RESEARCH ARTICLE



Design, synthesis, in vitro evaluation, and molecular modeling studies of *N*-substituted benzomorphans, analogs of LP2, as novel MOR ligands

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Abstract

6,7-Benzomorphans have been investigated in medicinal chemistry for developing new drugs. This nucleus could be considered a versatile scaffold. The physicochemical properties of benzomorphan *N*-substituent are crucial in achieving a definite pharmacological profile at opioid receptors. Thus, the dual-target MOR/ DOR ligands LP1 and LP2 were obtained through *N*-substituent modifications. Specifically, LP2, bearing as *N*-substituent the (2R/S)-2-methoxy-2- phenylethyl group, is a dual-target MOR/DOR agonist and is successful in animal models of inflammatory and neuropathic pain. To obtain new opioid ligands, we focused on the design and synthesis of LP2 analogs. First, the 2-methoxyl group of LP2 was replaced by an ester or acid functional group. Then, spacers of different lengths were introduced at *N*-substituent. In-vitro, their affinity profile versus opioid receptors has been performed through competition binding assays. Molecular modeling studies were conducted to deeply analyze the binding mode and the interactions between the new ligands and all opioid receptors.

K E Y W O R D S

6,7-benzomorphan, docking studies, opioid receptor, radioligand competition-binding

1 | INTRODUCTION

6,7-Benzomorphans, originated from morphine structure simplification, have been investigated in medicinal chemistry for developing new drugs (Turnaturi et al., 2018). In the interaction of opioid receptors with benzomorphan-based compounds, three different pharmacophoric requirements were identified: the aromatic ring, the saturated segment, and the basic nitrogen. In fact, the basic nitrogen and phenolic group of Tyr1 of endogenous opioid peptides could be mimic by the rigid benzomorphan scaffold. The (-)-(2R,6R,11R) configuration of the benzomorphan nucleus is preferred for opioid receptors interaction and is identical to the (-)-morphine configuration (Pathan & Williams, 2012). Moreover, this nucleus could be considered a versatile scaffold, and the

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modifications of the functional groups attached to basic nitrogen led to different opioid ligands. Thus, the physicochemical properties of benzomorphan *N*-substituent are crucial in achieving a definite pharmacological profile at MOR (mu opioid receptor), DOR (delta opioid receptor), and KOR (kappa opioid receptor).

To obtain different functional profiles, Pasquinucci et al. (Pasquinucci et al., 2021) designed and synthesized different benzomorphan-based compounds. The dualtarget MOR/DOR ligands LP1 and LP2 were discovered (Figure 1). LP1, bearing an *N*-phenylpropanamide substituent (Pasquinucci et al., 2010), is a potent MOR agonist/ DOR antagonist able to counteract nociceptive pain and behavioral signs of persistent pain with low toleranceinducing capability (Parenti et al., 2013; Pasquinucci et al., 2012). Moreover, LP2, with a flexible 2-methoxyethyl spacer at N-substituent, is a dual-target MOR/DOR agonist (Pasquinucci et al., 2017) that exhibited antinociceptive effect in nociceptive and persistent pain models (Pasquinucci et al., 2019; Vicario et al., 2019). Thus, LP1 and LP2 could represent helpful examples of the discovery process of new opioid ligands with a safer profile through N-substituent modification of the benzomorphan scaffold.

To improve the understanding of molecular interaction between the *N*-substituent and the opioid receptors, here we report the design and synthesis of **LP2** analogs. First, the structure-affinity profile of new ligands was evaluated, and the 2-methoxyl group of **LP2** was replaced by an ester or acid functional group. Then, spacers of different lengths were introduced at *N*-substituent (Figure 1). In all new **LP2** analogs the phenyl ring in the *N*-substituent of the benzomorphan scaffold was retained. In-vitro, their affinity profile versus opioid receptors has been performed through competition binding assays. Molecular modeling was conducted to deeply analyze the binding mode and the interactions between the new ligands and MOR, DOR, and KOR.

2 | RESULTS AND DISCUSSION

2.1 | Chemistry

The new compounds **2**, **3**, and **10–13** were synthesized according to Schemes 1 and 2, respectively. Compound **1** was prepared by primary alcohol sulfonylation with methanesulfonyl chloride and triethylamine in CH_2Cl_2 . Target ester **2** was obtained by alkylation of (–)-*cis*-*N*-normetazocine (Brine et al., 1990) with the mesylated alcohol **1**, and the respective acid derivate **3** was obtained by the ester hydrolysis in basic condition.

Intermediate compounds **8** and **9** were synthesized as previously reported (Cheng et al., 1990). Target compounds **10** and **11** were synthesized by alkylation of (-)-*cis*-*N*-normetazocine with intermediate compounds **8** and **9**. The respective acids **12** and **13** were obtained by esters hydrolysis in basic condition. New synthesized compounds possess a chiral stereocenter at the *N*-substituent of (-)-*cis*-*N*-normetazocine scaffold and are a mixture of diastereoisomers, not separable in our purification procedures. They were characterized by ¹H NMR, ¹³C NMR, and elemental analysis.

2.2 | Radioligand binding assays

Target compounds **2**, **3**, and **10–13** were assayed on opioid receptors, as previously described (Rita Turnaturi et al., 2022). Inhibition constant (K_i) values (Table 1) were calculated using nonlinear regression analysis (GraphPad Prism, version 6.0, GraphPad Software In.). (Figures S1–S3).

Compared to LP2, K_i values versus MOR, DOR, and KOR are higher for all newly synthesized compounds. However, regarding their affinity to opioid receptors, the new molecules behaved differently based on Nsubstituents. LP2 has the phenyl ring in the N-substituent of the benzomorphan scaffold linked to an ethyl spacer bearing a methoxyl group at carbon 2. In compound 2, the 2-methoxyl group of LP2 was replaced by an ethyl ester functional group. This substitution is well tolerated, and compound **2** confirmed a MOR profile with the K_i values of 38.2 nM. Instead, a reduction of DOR and KOR affinity in comparison to LP2 was observed. Compounds 10 and 11 also retained an ethyl ester functionality in Nsubstituent, but the distance with respect to basic nitrogen of the benzomorphan nucleus was incremented by 1 and 2 methylene units, respectively. The nanomolar MOR affinity, exhibited by compounds **10** and **11** ($K_i = 10$ nM

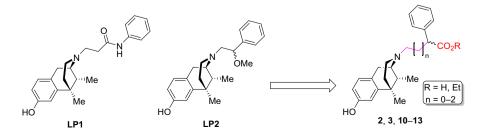
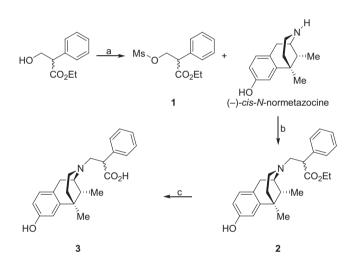


FIGURE 1 Chemical structures of **LP1**, **LP2** and the newly synthesized compounds.

and $K_i = 53.7$ nM, respectively), confirmed that in MOR interaction, the ethyl ester in *N*-substituent was tolerated highlighting an optimal two methylene units spacer. In compounds **10** and **11**, the affinity at DOR (14- and 50-fold, respectively) and KOR (2- and 6-fold) was higher with respect to compound **2**, with a significative incremental of KOR affinity in compound **11** ($K_i = 32.1 \pm 4$), showing a selectivity ratio of $K_i^{\text{DOR}}/K_i^{\text{MOR}} = 2.31$ and $K_i^{\text{KOR}}/K_i^{\text{MOR}} = 0.6$.

An acid functional group characterizes the compounds **3**, **12**, and **13**. Compound **12** showed a nanomolar K_i value at MOR comparable to its ester analog **10**. Instead, the acid functional group in compounds **3** and **13** led to a detrimental effect on all opioid receptors compared to compounds **2** and **11**. Thus, replacing the ester group with the corresponding acid group seems not tolerated depending on chain lengths.



SCHEME 1 Synthesis of target compounds **2** and **3**. Reagents and conditions: (a) MsCl, TEA, anhydrous DCM, rt, 3 h; (b) NaHCO₃, KI, DMF, 55 °C, 24 h; (c) NaOH 1 N, 110 °C, 5 h.

2.3 | Molecular modeling

2.3.1 | Docking studies

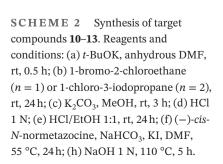
In silico studies were performed using AutoDock implemented in the YASARA software. The docking studies were performed on all the protonation states of the compounds at pH 7.4, previously calculated using the Marvin software (Rescifina et al., 2014; Szczepańska et al., 2021; Varrica et al., 2018).

All compounds at pH 7.4 possess positively charged nitrogen, forming a nitrogen stereocenter for each molecule in addition to the one already present. To avoid docking studies on four stereoisomers for each compound, we first docked the four stereoisomers of compound **LP2**, using it as a reference (Figure 2). Then, regarding the nitrogen stereocenter, the chosen stereoisomer had the calculated K_i value closest to the experimental one, in our case (R)-N. Based on this assumption, we kept the stereocenter on nitrogen fixed (R) for all other compounds and performed docking studies on the two isomers of each compound for the three receptors.

In Table 2, we have only listed the ΔG and K_i values of the isomers whose calculated K_i value is closest to the experimental one; in almost all compounds, however, there is not much difference in inhibitory activity between the two isomers.

Focusing on the MOR, we carefully analyzed the poses within the receptor site of compounds **10** and **12**, which possess a ΔG (kcal/moL) of -9.91 and -9.67, respectively.

In Figure 3, we show the 3D and 2D poses of compounds **10**, **12**, and LP2 within the MOR, where we can see that the presence of the two groups, ester, and carboxylic acid, alters the pose within the site but still retains excellent inhibitory activity. Compound **10** (Figure 3a), in addition to hydrophobic interactions with residues Ile296 and 322, in common with compound **12**, possesses a salt



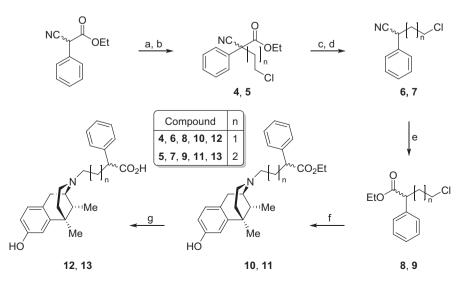


 TABLE 1
 Opioid receptors binding affinity and selectivity of compounds 2, 3, and 10–13.

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Compound	N-substituent	$K_{i}(nM) \pm SD^{a}$					
		MOR	DOR	KOR	$K_{\rm i}^{\rm DOR}/K_{\rm i}^{\rm MOR}$	$K_{\rm i}^{\rm KOR}/K_{\rm i}^{\rm MOR}$	
2	200 NG	38.2 ±4	6,170 ± 70	211.0 ±10	161.5	5.5	
3	HO PO	980.6 ± 8	2,769.0 ± 50	715.0 ±4	2.8	0.7	
10	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	10.8 ± 2	440.0 ± 22	130.0 ±15	40.7	12.0	
11	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	53.7 ±7	124.0 ± 8	32.1 ±4	2.3	0.6	
12	HOLO	11.8 ±2	275.0 ± 11	3,216.0 ± 60	23.3	252.7	
13	HOLO	477.0 ± 27	852.0 ± 10	2,716.0 ± 20	1.8	4.6	
2 <i>R</i> / <i>S</i> - LP2 ^b		1.1 ± 0.1	6.6 ± 0.6	15.2 ± 0.8	6.1	14.0	
DAMGO		1.2 ± 0.1	_	_	_	_	
Naltrindole		_	1.1 ± 0.1	—	—	-	
U69,593		_	—	0.3 ± 0.1	—		

^aEach value is the mean ± SD of at least two experiments performed in duplicate. Reference compounds were tested with the same membrane homogenates. ^bReference (Pasquinucci et al., 2017).

bridge with residue Asp147, a typical interaction for the MOR; it also establishes hydrogen bonds with residues Gln124 and Asn127. The presence of the carboxyl group in compound **12**, on the other hand, allows the formation of a hydrogen bond with His297 and prevents the formation of the salt bridge with Asp147, with which electrostatic interactions are formed (Ricarte et al., 2021; Zádor et al., 2022) (Figure 3b).

Another aspect being emphasized is the different affinity that **10** and **12** have compared to the other ligands studied; this is to be found in the spacer chain. The presence of an alkyl chain of two carbon atoms allows ligands **10** and **12** to accommodate themselves within the receptor site in such a way as to maximize interactions and strengthen anchoring.

2.3.2 | Molecular dynamics

To study better the formation of the ligand@protein complex, 100 ns molecular dynamics of ligands **10** and **12** were performed within the receptor pocket of the MOR protein. From the graphs of the total energy and RMSD (rootmean-square deviation) of the complexes (Figure 4), it is clear that both ligands form a very stable complex with the MOR; this is due to the dense network of interactions, previously commented, that keep the ligands anchored within the receptor pocket (Gentile et al., 2022).

3 | CONCLUSION

Novel **LP2** analogs with a benzomorphan scaffold were designed and synthesized. Based on in vitro competition binding assays and molecular modeling studies, compounds **10** and **12** showed a high affinity for MOR. Both molecules are appropriately located in the binding pocket of MOR, and maintain a similar docking pose. An alkyl chain of two methylene units maximizes their interactions with MOR, highlighting the critical role of the distance between the basic nitrogen of the benzomorphan scaffold

and the ester or acid functional groups. Thus, the combination of different functional groups and the spacing carbon chain length appeared to determine the affinity profile of compounds. In this study, we confirmed that *N*-substituent nature modulates the nature of interactions towards the opioid receptors.

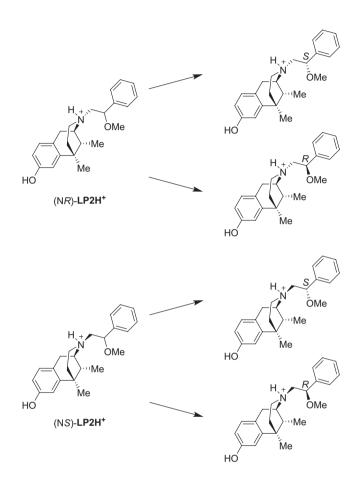


FIGURE 2 Protonation states of **LP2** and relative stereoisomers.

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4 | EXPERIMENTAL SECTION

4.1 | General remarks

Reagent-grade chemicals were purchased from Merck (Darmstadt, Germany) and (\pm) -*cis-N*-normetazocine was obtained from Fabbrica Italiana Sintetici. All new compounds were purified by flash column chromatography on Merck silica gel 60 (230–400 mesh). Melting points were determined in open capillary tubes with a Büchi 530 apparatus and are uncorrected. Optical rotations were determined in EtOH solution with a Perkin-Elmer 241 polarimeter. ¹H and ¹³C NMR spectra were recorded at 200 and 500 MHz on Varian Inova spectrometers in CDCl₃, CD₃OD or DMSO-*d*₆. Elemental analyses (C, H, N) were performed on a Carlo Erba 1106 analyzer, and the results were within ±0.4% of the theoretical values.

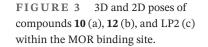
4.2 | Preparation of the target compounds 2 and 3

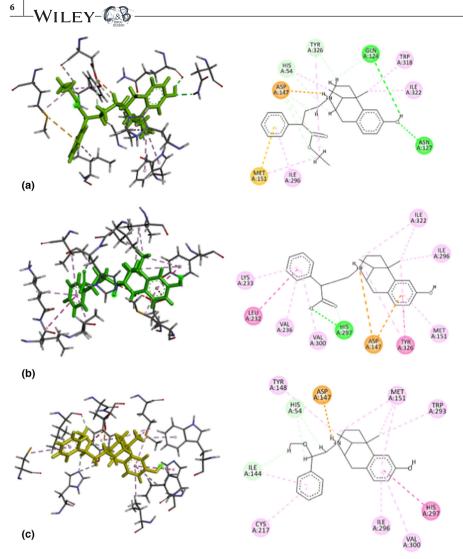
4.2.1 | Ethyl 3-((methylsulfonyl)oxy)-2-(R/S)-phenylpropanoate (**1**)

Ethyl tropate (2.77 mmoL, 1 eq) was dissolved in CH₂Cl₂ (6 mL), the solution was cooled at 0 °C under a nitrogen atmosphere, and triethylamine (5.54 mmoL, 2 eq) and methanesulfonyl chloride (3.33 mmoL, 1.2 eq) were added. The reaction mixture was stirred overnight at rt. After, the mixture was transferred to a separatory funnel and partitioned (CH₂Cl₂/H₂O). The organic phase was washed with brine, dried over Na₂SO₄, and concentrated in vacuo to give an amber oil that, purified by flash chromatography (EtOAc/C₆H₁₂ 1:9), gave a light-yellow oil. Yield: 42%; TLC EtOAc/C₆H₁₂ (2:8 ν/ν) $R_f = 0.50$; ¹H NMR

TABLE 2 Calculated free energies of binding, ΔG (kcal/moL), and constants of binding, K_i (nM), for the binding sites of MOR, DOR, and KOR for all compounds.

	MOR			DOR			KOR		
Compound	Calcd ΔG	Calcd K _i	Expected K _i	Calcd ΔG	Calcd K _i	Expected K _i	Calcd ΔG	Calcd K _i	Expected K _i
2	-9.86	58.7	38.2 ± 4	-6.97	7733.4	6170.0 ± 70	-9.16	191.5	211.0 ± 10
3	-8.04	1269.6	980.6 ± 8	-7.58	2760.7	2769.0 ± 50	-8.34	765.0	715 ± 4
10	-9.91	54.0	10.8 ± 2	-8.59	501.5	$440\pm\!22$	-9.32	146.2	130 ± 15
11	-9.78	67.2	53.7 ± 7	-8.78	363.9	124 ± 8	-9.89	55.8	32.1 ± 4
12	-9.67	81.0	11.8 ± 2	-8.96	268.5	275 ± 11	-7.57	2807.7	3216 ± 60
13	-8.69	423.6	477 ± 27	-8.20	969.0	852 ± 10	-7.74	2107.0	2716 ± 20
2R/S-LP2	-9.64	85.2	1.08 ± 0.10	-9.86	58.7	6.61 ± 0.60	-9.13	212.0	15.22 ± 0.80
DAMGO	-11.83	2.1	1.16 ± 0.10			-			-
Naltrindole			-	-9.68	79.6	1.13 ± 0.10			-
U69,593			-			-	-9.91	54.0	0.34 ± 0.10





(200 MHz, CDCl₃): δ = 7.34–7.21 (m, 5H), 4.56–4.47 (m, 1H), 4.21–4.06 (m, 1H), 3.73–3.68 (m, 3H), 2.97 (s, 3H), 1.21 (t, 3H).

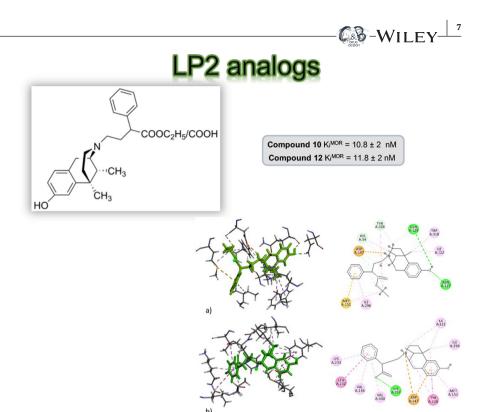
4.2.2 | Ethyl 3-((2R,6R,11R)-8-hydroxy-6,11-dimethyl-1,4,5,6-tetrahydro-2,6methanobenzo[d]azocin-3(2H)-yl)-2-(R/S)phenylpropanoate (**2**)

(-)-*cis*-*N*-normetazocine (0.92 mmol, 1 eq) was dissolved in DMF (5 mL) and ethyl 3-((methylsulfonyl) oxy)-2-phenylpropanoate (0.92 mmol, 1 eq), NaHCO₃ (1.38 mmoL, 1.5 eq) and KI (catalytic quantity) were added. The reaction mixture was stirred overnight at 55 °C. At the reaction mixture, 4 mL of H₂O were added, and the aqueous phase was extracted with EtOAc; the organic phase was dried over Na₂SO₄ and concentrated in vacuo. The reaction crude was purified by flash chromatography (CH₂Cl₂/MeOH 95:5) to obtain a white solid. Yield: 96%; Mp: 178–181 °C; $[\alpha]_D^{25} = -61.7^\circ$ (*c* 1.005, EtOH); TLC CH₂Cl₂/MeOH (95:5 *v*/*v*) $R_f = 0.49$; ¹H NMR (500 MHz, CDCl₃): $\delta = 7.40-7.25$ (m, 5H), 6.87 (d, 1H,

$$\begin{split} J &= 8.3 \text{ Hz}), 6.70 \text{ (d, 1H, } J &= 3.4 \text{ Hz}), 6.60 \text{ (dd, 1H, } J &= 8.3, \\ 3.4 \text{ Hz}), 5.90 \text{ (s, 1H)}, 4.25 \text{ (d, 2H, } J &= 7.2 \text{ Hz}), 3.28 \text{ (d, 1H, } \\ J &= 7.1 \text{ Hz}), 2.99-2.79 \text{ (m, 4H)}, 2.70-2.59 \text{ (m, 4H)}, 1.84- \\ 1.70 \text{ (m, 2H)}, 1.29 \text{ (s, 3H)}, 1.27 \text{ (t, 3H, } J &= 7.2 \text{ Hz}), 0.80 \\ \text{ (d, 3H, } J &= 7.0). \text{ (Figure S4)}^{13} \text{C NMR} (125 \text{ MHz, CDCl}_3): \\ \delta &= 173.95, 154.46, 137.41, 18.98, 128.51, 128.26, 128.12, \\ 128.09, 128.00, 127.94, 127.33, 126.88, 113.89, 112.83, \\ 112.43, 112.23, 66.12, 57.18, 51.80, 51.31, 46.24, 45.75, \\ 42.20, 41.96, 41.46, 36.40, 25.55, 24.79, 14.17, 13.55. \text{ Anal} \\ \text{ (C}_{25}\text{H}_{31}\text{NO}_3) \text{ C, H, N} \text{ (Table S1).} \end{split}$$

4.2.3 | 3-[(2R,6R,11R)-8-hydroxy-6,11-dimethyl-1,4,5,6-tetrahydro-2,6methanobenzo[*d*]azocin-3(2H)-yl]-2(*R*/*S*)phenylpropanoic acid (**3**).

1 N NaOH solution (3.06 mmol, 9.27 eq) was added to ethyl 3-((2*R*,6*R*,11*R*)-8-hydroxy-6,11-dimethyl-1,4,5,6-tetrahydro-2,6-methanobenzo[*d*]azocin-3(2*H*)yl)-2-phenylpropanoate (0.33 mmoL, 1 eq). The resulting suspension was vigorously stirred and refluxed at 110 °C for 5 h. After cooling, the reaction mixture was partitioned (CHCl₃/H₂O). A 1 N solution **FIGURE 4** Total energy and RMSD of the ligand@protein complexes for compounds **10** (upper) and **12** (down) within the MOR.



of HCl was added to the aqueous phase to a pH of 5–6. The obtained yellow precipitate was crystallized by EtOH/diethyl ether. Yield: 90%; Mp: 180–183 °C; $[\alpha]_D^{25} = -57.2^{\circ}$ (*c* 1.03, EtOH); TLC CH₂Cl₂/MeOH (95:5 ν/ν): $R_f = 0.30$; ¹H NMR (500 MHz, DMSO- d_6): $\delta = 9.12$ (br, 1H), 7.31–7.28 (m, 5H), 6.87–6.84 (m, 1H), 6.60–6.59 (m, 1H), 6.55–6.53 (m, 1H), 4.11–3.98 (m, 1H), 3.30–3.19 (m, 4H), 3.01–2.74 (m, 3H), 2.35–2.24 (m, 1H), 1.99–1.76 (m, 2H), 1.23 (s, 3H), 0.72 (d, 3H). (Figure S5) ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 173.44$, 155.95, 140.66, 137.81, 128.54, 128.48, 128.06, 128.02, 127.94, 127.88, 127.22, 126.04, 113.50, 111.83, 92.52, 56.26, 51.00, 46.74, 45.08, 35.17, 26.58, 25.39, 12.93. Anal (C₂₃H₂₇NO₃) C, H, N (Table S1).

4.3 | **Preparation of the target** compounds 10–13

4.3.1 | Ethyl 4-chloro-2-(R/S)-cyano-2-phenylbutanoate (4)

To a suspension of potassium *t*-butoxide (34 mmoL, 2 eq) in anhydrous DMF (18 mL), ethyl phenyl cyanoacetate (17 mmoL, 1 eq) was added dropwise, under argon. After 30 min, 1-bromo-2-chloroethane (25.5 mmoL, 1.5 eq) was added dropwise. The reaction mixture was stirred at rt for 24 h. Then, the DMF is removed under vacuum. To the reaction mixture, an aqueous solution of NH₄Cl was added. The mixture was transferred to a separatory funnel and partitioned with diethyl ether. The crude obtained was purified by flash chromatography on silica gel (C₆H₁₂/

EtOAc, 9:1–8:2) to give a yellow oil. Yield: 63.25%. TLC $C_6H_{12}/EtOAc$ (8:2 ν/ν): $R_f = 0.33$; ¹H NMR (200 MHz, CDCl₃): $\delta = 7.56-7.40$ (m, 5H), 4.28 (q, 1H, J = 7.2 Hz), 4.21 (q, 1H, J = 7.2 Hz), 3.71–3.45 (m, 2H), 2.96–2.81 (m, 1H), 2.68–2.53 (m, 1H), 1.25 (t, 3H, J = 7.2 Hz).

4.3.2 | Ethyl 5-chloro-2-(R/S)-cyano-2-phenylpentanoate (5)

To a suspension of potassium *t*-butoxide (42.28 mmoL, 2 eq) in anhydrous DMF (18 mL), ethyl phenyl cyanoacetate (21.14 mmoL, 1 eq) was added dropwise, under argon. After 30 min, 1-chloro-3-iodopropane (41.10 mmoL, 1.5 eq) was added dropwise. The reaction mixture was stirred at rt for 24h. Then, the DMF is removed under vacuum. To the reaction mixture, an aqueous solution of NH₄Cl was added. The mixture was transferred to a separatory funnel and partitioned with diethyl ether. The crude obtained was purified by flash chromatography on silica gel (C₆H₁₂/EtOAc, 9:1–8:2) to give a yellow oil. Yield: 54%; TLC C₆H₁₂/EtOAc (8:2 *v/v*): $R_{\rm f} = 0.45$; ¹H NMR (200 MHz, CDCl₃): $\delta = 7.57-7.40$ (m, 5H), 4.27 (q, 1H, J = 7.2 Hz), 4.21 (q, 1H, J = 7.2 Hz), 3.57 (t, 2H), 2.57–2.27 (m, 2H), 2.05–1.80 (m, 2H), 1.26 (t, 3H, J = 7.2 Hz).

4.3.3 | 4-Chloro-2-(*R/S*)-phenylbutanenitrile(6)

Ethyl 4-chloro-2-(R/S)-cyano-2-phenylbutanoate (4.64 mmoL, 1 eq) was dissolved in CH₃OH (20 mL), and a

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a transparent oil. Yield: 52%; TLC C₆H₁₂/EtOAc (8:2 ν/ν): $R_{\rm f} = 0.54$; ¹H NMR (200 MHz, CDCl₃): $\delta = 7.38-7.27$ (m, 5H), 4.18 (q, 1H, J = 7.4 Hz), 4.05 (q, 1H, J = 7.4 Hz), 3.58– 3.51 (m, 3H), 2.30-2.19 (m, 1H), 2.00-1.93 (m, 1H), 1.79-1.68 (m, 2H), 1.22 (t, 3H, J = 7.4 Hz).4.3.7 Ethyl 4-((2*R*,6*R*,11*R*)-8-hydroxy-6,11-dimethyl-1,4,5,6-tetrahydro-2,6methanobenzo[d]azocin-3(2H)-yl)-2-(R/S)phenylbutanoate (10) (-)-cis-N-normetazocine (2.17 mmoL, 1 eq) was dissolved inDMF(6mL), and ethyl4-chloro-2(R/S)-phenylbutanoate (2.17 mmoL, 1 eq), NaHCO₃ (3.25 mmoL, 1.5 eq) and KI (catalytic quantity) were added. The reaction mixture was stirred at 65 °C for 72 h. After, the mixture was transferred to a separatory funnel and partitioned (EtOAc/ H₂O). The organic phase was washed with brine, dried on Na₂SO₄, and concentrated under vacuum. The reaction

crude was purified by flash chromatography (CH₂Cl₂/ MeOH 95:5) to obtain an orange solid. Yield: 30%; Mp: 157–160 °C; $[\alpha]_{\rm D}^{25} = -56.6^{\circ}$ (c 1.1, EtOH); TLC CH₂Cl₂/ MeOH (95:5 v/v) $R_f = 0.35$; ¹H NMR (200 MHz, CDCl₃): $\delta = 7.30-7.19$ (m, 5H), 6.82 (d, 1H J = 8.0 Hz), 6.63 (d, 1HJ = 2.2 Hz), 6.53 (dd, 1H, J = 2.2, 8.0 Hz), 4.03 (q, 2H, J = 7.2 Hz), 3.56 (t, 1H, J = 7.1 Hz), 2.83–2.22 (m, 7H), 2.11-1.72 (m, 4H), 1.61-1.46 (m, 1H), 1.23 (s, 3H) 1.12 (t, 3H, *J* = 7.2 Hz), 0.75 (d, 3H, *J* = 7.0 Hz). (Figure S6) ¹³C NMR (50 MHz, CDCl₃): δ = 173.87, 154.56, 142.79, 138.85, 129.01, 128.59, 128.14, 128.02, 127.85, 127.20, 113.24, 112.40, 60.84, 57.57, 52.54, 49.78, 45.87, 41.85, 41.49, 41.38, 40.85, 36.19, 33.42, 25.23, 23.30, 14.05. Anal

4.3.8 Ethyl 5-((2R,6R,11R)-8-hydroxy-6,11dimethyl-1,4,5,6-tetrahydro-2,6-methanobenzo[d] azocin-3(2H)-yl)-2-(R/S)-phenylpentanoate (11)

(C₂₆H₃₃NO₃) C, H, N (Table S1).

(-)-cis-N-normetazocine (4.15 mmoL, 1 eq) was dissolved in DMF (6 mL) and ethyl 5-chloro-2(R/S)-phenylpentanoa te (4.15 mmoL, 1 eq), NaHCO₃ (6.23 mmoL, 1.5 eq) and KI (catalytic quantity) were added. The reaction mixture was stirred at 65 °C for 72 h. After, the mixture was transferred to a separatory funnel and partitioned (EtOAc/H₂O). The organic phase was washed with brine, dried on Na₂SO₄, and concentrated in vacuum. The reaction crude was purified by flash chromatography (CH₂Cl₂/MeOH 95:5) to obtain an orange solid. Yield: 46%; Mp: 161–164 °C; $[\alpha]_{D}^{25} = -54.2^{\circ}$ (*c* 1.05, EtOH); TLC CH₂Cl₂/MeOH (95:5 ν/ν) $R_{\rm f} = 0.48$; ¹H NMR (500 MHz, CDCl₃): δ = 7.56–7.54 (m, 2H), 7.41–7.36 (m, 3H), 6.91 (d, 1H, J = 10.0 Hz), 6.69 (d, 1H, J = 5.0 Hz),

saturated aqueous solution of K₂CO₃ (4 mL) was added at 0 °C. The reaction mixture was stirred at rt for 3 h, and HCl was added to pH 7. The mixture was partitioned with diethyl ether, the organic phase was washed with brine, dried over Na₂SO₄, and concentrated in vacuo to obtain a light-yellow oil. Yield: 75%; TLC C_6H_{12} /EtOAc (8:2 ν/ν): $R_{\rm f} = 0.36$; ¹H NMR (200 MHz, CDCl₃): $\delta = 7.38-7.27$ (m, 5H), 4.14 (t, 1H), 3.72-3.48 (m, 1H), 2.45-2.22 (m, 1H), 1.73-1.40 (m, 2H).

4.3.4 5-Chloro-2-(R/S)-pentanenitrile (7)

Ethyl 5-chloro-2-(R/S)-cyano-2-henylpentanoate (11.51 mmoL, 1 eq) was dissolved in CH₃OH (40 mL), and a saturated aqueous solution of K₂CO₃ (8 mL) was added at 0 °C. The reaction mixture was stirred at rt for 3 h, and HCl was added to pH 7. The mixture was transferred to a separatory funnel and partitioned with diethyl ether. The organic phase was washed with brine, dried over Na₂SO₄, and concentrated in vacuo to obtain a light-yellow oil. Yield: 61%; TLC C_6H_{12} /EtOAc (8:2 v/v): $R_f = 0.40$; ¹H NMR $(200 \text{ MHz}, \text{CDCl}_3)$: $\delta = 7.40-7.33 \text{ (m, 5H)}, 3.82 \text{ (t, 1H)}, 3.57$ (t, 2H), 2.15-1.94 (m, 4H).

4.3.5 Ethyl 4-chloro-2-(R/S)phenylbutanoate (8)

4-Chloro-2(R/S)-phenylbutanonitrile (3.46 mmoL, 1 eq) was added to a 1:1 mixture of HCl (12 N) and CH₃CH₂OH, and the resulting suspension was stirred and refluxed for 24 h. After, the reaction mixture was concentrated under vacuum, transferred to a separatory funnel, and partitioned with CH₂Cl₂. The organic phase was washed with brine, dried on Na2SO4, and concentrated to obtain a colorless oil. Yield: 63%; TLC C₆H₁₂/EtOAc (8:2 ν/ν): $R_f = 0.48$; ¹H NMR (200 MHz, CDCl₃): δ = 7.39–7.27 (m, 5H), 4.15 (q, 1H, J = 7.4 Hz), 4.09 (q, 1H, J = 7.4 Hz), 3.87 (t, 1H),3.57-3.43 (m, 1H), 2.59-2.42 (m, 1H), 2.35-2.17 (m, 1H), 1.77–1.74 (m, 1H), 1.20 (t, 3H, *J* = 7.4 Hz).

4.3.6 Ethyl 5-chloro-2-(R/S)phenylpentanoate (9)

5-Chloro-2-(R/S)-pentanenitrile (7.94 mmoL, 1 eq) was added to a 1:1 mixture of HCl (12N) and CH₃CH₂OH, and the resulting suspension was stirred and refluxed for 24 h. Afterward, the reaction mixture was concentrated under vacuum, transferred to a separatory funnel, and partitioned with CH₂Cl₂. The organic phase was washed with brine, dried on Na2SO4, and concentrated to obtain

6.59 (dd, 1H, J = 5.0, 10.0 Hz), 4.13–4.12 (m, 1H), 2.64– 2.40 (m, 5H), 2.02–1.98 (m, 2H), 1.85–1.70 (m, 4H), 1.64– 1.56 (m, 4H), 1.32 (s, 3H), 1.27–1.24 (m, 3H), 0.81 (d, 3H). (Figure S7) ¹³C NMR (125 MHz, CDCl₃): $\delta = 167.60$, 167.47, 155.91, 133.70, 133.30, 129.49, 129.44, 129.35, 129.33, 128.53, 126.00, 118.02, 114.43, 112.67, 85.11, 83.62, 63.56, 54.29, 54.25, 53.61, 53.47, 35.48, 35.09, 34.79, 29.73, 24.27, 13.74. Anal ($C_{27}H_{35}NO_3$) C, H, N (Table S1).

4.3.9 | 4–((2R,6R,11R)-8-hydroxy-6,11-dimethyl-1,4,5,6-tetrahydro-2,6methanobenzo[d]azocin-3(2H)-yl)-2-(R/S)phenylbutanoic acid (**12**)

1N NaOH solution (3.06 mmoL, 11.3 eq) was added to ethyl 4-((2R,6R,11R)-8-hydroxy-6,11-dimethyl-1,4,5,6-te trahydro-2,6-methanobenzo[d]azocin-3(2H)-yl)-2-(R/S)phenylbutanoate (0.27 mmoL, 1 eq). The resulting suspension was vigorously stirred and refluxed at 110 °C for 5 h. After, the mixture was cooled at rt, transferred to a separatory funnel, and partitioned (CHCl₃/H₂O). A 1 N solution of HCl was added to the aqueous phase until a pH range of 5-6. The obtained yellow precipitate was separated from the aqueous phase by vacuum filtration. Yield: 43%, Mp: 192 °C dec; $[\alpha]_D^{25} = -42.6^\circ$ (*c* 1.02, EtOH); TLC CH₂Cl₂/ MeOH (95:5 v/v) $R_f = 0.29$; ¹H NMR (500 MHz, DMSO- d_6): $\delta = 8.49$ (brs, 1H), 7.28–7.26 (m, 2H), 7.19–7.16 (m, 2H), 7.09–7.07 (m, 1H), 6.99 (d, 1H, J = 10.0 Hz), 6.81–6.79 (d, 1H, J = 10 Hz), 6.49–6.47 (dd, 1H, J = 10 Hz), 3.19–3.06 (m, 2H), 2.74-2.68 (m, 2H), 2.44-2.28 (m, 4H), 1.86-1.83 (m, 2H), 1.70-1.58 (m, 3H), 1.17-1.14 (m, 2H), 0.84-0.83 (m, 1H), 0.68 (d, 3H, J = 7.2 Hz). (Figure S8) ¹³C NMR $(125 \text{ MHz}, \text{DMSO-}d_6): \delta = 177.13, 172.88, 156.00, 144.82,$ 142.37, 128.08, 127.84, 127.70, 126.79, 125.62, 113.32, 112.12, 86.24, 67.55, 56.63, 52.94, 45.00, 41.97, 36.41, 33.00, 29.08, 25.47, 22.62, 13.64. Anal (C₂₄H₂₉NO₃) C, H, N (Table S1).

4.3.10 | 5-((2R,6R,11R)-8-hydroxy-6,11-dimethyl-1,4,5,6-tetrahydro-2,6-methanobenzo[d]azocin-3(2H)-yl)-2-(R/S)-phenylpentanoic acid (13)

1 N NaOH solution (10.53 mmoL, 11.3 eq) was added to ethyl 5-((2*R*, 6*R*, 11*R*)-8-hydroxy-6,11-dimethyl-1,4,5,6tetrahydro-2,6-methanobenzo[*d*]azocin-3(2*H*)-yl)-2-(*R*/*S*)-phenylpentanoate (0.93 mmoL, 1 eq). The resulting suspension was vigorously stirred and refluxed at 110 °C for 5 h. After, the mixture was cooled at rt, transferred to a separatory funnel, and partitioned (CHCl₃/H₂O). A 1 N solution of HCl was added to the aqueous phase until a pH range of 5–6. The obtained yellow precipitate was separated from the aqueous phase by vacuum filtration. Yield: 52%; Mp: 195 °C dec; $[α]_D^{25} = -40.9°$ (*c* 1.08, EtOH); TLC CH₂Cl₂/MeOH (95:5 *v/v*) $R_f = 0.20$; ¹H NMR (200 MHz, CD₃OD): $\delta = 7.47-7.22$ (m, 5H), 7.05 (d, 1H, J = 10.0 Hz), 6.80 (d, 1H, J = 5.0 Hz), 6.72 (dd, 1H, J = 5.0, 10.0 Hz), 4.28–4.24 (m, 1H), 3.72–3.54 (m, 1H), 3.23–3.11 (m, 5H), 2.20–2.00 (m, 4H), 1.82–1.59 (m, 4H), 1.46 (s, 3H), 0.98 (d, 3H). (Figure S9) ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 177.57$, 174.23, 161.29,155.83, 142.97, 139.52, 128.41,128.12, 127.77,127.61, 127.30, 126.87, 113.48, 112.23, 94.66, 91.20, 72.87, 57.00, 50.97, 44.83, 30.95, 28.82, 24.98, 19.45, 13.47. Anal (C₂₅H₃₁NO₃) C, H, N (Table S1).

4.4 | Radioligand binding assays

The radioligand binding assays and the data analysis were performed as previously reported (Rita Turnaturi et al., 2022).

4.5 | Molecular modeling

4.5.1 | Structures preparation and minimization

All the molecules used in this study were built using Marvin Sketch (18.24, ChemAxon Ltd.). The PM6-D3H4 Hamiltonian, implemented in the MOPAC package (MOPAC2016 v. 18.151, Stewart Computational Chemistry, Colorado Springs), was then used to further optimize the 3D structures before the alignment for the docking calculations.

4.5.2 | Docking and molecular dynamics studies

Flexible ligands docking experiments were performed employing AutoDock implemented in YASARA, using the crystal structure of MOR (PDB code: 5C1M) (Huang et al., 2015), the crystal structure of DOR (PDB code: 4EJ4) (Granier et al., 2012) and crystal structure of KOR (PDB ID: 4DJH) (Wu et al., 2012) retrieved from the PDB_REDO Data Bank. A periodic simulation cell with boundaries extending 5 Å (Duan et al., 2003) from the surface of the ligand was employed.

The molecular dynamics (MD) simulations of the complexes were performed with the YASARA structure package according to our previously reported procedures (Floresta et al., 2019; Patamia et al., 2023).

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CONFLICT OF INTEREST STATEMENT

The authors declare no competing financial interest.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available on request from the corresponding author. The data are not publicly available due to privacy or ethical restrictions.

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SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

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