi D. Anatomy and cellular constituents of the human olfactory mucosa: A review. *J Neurol Surg B Skull Base* (2014) 75(5):293–300. doi: 10.1055/s-0033-1361837

11. Schwob JE, Jang W, Holbrook EH, Lin B, Herrick DB, Peterson JN, et al. Stem and progenitor cells of the mammalian olfactory epithelium: Taking poietic license. *J Comp Neurol* (2017) 525(4):1034–54. doi: 10.1002/cne.24105

12. Rieger A, Deitmer JW, Lohr C. Axon-glia communication evokes calcium signaling in olfactory ensheathing cells of the developing olfactory bulb. *Glia* (2007) 55(4):352–9. doi: 10.1002/glia.20460

13. Ursavas S, Darici H, Karaoz E. Olfactory ensheathing cells: Unique glial cells promising for treatments of spinal cord injury. *J Neurosci Res* (2021) 99(6):1579–97. doi: 10.1002/jnr.24817

14. Roet KC, Verhaagen J. Understanding the neural repair-promoting properties of olfactory ensheathing cells. *Exp Neurol* (2014) 261:594-609. doi: 10.1016/j.expneurol.2014.05.007

15. Shields SD, Moore KD, Phelps PE, Basbaum AI. Olfactory ensheathing glia express aquaporin 1. J Comp Neurol (2010) 518(21):4329–41. doi: 10.1002/cne.22459

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

16. Barraud P, Seferiadis AA, Tyson LD, Zwart MF, Szabo-Rogers HL, Ruhrberg C, et al. Neural crest origin of olfactory ensheathing glia. *Proc Natl Acad Sci U S A* (2010) 107(49):21040–5. doi: 10.1073/pnas.1012248107

17. Forni PE, Taylor-Burds C, Melvin VS, Williams T, Wray S. Neural crest and ectodermal cells intermix in the nasal placode to give rise to GnRH-1 neurons, sensory neurons, and olfactory ensheathing cells. *J Neurosci* (2011) 31(18):6915–27. doi: 10.1523/JNEUROSCI.6087-10.2011

18. Katoh H, Shibata S, Fukuda K, Sato M, Satoh E, Nagoshi N, et al. The dual origin of the peripheral olfactory system: Placode and neural crest. *Mol Brain* (2011) 4:34. doi: 10.1186/1756-6606-4-34

19. Yao R, Murtaza M, Velasquez JT, Todorovic M, Rayfield A, Ekberg J, et al. Olfactory ensheathing cells for spinal cord injury: Sniffing out the issues. *Cell Transpl* (2018) 27(6):879–89. doi: 10.1177/0963689718779353

20. Pellitteri R, Spatuzza M, Stanzani S, Zaccheo D. Biomarkers expression in rat olfactory ensheathing cells. *Front Biosci (Schol Ed)* (2010) 2(1):289–98. doi: 10.2741/s64

21. Barton MJ, John JS, Clarke M, Wright A, Ekberg J. The glia response after peripheral nerve injury: A comparison between schwann cells and olfactory ensheathing cells and their uses for neural regenerative therapies. *Int J Mol Sci* (2017) 18(2):287–306. doi: 10.3390/ijms18020287

22. Boruch AV, Conners JJ, Pipitone M, Deadwyler G, Storer PD, Devries GH, et al. Neurotrophic and migratory properties of an olfactory ensheathing cell line. *Glia* (2001) 33(3):225–9. doi: 10.1002/1098-1136(200103)33:3<225::AID-GLIA1021>3.0.CO;2-Y

23. Stepanova OV, Voronova AD, Chadin AV, Fursa GA, Karsuntseva EK, Valikhov MP, et al. Neurotrophin-3 enhances the effectiveness of cell therapy in chronic spinal cord injuries. *Bull Exp Biol Med* (2022) 173(1):114–8. doi: 10.1007/s10517-022-05504-4

24. Russo C, Patane M, Russo A, Stanzani S, Pellitteri R. Effects of ghrelin on olfactory ensheathing cell viability and neural marker expression. *J Mol Neurosci* (2021) 71(5):963–71. doi: 10.1007/s12031-020-01716-3

25. Lo Furno D, Mannino G, Giuffrida R. Functional role of mesenchymal stem cells in the treatment of chronic neurodegenerative diseases. *J Cell Physiol* (2018) 233(5):3982–99. doi: 10.1002/jcp.26192

26. Golchin A, Shams F, Kangari P, Azari A, Hosseinzadeh S. Regenerative medicine: Injectable cell-based therapeutics and approved products. *Adv Exp Med Biol* (2020) 1237:75–95. doi: 10.1007/5584\_2019\_412

27. Mannino G, Russo C, Longo A, Anfuso CD, Lupo G, Lo Furno D, et al. Potential therapeutic applications of mesenchymal stem cells for the treatment of eye diseases. *World J Stem Cells* (2021) 13(6):632-44. doi: 10.4252/wjsc.v13.i6.632

 Mannino G, Cristaldi M, Giurdanella G, Perrotta RE, Lo Furno D, Giuffrida R, et al. ARPE-19 conditioned medium promotes neural differentiation of adiposederived mesenchymal stem cells. World J Stem Cells (2021) 13(11):1783–96. doi: 10.4252/wjsc.v13.i11.1783 29. Menezes K, Nascimento MA, Goncalves JP, Cruz AS, Lopes DV, Curzio B, et al. Human mesenchymal cells from adipose tissue deposit laminin and promote regeneration of injured spinal cord in rats. *PloS One* (2014) 9(5):e96020. doi: 10.1371/journal.pone.0096020

30. Lu J, Feron F, Mackay-Sim A, Waite PM. Olfactory ensheathing cells promote locomotor recovery after delayed transplantation into transected spinal cord. *Brain* (2002) 125(Pt 1):14–21. doi: 10.1093/brain/awf014

31. Pearse DD, Sanchez AR, Pereira FC, Andrade CM, Puzis R, Pressman Y, et al. Transplantation of schwann cells and/or olfactory ensheathing glia into the contused spinal cord: Survival, migration, axon association, and functional recovery. *Glia* (2007) 55(9):976–1000. doi: 10.1002/glia.20490

32. Taylor MJ, Thompson AM, Alhajlah S, Tuxworth RI, Ahmed Z. Inhibition of Chk2 promotes neuroprotection, axon regeneration, and functional recovery after CNS injury. *Sci Adv* (2022) 8(37):eabq2611. doi: 10.1126/sciadv.abq2611

33. He BR, Xie ST, Wu MM, Hao DJ, Yang H. Phagocytic removal of neuronal debris by olfactory ensheathing cells enhances neuronal survival and neurite outgrowth *via* p38MAPK activity. *Mol Neurobiol* (2014) 49(3):1501–12. doi: 10.1007/s12035-013-8588-2

34. Hao DJ, Liu C, Zhang L, Chen B, Zhang Q, Zhang R, et al. Lipopolysaccharide and curcumin Co-stimulation potentiates olfactory ensheathing cell phagocytosis *Via* enhancing their activation. *Neurotherapeutics* (2017) 14(2):502–18. doi: 10.1007/s13311-016-0485-8

35. Su Z, Chen J, Qiu Y, Yuan Y, Zhu F, Zhu Y, et al. Olfactory ensheathing cells: the primary innate immunocytes in the olfactory pathway to engulf apoptotic olfactory nerve debris. *Glia* (2013) 61(4):490–503. doi: 10.1002/glia.22450

36. Vargas ME, Barres BA. Why is wallerian degeneration in the CNS so slow? Annu Rev Neurosci (2007) 30:153–79. doi: 10.1146/annurev.neuro.30.051606.094354

37. Nazareth L, Shelper TB, Chacko A, Basu S, Delbaz A, Lee JYP, et al. Key differences between olfactory ensheathing cells and schwann cells regarding phagocytosis of necrotic cells: implications for transplantation therapies. *Sci Rep* (2020) 10(1):18936. doi: 10.1038/s41598-020-75850-8

38. Nazareth L, Lineburg KE, Chuah MI, Tello Velasquez J, Chehrehasa F, St John JA, et al. Olfactory ensheathing cells are the main phagocytic cells that remove axon debris during early development of the olfactory system. *J Comp Neurol* (2015) 523(3):479–94. doi: 10.1002/cne.23694

39. Wewetzer K, Kern N, Ebel C, Radtke C, Brandes G. Phagocytosis of O4+ axonal fragments *in vitro* by p75- neonatal rat olfactory ensheathing cells. *Glia* (2005) 49(4):577–87. doi: 10.1002/glia.20149

40. Panni P, Ferguson IA, Beacham I, Mackay-Sim A, Ekberg JA, St John JA. Phagocytosis of bacteria by olfactory ensheathing cells and schwann cells. *Neurosci Lett* (2013) 539:65–70. doi: 10.1016/j.neulet.2013.01.052

41. Jiang Y, Guo J, Tang X, Wang X, Hao D, Yang H. The immunological roles of olfactory ensheathing cells in the treatment of spinal cord injury. *Front Immunol* (2022) 13:881162. doi: 10.3389/fimmu.2022.881162

42. Li Y, Huo S, Fang Y, Zou T, Gu X, Tao Q, et al. ROCK inhibitor Y27632 induced morphological shift and enhanced neurite outgrowth-promoting property of olfactory ensheathing cells *via* YAP-dependent up-regulation of L1-CAM. *Front Cell Neurosci* (2018) 12:489. doi: 10.3389/fncel.2018.00489

43. Hartlehnert M, Derksen A, Hagenacker T, Kindermann D, Schafers M, Pawlak M, et al. Schwann cells promote post-traumatic nerve inflammation and neuropathic pain through MHC class II. *Sci Rep* (2017) 7(1):12518. doi: 10.1038/s41598-017-12744-2

44. Nazareth L, St John J, Murtaza M, Ekberg J. Phagocytosis by peripheral glia: Importance for nervous system functions and implications in injury and disease. *Front Cell Dev Biol* (2021) 9:660259. doi: 10.3389/fcell.2021.660259

45. Wright AA, Todorovic M, Murtaza M, St John JA, Ekberg JA. Macrophage migration inhibitory factor and its binding partner HTRA1 are expressed by olfactory ensheathing cells. *Mol Cell Neurosci* (2020) 102:103450. doi: 10.1016/j.mcn.2019.103450

46. Vincent AJ, Choi-Lundberg DL, Harris JA, West AK, Chuah MI. Bacteria and PAMPs activate nuclear factor kappaB and gro production in a subset of olfactory ensheathing cells and astrocytes but not in schwann cells. *Glia* (2007) 55 (9):905–16. doi: 10.1002/glia.20512

47. Carvalho LA, Nobrega AF, Soares ID, Carvalho SL, Allodi S, Baetas-da-Cruz W, et al. The mannose receptor is expressed by olfactory ensheathing cells in the rat olfactory bulb. *J Neurosci Res* (2013) 91(12):1572–80. doi: 10.1002/jnr.23285

48. Hanayama R, Tanaka M, Miwa K, Shinohara A, Iwamatsu A, Nagata S. Identification of a factor that links apoptotic cells to phagocytes. *Nature* (2002) 417 (6885):182–7. doi: 10.1038/417182a

 Li Y, Zou T, Xue L, Yin ZQ, Huo S, Xu H. TGF-beta1 enhances phagocytic removal of neuron debris and neuronal survival by olfactory ensheathing cells via integrin/MFG-E8 signaling pathway. *Mol Cell Neurosci* (2017) 85:45–56. doi: 10.1016/j.mcn.2017.08.006 50. Lankford KL, Sasaki M, Radtke C, Kocsis JD. Olfactory ensheathing cells exhibit unique migratory, phagocytic, and myelinating properties in the X-irradiated spinal cord not shared by schwann cells. *Glia* (2008) 56(15):1664–78. doi: 10.1002/glia.20718

51. Orr MB, Gensel JC. Spinal cord injury scarring and inflammation: Therapies targeting glial and inflammatory responses. *Neurotherapeutics* (2018) 15(3):541–53. doi: 10.1007/s13311-018-0631-6

52. Yu SS, Li ZY, Xu XZ, Yao F, Luo Y, Liu YC, et al. M1-type microglia can induce astrocytes to deposit chondroitin sulfate proteoglycan after spinal cord injury. *Neural Regener Res* (2022) 17(5):1072–9. doi: 10.4103/1673-5374.324858

53. Ramesh G, MacLean AG, Philipp MT. Cytokines and chemokines at the crossroads of neuroinflammation, neurodegeneration, and neuropathic pain. *Mediators Inflamm* (2013) 2013:480739. doi: 10.1155/2013/480739

54. Jin X, Yamashita T. Microglia in central nervous system repair after injury. J Biochem (2016) 159(5):491–6. doi: 10.1093/jb/mvw009

55. David S, Kroner A. Repertoire of microglial and macrophage responses after spinal cord injury. *Nat Rev Neurosci* (2011) 12(7):388–99. doi: 10.1038/nrn3053

56. Yan Y, Su J, Zhang Z. The CXCL12/CXCR4/ACKR3 response axis in chronic neurodegenerative disorders of the central nervous system: Therapeutic target and biomarker. *Cell Mol Neurobiol* (2022) 42(7):2147–56. doi: 10.1007/s10571-021-01115-1

57. Vincent AJ, Taylor JM, Choi-Lundberg DL, West AK, Chuah MI. Genetic expression profile of olfactory ensheathing cells is distinct from that of schwann cells and astrocytes. *Glia* (2005) 51(2):132–47. doi: 10.1002/glia.20195

58. Hisaoka T, Morikawa Y, Kitamura T, Senba E. Expression of a member of tumor necrosis factor receptor superfamily, TROY, in the developing mouse brain. *Brain Res Dev Brain Res* (2003) 143(1):105–9. doi: 10.1016/S0165-3806(03)00101-9

59. Su Z, He C. Olfactory ensheathing cells: biology in neural development and regeneration. *Prog Neurobiol* (2010) 92(4):517-32. doi: 10.1016/j.pneurobio.2010.08.008

60. Donnelly DJ, Popovich PG. Inflammation and its role in neuroprotection, axonal regeneration and functional recovery after spinal cord injury. *Exp Neurol* (2008) 209(2):378–88. doi: 10.1016/j.expneurol.2007.06.009

61. Cataldi C, Mari NL, Lozovoy MAB, Martins LMM, Reiche EMV, Maes M, et al. Proinflammatory and anti-inflammatory cytokine profiles in psoriasis: use as laboratory biomarkers and disease predictors. *Inflammation Res* (2019) 68(7):557–67. doi: 10.1007/s00011-019-01238-8

62. Daines JM, Schellhardt L, Wood MD. The role of the IL-4 signaling pathway in traumatic nerve injuries. *Neurorehabil Neural Repair* (2021) 35(5):431-43. doi: 10.1177/15459683211001026

63. Zhang L, Zhuang X, Kotitalo P, Keller T, Krzyczmonik A, Haaparanta-Solin M, et al. Intravenous transplantation of olfactory ensheathing cells reduces neuroinflammation after spinal cord injury *via* interleukin-1 receptor antagonist. *Theranostics* (2021) 11(3):1147–61. doi: 10.7150/thno.52197

64. Park J, Decker JT, Margul DJ, Smith DR, Cummings BJ, Anderson AJ, et al. Local immunomodulation with anti-inflammatory cytokine-encoding lentivirus enhances functional recovery after spinal cord injury. *Mol Ther* (2018) 26(7):1756–70. doi: 10.1016/j.ymthe.2018.04.022

65. Khankan RR, Griffis KG, Haggerty-Skeans JR, Zhong H, Roy RR, Edgerton VR, et al. Olfactory ensheathing cell transplantation after a complete spinal cord transection mediates neuroprotective and immunomodulatory mechanisms to facilitate regeneration. *J Neurosci* (2016) 36(23):6269–86. doi: 10.1523/JNEUROSCI.0085-16.2016

66. Duan D, Rong M, Zeng Y, Teng X, Zhao Z, Liu B, et al. Electrophysiological characterization of NSCs after differentiation induced by OEC conditioned medium. *Acta Neurochir (Wien)* (2011) 153(10):2085–90. doi: 10.1007/s00701-011-0955-z

67. Sethi R, Sethi R, Redmond A, Lavik E. Olfactory ensheathing cells promote differentiation of neural stem cells and robust neurite extension. *Stem Cell Rev Rep* (2014) 10(6):772–85. doi: 10.1007/s12015-014-9539-7

68. Lipson AC, Widenfalk J, Lindqvist E, Ebendal T, Olson L. Neurotrophic properties of olfactory ensheathing glia. *Exp Neurol* (2003) 180(2):167–71. doi: 10.1016/S0014-4886(02)00058-4

69. Cao L, Zhu YL, Su Z, Lv B, Huang Z, Mu L, et al. Olfactory ensheathing cells promote migration of schwann cells by secreted nerve growth factor. *Glia* (2007) 55 (9):897–904. doi: 10.1002/glia.20511

70. Au E, Richter MW, Vincent AJ, Tetzlaff W, Aebersold R, Sage EH, et al. SPARC From olfactory ensheathing cells stimulates schwann cells to promote neurite outgrowth and enhances spinal cord repair. *J Neurosci* (2007) 27(27):7208–21. doi: 10.1523/JNEUROSCI.0509-07.2007

71. Higginson JR, Barnett SC. The culture of olfactory ensheathing cells (OECs)-a distinct glial cell type. *Exp Neurol* (2011) 229(1):2–9. doi: 10.1016/ j.expneurol.2010.08.020

72. Zhang X, Klueber KM, Guo Z, Cai J, Lu C, Winstead WI, et al. Induction of neuronal differentiation of adult human olfactory neuroepithelial-derived progenitors. *Brain Res* (2006), 1073–1074:109–19. doi: 10.1016/j.brainres.2005.12.059

73. Kearns SM, Laywell ED, Kukekov VK, Steindler DA. Extracellular matrix effects on neurosphere cell motility. *Exp Neurol* (2003) 182(1):240–4. doi: 10.1016/S0014-4886(03)00124-9

74. Wilkinson AE, Kobelt LJ, Leipzig ND. Immobilized ECM molecules and the effects of concentration and surface type on the control of NSC differentiation. *J BioMed Mater Res A* (2014) 102(10):3419–28. doi: 10.1002/jbm.a.35001

75. Wang L, Zhang ZG, Zhang RL, Gregg SR, Hozeska-Solgot A, LeTourneau Y, et al. Matrix metalloproteinase 2 (MMP2) and MMP9 secreted by erythropoietinactivated endothelial cells promote neural progenitor cell migration. *J Neurosci* (2006) 26(22):5996–6003. doi: 10.1523/JNEUROSCI.5380-05.2006

76. Ge L, Zhang C, Xie H, Zhuo Y, Xun C, Chen P, et al. A phosphoproteomics study reveals a defined genetic program for neural lineage commitment of neural stem cells induced by olfactory ensheathing cell-conditioned medium. *Pharmacol Res* (2021) 172:105797. doi: 10.1016/j.phrs.2021.105797

77. Shukla S, Chaturvedi RK, Seth K, Roy NS, Agrawal AK. Enhanced survival and function of neural stem cells-derived dopaminergic neurons under influence of olfactory ensheathing cells in parkinsonian rats. *J Neurochem* (2009) 109(2):436–51. doi: 10.1111/j.1471-4159.2009.05983.x

78. Srivastava N, Seth K, Khanna VK, Ansari RW, Agrawal AK. Long-term functional restoration by neural progenitor cell transplantation in rat model of cognitive dysfunction: co-transplantation with olfactory ensheathing cells for neurotrophic factor support. *Int J Dev Neurosci* (2009) 27(1):103–10. doi: 10.1016/j.ijdevneu.2008.08.002

79. Qu Q, Sun G, Li W, Yang S, Ye P, Zhao C, et al. Orphan nuclear receptor TLX activates wnt/beta-catenin signalling to stimulate neural stem cell proliferation and self-renewal. *Nat Cell Biol* (2010) 12(1):31-40. doi: 10.1038/ncb2001

80. Mannino G, Russo C, Maugeri G, Musumeci G, Vicario N, Tibullo D, et al. Adult stem cell niches for tissue homeostasis. *J Cell Physiol* (2022) 237(1):239–57. doi: 10.1002/jcp.30562

81. Yue Y, Xue Q, Yang J, Li X, Mi Z, Zhao G, et al. Wnt-activated olfactory ensheathing cells stimulate neural stem cell proliferation and neuronal differentiation. *Brain Res* (2020) 1735:146726. doi: 10.1016/j.brainres.2020.146726

82. Peterson KM, Aly A, Lerman A, Lerman LO, Rodriguez-Porcel M. Improved survival of mesenchymal stromal cell after hypoxia preconditioning: role of oxidative stress. *Life Sci* (2011) 88(1-2):65–73. doi: 10.1016/j.lfs.2010.10.023

83. Stubbs SL, Hsiao ST, Peshavariya HM, Lim SY, Dusting GJ, Dilley RJ. Hypoxic preconditioning enhances survival of human adipose-derived stem cells and conditions endothelial cells *in vitro*. *Stem Cells Dev* (2012) 21(11):1887–96. doi: 10.1089/scd.2011.0289

84. Wang L, Jiang M, Duan D, Zhao Z, Ge L, Teng X, et al. Hyperthermiaconditioned OECs serum-free-conditioned medium induce NSC differentiation into neuron more efficiently by the upregulation of HIF-1 alpha and binding activity. *Transplantation* (2014) 97(12):1225–32. doi: 10.1097/TP.000000000000118

85. Zhuo Y, Wang L, Ge L, Li X, Duan D, Teng X, et al. Hypoxic culture promotes dopaminergic-neuronal differentiation of nasal olfactory mucosa mesenchymal stem cells *via* upregulation of hypoxia-inducible factor-1alpha. *Cell Transpl* (2017) 26(8):1452–61. doi: 10.1177/0963689717720291

86. Calabrese G, Giuffrida R, Lo Furno D, Parrinello NL, Forte S, Gulino R, et al. Potential effect of CD271 on human mesenchymal stromal cell proliferation and differentiation. *Int J Mol Sci* (2015) 16(7):15609–24. doi: 10.3390/ijms160715609

87. Lo Furno D, Mannino G, Cardile V, Parenti R, Giuffrida R. Potential therapeutic applications of adipose-derived mesenchymal stem cells. *Stem Cells Dev* (2016) 25(21):1615–28. doi: 10.1089/scd.2016.0135

88. Mannino G, Longo A, Gennuso F, Anfuso CD, Lupo G, Giurdanella G, et al. Effects of high glucose concentration on pericyte-like differentiated human adipose-derived mesenchymal stem cells. *Int J Mol Sci* (2021) 22(9):4604–19. doi: 10.3390/ijms22094604

89. Lo Furno D, Graziano AC, Caggia S, Perrotta RE, Tarico MS, Giuffrida R, et al. Decrease of apoptosis markers during adipogenic differentiation of mesenchymal stem cells from human adipose tissue. *Apoptosis* (2013) 18(5):578-88. doi: 10.1007/s10495-013-0830-x

90. Musumeci G, Mobasheri A, Trovato FM, Szychlinska MA, Graziano AC, Lo Furno D, et al. Biosynthesis of collagen I, II, RUNX2 and lubricin at different time points of chondrogenic differentiation in a 3D *in vitro* model of human mesenchymal stem cells derived from adipose tissue. *Acta Histochem* (2014) 116 (8):1407–17. doi: 10.1016/j.acthis.2014.09.008

91. Russo C, Mannino G, Patane M, Parrinello NL, Pellitteri R, Stanzani S, et al. Ghrelin peptide improves glial conditioned medium effects on neuronal differentiation of human adipose mesenchymal stem cells. *Histochem Cell Biol* (2021) 156(1):35–46. doi: 10.1007/s00418-021-01980-3

92. Lo Furno D, Pellitteri R, Graziano AC, Giuffrida R, Vancheri C, Gili E, et al. Differentiation of human adipose stem cells into neural phenotype by neuroblastoma- or olfactory ensheathing cells-conditioned medium. *J Cell Physiol* (2013) 228(11):2109–18. doi: 10.1002/jcp.24386

93. Lo Furno D, Mannino G, Giuffrida R, Gili E, Vancheri C, Tarico MS, et al. Neural differentiation of human adipose-derived mesenchymal stem cells induced by glial cell conditioned media. *J Cell Physiol* (2018) 233(10):7091–100. doi: 10.1002/jcp.26632

94. Lo Furno D, Mannino G, Pellitteri R, Zappala A, Parenti R, Gili E, et al. Conditioned media from glial cells promote a neural-like connexin expression in human adipose-derived mesenchymal stem cells. *Front Physiol* (2018) 9:1742. doi: 10.3389/fphys.2018.01742

95. Vicario N, Castrogiovanni P, Imbesi R, Giallongo S, Mannino G, Furno DL, et al. GJA1/CX43 high expression levels in the cervical spinal cord of ALS patients correlate to microglia-mediated neuroinflammatory profile. *Biomedicines* (2022) 10 (9):2246–64. doi: 10.3390/biomedicines10092246

96. Vicario N, Parenti R. Connexins signatures of the neurovascular unit and their physio-pathological functions. *Int J Mol Sci* (2022) 23(17):9510–26. doi: 10.3390/ijms23179510

97. Vicario N, Turnaturi R, Spitale FM, Torrisi F, Zappala A, Gulino R, et al. Intercellular communication and ion channels in neuropathic pain chronicization. *Inflammation Res* (2020) 69(9):841–50. doi: 10.1007/s00011-020-01363-9

98. Mannino G, Vicario N, Parenti R, Giuffrida R, Lo Furno D. Connexin expression decreases during adipogenic differentiation of human adipose-derived mesenchymal stem cells. *Mol Biol Rep* (2020) 47(12):9951–8. doi: 10.1007/s11033-020-05950-1

99. Wang B, Han J, Gao Y, Xiao Z, Chen B, Wang X, et al. The differentiation of rat adipose-derived stem cells into OEC-like cells on collagen scaffolds by co-culturing with OECs. *Neurosci Lett* (2007) 421(3):191-6. doi: 10.1016/j.neulet.2007.04.081

100. Garcia-Alias G, Lopez-Vales R, Fores J, Navarro X, Verdu E. Acute transplantation of olfactory ensheathing cells or schwann cells promotes recovery after spinal cord injury in the rat. *J Neurosci Res* (2004) 75(5):632–41. doi: 10.1002/jnr.20029

101. Lopez-Vales R, Fores J, Verdu E, Navarro X. Acute and delayed transplantation of olfactory ensheathing cells promote partial recovery after complete transection of the spinal cord. *Neurobiol Dis* (2006) 21(1):57–68. doi: 10.1016/j.nbd.2005.06.011

102. Lopez-Vales R, Garcia-Alias G, Fores J, Navarro X, Verdu E. Increased expression of cyclo-oxygenase 2 and vascular endothelial growth factor in lesioned spinal cord by transplanted olfactory ensheathing cells. *J Neurotrauma* (2004) 21 (8):1031–43. doi: 10.1089/0897715041651105

103. Ramon-Cueto A, Cordero MI, Santos-Benito FF, Avila J. Functional recovery of paraplegic rats and motor axon regeneration in their spinal cords by olfactory ensheathing glia. *Neuron* (2000) 25(2):425–35. doi: 10.1016/S0896-6273 (00)80905-8

104. Rossignol S, Schwab M, Schwartz M, Fehlings MG. Spinal cord injury: time to move? J Neurosci (2007) 27(44):11782–92. doi: 10.1523/JNEUROSCI.3444-07.2007

105. Andrews MR, Stelzner DJ. Evaluation of olfactory ensheathing and schwann cells after implantation into a dorsal injury of adult rat spinal cord. J Neurotrauma (2007) 24(11):1773–92. doi: 10.1089/neu.2007.0353

106. Ramer LM, Au E, Richter MW, Liu J, Tetzlaff W, Roskams AJ. Peripheral olfactory ensheathing cells reduce scar and cavity formation and promote regeneration after spinal cord injury. *J Comp Neurol* (2004) 473(1):1–15. doi: 10.1002/cne.20049

107. Gomez RM, Sanchez MY, Portela-Lomba M, Ghotme K, Barreto GE, Sierra J, et al. Cell therapy for spinal cord injury with olfactory ensheathing glia cells (OECs). *Glia* (2018) 66(7):1267–301. doi: 10.1002/glia.23282

108. Li Y, Li D, Raisman G. Interaction of olfactory ensheathing cells with astrocytes may be the key to repair of tract injuries in the spinal cord: the 'pathway hypothesis'. J Neurocytol (2005) 34(3-5):343–51. doi: 10.1007/s11068-005-8361-1

109. Franssen EH, Roet KC, de Bree FM, Verhaagen J. Olfactory ensheathing glia and schwann cells exhibit a distinct interaction behavior with meningeal cells. J Neurosci Res (2009) 87(7):1556–64. doi: 10.1002/jnr.21979

110. Pastrana E, Moreno-Flores MT, Gurzov EN, Avila J, Wandosell F, Diaz-Nido J. Genes associated with adult axon regeneration promoted by olfactory ensheathing cells: a new role for matrix metalloproteinase 2. *J Neurosci* (2006) 26 (20):5347–59. doi: 10.1523/JNEUROSCI.1111-06.2006

111. Chuah MI, Choi-Lundberg D, Weston S, Vincent AJ, Chung RS, Vickers JC, et al. Olfactory ensheathing cells promote collateral axonal branching in the injured adult rat spinal cord. *Exp Neurol* (2004) 185(1):15–25. doi: 10.1016/j.expneurol.2003.09.008

112. Imaizumi T, Lankford KL, Kocsis JD. Transplantation of olfactory ensheathing cells or schwann cells restores rapid and secure conduction across

the transected spinal cord. Brain Res (2000) 854(1-2):70-8. doi: 10.1016/S0006-8993(99)02285-4

113. Toft A, Scott DT, Barnett SC, Riddell JS. Electrophysiological evidence that olfactory cell transplants improve function after spinal cord injury. *Brain* (2007) 130(Pt 4):970–84. doi: 10.1093/brain/awm040

114. Xu X, Liang Z, Lin Y, Rao J, Lin F, Yang Z, et al. Comparing the efficacy and safety of cell transplantation for spinal cord injury: A systematic review and Bayesian network meta-analysis. *Front Cell Neurosci* (2022) 16:860131. doi: 10.3389/fncel.2022.860131

115. Kawaja MD, Boyd JG, Smithson LJ, Jahed A, Doucette R. Technical strategies to isolate olfactory ensheathing cells for intraspinal implantation. *J Neurotrauma* (2009) 26(2):155–77. doi: 10.1089/neu.2008.0709

116. Murtaza M, Mohanty L, Ekberg JAK, St John JA. Designing olfactory ensheathing cell transplantation therapies: Influence of cell microenvironment. *Cell Transpl* (2022) 31:9636897221125685. doi: 10.1177/09636897221125685

117. Miah M, Ferretti P, Choi D. Considering the cellular composition of olfactory ensheathing cell transplants for spinal cord injury repair: A review of the literature. *Front Cell Neurosci* (2021) 15:781489. doi: 10.3389/fncel.2021.781489

118. Wang X, Kuang N, Chen Y, Liu G, Wang N, Kong F, et al. Transplantation of olfactory ensheathing cells promotes the therapeutic effect of neural stem cells on spinal cord injury by inhibiting necrioptosis. *Aging (Albany NY)* (2021) 13 (6):9056–70. doi: 10.18632/aging.202758

119. He Y, Jiang Y, Dong L, Jiang C, Zhang L, Zhang G, et al. The aOECs facilitate the neuronal differentiation of neural stem cells in the inflammatory microenvironment through up-regulation of bioactive factors and activation of Wnt3/beta-catenin pathway. *Mol Neurobiol* (2022). doi: 10.1007/s12035-022-03113-w

120. Gu M, Gao Z, Li X, Guo L, Lu T, Li Y, et al. Conditioned medium of olfactory ensheathing cells promotes the functional recovery and axonal

regeneration after contusive spinal cord injury. Brain Res (2017) 1654(Pt A):43-54. doi: 10.1016/j.brainres.2016.10.023

121. Fan H, Chen Z, Tang HB, Shan LQ, Chen ZY, Wang XH, et al. Exosomes derived from olfactory ensheathing cells provided neuroprotection for spinal cord injury by switching the phenotype of macrophages/microglia. *Bioeng Transl Med* (2022) 7(2):e10287. doi: 10.1002/btm2.10287

122. Saglam A, Calof AL, Wray S. Novel factor in olfactory ensheathing cellastrocyte crosstalk: Anti-inflammatory protein alpha-crystallin b. *Glia* (2021) 69 (4):1022–36. doi: 10.1002/glia.23946

123. Kigerl KA, Gensel JC, Ankeny DP, Alexander JK, Donnelly DJ, Popovich PG. Identification of two distinct macrophage subsets with divergent effects causing either neurotoxicity or regeneration in the injured mouse spinal cord. *J Neurosci* (2009) 29(43):13435–44. doi: 10.1523/JNEUROSCI.3257-09.2009

124. Kroner A, Greenhalgh AD, Zarruk JG, Passos Dos Santos R, Gaestel M, David S. TNF and increased intracellular iron alter macrophage polarization to a detrimental M1 phenotype in the injured spinal cord. *Neuron* (2014) 83(5):1098–116. doi: 10.1016/j.neuron.2014.07.027

125. Basu S, Choudhury IN, Lee JYP, Chacko A, Ekberg JAK, St John JA. Macrophages treated with VEGF and PDGF exert paracrine effects on olfactory ensheathing cell function. *Cells* (2022) 11(15):2408–28. doi: 10.3390/cells11152408

126. Reshamwala R, Shah M, St John J, Ekberg J. Survival and integration of transplanted olfactory ensheathing cells are crucial for spinal cord injury repair: Insights from the last 10 years of animal model studies. *Cell Transpl* (2019) 28 (1\_suppl):132S-59S. doi: 10.1177/0963689719883823

127. Lindsay SL, Toft A, Griffin J, MME A, Barnett SC, Riddell JS. Human olfactory mesenchymal stromal cell transplants promote remyelination and earlier improvement in gait co-ordination after spinal cord injury. *Glia* (2017) 65(4):639–56. doi: 10.1002/glia.23117