Correlation between the overexpression of Yin Yang 1 and the expression levels of miRNAs in Burkitt's lymphoma: A computational study

SAMEH HAFSI^{1,2}, SAVERIO CANDIDO¹, ROBERTA MAESTRO³, LUCA FALZONE¹, ZORA SOUA², BENJAMIN BONAVIDA⁴, DEMETRIOS A. SPANDIDOS⁵ and MASSIMO LIBRA¹

¹Department of Biomedical and Biotechnological Sciences, Section of General and Clinical Pathology, and Oncology, University of Catania, I-95124 Catania, Italy; ²U.R. Molecular Biology of Leukemia and Lymphoma, Faculty of Medicine, University of Sousse, Sousse 4002, Tunisia; ³Experimental Oncology 1, CRO Aviano National Cancer Institute, IRCCS, I-33081 Aviano, Italy; ⁴Department of Microbiology, Immunology and Molecular Genetics, Jonsson Comprehensive Cancer Center, University of California, Los Angeles, CA 90095, USA; ⁵Laboratory of Clinical Virology, University of Crete Medical School, Heraklion 71409, Greece

Received November 5, 2015; Accepted December 14, 2015

DOI: 10.3892/ol.2015.4031

Abstract. A growing number of studies have highlighted the role of microRNAs (miRNAs or miRs) in the development and progression of cancer. In particular, the aberrant expression of cancer-related proteins, such as oncogenes and tumor suppressors has been shown to correlate with the modulation of the expression of specific miRNAs. In the present study, we aimed to determine which downregulated miRNAs may be involved in modulating the expression of the oncogenic transcription factor, Yin Yang 1 (YY1). YY1 has been reported to be overexpressed in several malignancies and our previous studies have highlighted the significant correlation between the levels of YY1 and aggressive behavior in non-Hodgkin's lymphoma (NHL). A total of 57 miRNAs that are potentially capable of targeting YY1 was identified through in silico approaches. The search of publicly available NHL datasets, including paired mRNA and miRNA data (GSE23026) highlighted a significant correlation (Pearson's correlation, r>0.5) between the expression levels of YY1 and the expression levels of a limited set of miRNAs, including miR-363, miR-200a, miR-23b, miR-15a and miR-15b. Intriguingly, both hsa-miR-363 and hsa-miR-200a belong to the top 20 miRNAs that were found to be downregulated in Burkitt's lymphoma (BL) tissue compared to normal tissue. Although

E-mail: m.libra@unict.it

further validation studies are warranted, the identification of these two miRNAs associated with the upregulation of YY1 in BL may provide further insight into the pathogenesis of this tumor and may contribute to more personalized and targeted treatment approaches for patients with this disease.

Introduction

MicroRNAs (miRNAs or miRs) are small non-coding RNAs, comprising 19-25 nucleotides, that reduce the abundance and translational efficiency of mRNAs and play a major role in regulatory networks, influencing diverse biological processes through the effects of individual miRNAs on the translation of multiple mRNAs (1,2). Determining the roles of individual miRNAs in cellular regulatory processes poses a major challenge with the function of the majority of miRNAs that are currently unknown; even for relatively well-studied miRNAs, only a handful of targets have been rigorously characterized and these may differ by tissue type.

miRNAs post-transcriptionally regulate the expression of thousands of genes, including key genes in cell differentiation and cancer pathogenesis (3). A growing number of studies have highlighted the role of miRNAs in the development and progression of cancer (4,5). In particular, the aberrant expression of cancer-related proteins, such as oncogenes and tumor suppressors has been shown to correlate with the modulation of the expression specific miRNAs.

Non-Hodgkin's lymphomas (NHLs) are a heterogeneous group of malignancies of the lymphoid system. NHLs are classified based on cytology, immunophenotype and both genetic and clinical characteristics (6). Genetic aberration and molecular changes are used for a more appropriate diagnostic profile. The most commonly associated genetic abnormality in NHL is the translocation of t(14;18)(q32;q21) (7), mostly detected in low-grade lymphomas, such as follicular lymphoma; while molecular rearrangements involving Bcl-6 or c-Myc are frequently observed in diffuse large B-cell lymphoma (DLBCL)

Correspondence to: Dr Massimo Libra, Department of Biomedical and Biotechnological Sciences, Section of General and Clinical Pathology, and Oncology, University of Catania, 83 Via Androne, I-95124 Catania, Italy

Key words: hsa-miR-363, hsa-miR-200a, Yin Yang 1, Burkitt's lymphoma

and Burkitt's lymphoma (BL), respectively. Currently, a significant percentage of patients with NHL are successfully cured; however, a fraction of these patients may relapse following treatment due to the development of chemoresitance (8,9).

It has been demonstrated that the transcription factor, Yin Yang 1 (YY1), is involved in the regulation of resistance to chemotherapy and immunotherapy in NHL cell lines. In addition, the regulation of Fas resistance by nuclear factor (NF)- κ B is mediated via YY1 expression and activity (10). YY1 is known to play a critical role in normal biological processes, such as embryogenesis, differentiation, replication and cell proliferation. YY1 exerts its effects on genes involved in these processes through its ability to initiate, activate, or repress transcription, depending upon the context in which it binds (11).

It has also been reported that YY1 is overexpressed in several malignancies, including B-cell NHL (12,13), and this overexpression is associated with aggressive behaviour in NHL (14). Based on these findings, in the current study, miRNAs potentially capable of targeting and modulating YY1 were identified through *in silico* approaches.

Data collection methods

In order to identify putative miRNAs that target YY1 mRNA, several publicly available miRNA target prediction tools were used, including DIANA-microT (15), miRanda (16), PicTar (17) and TargetScanS (18). Publicly available Gene Expression Omnibus datasets of NHL specimens with the relative normal counterparts were analyzed. Only datasets with both mRNA and miRNA expression levels were considered for the analysis. According to these criteria, the dataset GSE23026 was used in our study (19). This dataset includes 12 samples of BL, 5 tonsil specimens, 7 lymph node specimens and 4 samples of CD19+CD10+B-cells.

Statistical analysis. The differential analysis of YY1 expression levels was performed using the ANOVA test. Correlation analysis between the miRNA expression and YY1 expression levels was performed using the Pearson correlation test.

Results

The analysis using the Diana-microT, miRanda, PicTar, TargetScanS tools revealed a total of 57 predicted miRNAs capable of targeting YY1 (Fig. 1).

The search of publicly available NHL datasets, including paired mRNA and miRNA data (GSE23026) highlighted a significant negative correlation (Pearson correlation coefficient \leq -0.5) between the YY1 expression levels and the expression of a limited set of miRNAs, including miR-363, miR-200a, miR-23b, miR-15a and miR-15b among the 57 predicted miRNAs capable of targeting YY1 (Table I).

As was expected, the transcript expression levels of YY1 were found to be significantly higher in the BL samples compared to the normal tissue samples (mean value expression for BL=0.458, tonsil=-0.248, lymph node=0.049 and CD19⁺CD10⁺ B-cells=0.197; ANOVA, P=0.007) (Fig. 2). Intriguingly, both hsa-miR-363 and hsa-miR-200a belong to the top 20 miRNAs that were found to be downregulated in the BL samples compared to their normal counterparts (Table II).

Table I. Pearson correlation analysis between microRNAs targeting YY1 and YY1 expression levels.

miRNA	Pearson correlation $(r \le -0.5)$	miRNA ID
^a hsa-miR-363	-0.611	A_25_P00010953
ªhsa-miR-200a	-0.608	A_25_P00010209
hsa-miR-23b	-0.577	A_25_P00010881
hsa-miR-200a	-0.576	A_25_P00010208
hsa-miR-15a	-0.552	A_25_P00010467
^a hsa-miR-363	-0.549	A_25_P00010954
hsa-miR-15b	-0.517	A_25_P00011101

^aMicroRNAs (miRNAs or miRs) belonging to the top 20 miRNAs that were found to be differentially expressed in Burkitt's lymphoma samples compared to the normal tissue samples. YY1, Yin Yang 1.

microRNA	DIANA-microT	miRanda	PicTar	TargetScanS	m	icroRNA	DIANA-microT	miRanda	PicTar	TargetScanS
hsa-miR-1					hs	sa-miR-181d				
hsa-miR-15a					hs	sa-miR-182				
hsa-miR-15b					hs	sa-miR-186				
hsa-miR-18					hs	sa-miR-188				
hsa-miR-18b					hs	sa-miR-200a				
hsa-miR-18a					hs	sa-miR-200c				
hsa-miR-22					hs	sa-miR-206				
hsa-miR-23a					hs	sa-miR-302c*				
hsa-miR-23b					hs	sa-miR-361				
hsa-miR-29a					hs	sa-miR-363				
hsa-miR-29b					hs	sa-miR-374				
hsa-miR-29c					hs	sa-miR-381				
hsa-miR-30e					hs	sa-miR-384				
hsa-miR-31					hs	sa-miR-384				
hsa-miR-34a					hs	sa-miR-410				
hsa-miR-34c					hs	sa-miR-433				
hsa-miR-92					hs	sa-miR-449				
hsa-miR-101					hs	sa-miR-493-3p				
hsa-miR-130a					hs	sa-miR-496				
hsa-miR-130b					hs	sa-miR-511				
hsa-miR-134					hs	sa-miR-515-5p				
hsa-miR-141					hs	sa-miR-519e*				
hsa-miR-142-5p					hs	sa-miR-520d*				
hsa-miR-144					hs	sa-miR-524*				
hsa-miR-147					hs	sa-miR-539				
hsa-miR-154					hs	sa-miR-548c				
hsa-miR-181a					hs	sa-miR-573				
hsa-miR-181b					hs	sa-miR-648				
hsa-miR-181c										

Figure 1. Predicted miRNAs capable of targeting YY1 identified using the Diana-microT, miRanda, PicTar and TargetScanS tools. The tools that recognized each predicted miRNA are indicated by the blue-colored boxes.

Discussion

A growing body of evidence has indicated that deregulated miRNA expression is observed in a variety of human diseases,



Figure 2. Yin Yang 1 transcript levels in Burkitt's lymphoma (GSE23026) are reported as boxed quartiles (median, 25th, and 75th percentile) and whiskers (minimum and maximum). The differences between tumor and normal samples are compared by the ANOVA test.

Table II. Top 20 microRNAs downregulated in Burkitt's lymphoma
samples compared to normal tissue samples.

miRNA	P-value	FC (log)	miRNA ID
hsa-miR-150	2.32E-04	-3.991	A_25_P00010490
hsa-miR-150	3.13E-04	-3.869	A_25_P00010489
hsa-miR-363	1.71E-04	-3.745	A_25_P00010954
hsa-miR-99a	1.10E-06	-3.686	A_25_P00010471
hsa-miR-10b_v9.1	5.03E-05	-3.657	A_25_P00010136
hsa-miR-10b_v9.1	3.53E-05	-3.607	A_25_P00010135
hsa-miR-139_v9.1	2.73E-05	-3.594	A_25_P00010349
hsa-miR-363	9.85E-05	-3.547	A_25_P00010953
hsa-let-7g_v9.1	7.09E-06	-3.431	A_25_P00010306
hsa-miR-31_v9.1	6.93E-05	-3.426	A_25_P00010510
hsa-let-7g_v9.1	5.21E-06	-3.404	A_25_P00010305
hsa-miR-200b_v9.1	1.01E-03	-3.389	A_25_P00010908
hsa-let-7a	7.41E-07	-3.37	A_25_P00011584
hsa-let-7f	1.59E-06	-3.335	A_25_P00010088
hsa-miR-195	5.00E-06	-3.25	A_25_P00010769
hsa-miR-133b	7.41E-04	-3.233	A_25_P00010963
hsa-miR-195	9.00E-06	-3.198	A_25_P00010770
hsa-let-7e_v9.1	1.04E-06	-3.192	A_25_P00010280
hsa-miR-200a	8.78E-04	-3.184	A_25_P00010209
hsa-let-7e_v9.1	6.81E-07	-3.181	A_25_P00010279

Analysis of microRNAs (miRNAs or miRs) was performed using GEO2R tools. FC, fold change.

including lymphomas, suggesting an important role in their pathogenesis (20,21). The data from several experimental studies have supported an important role for miRNAs in lymphomagenesis through systematic miRNA profiling in lymphoma samples using a variety of miRNA expression platforms (22-24). YY1 is an activator and a repressor transcription factor belonging to the Zinc-finger proteins (25). It is a positive regulator of several oncogenes, such as c-Fos (26), c-Myc (27) and Erb-B2 receptor tyrosine kinase 2 (ERBB2) (28,29). However, it is also a negative regulator of a fraction of tumor suppressor genes, including p27 (30), p16 (31), p73 (32) and p53 (33). YY1 plays a role in the malignant transformation of several diseases, including B-cell lymphomas. Based on these findings, the aim of the present study was to identify, through *in silico* approaches, which miRNAs are potentially capable of targeting and modulating YY1.

Computational evaluation is an appropriate approach for an initial screening to identify deregulated miRNAs. Based on our results, we speculated that YY1 overexpression may be caused by the downregulation of both hsa-miR-363 and hsa-miR-200a, at least in BL. In fact, the overexpression of YY1 negatively correlated with the expression of hsa-miR-363 and hsa-miR-200a. In turn, the expression levels of hsa-miR-363 and hsa-miR-200a were lower in the BL samples when compared with the normal tissue samples. The increased transcript levels of YY1 in the BL samples is in agreement with the observations of a previous study in which the expression of YY1 was found to be associated with high-grade lymphomas, suggesting that it is linked with post-germinal center transit, where genetic alterations may occur, such as Bcl-6 mutations and/or Bcl-6 translocations (34). These findings may provide further insight into the pathogenesis of BL, and YY1 may thus be considered as a molecular signature of aggressiveness in high-grade lymphomas.

Notably, it has been previously reported that the overexpression of YY1 may promote p53 degradation or inhibit its transcriptional activity (35). Furthermore, the inhibition of YY1 in cancer cells, which lack functional p53, also sensitizes them to cell death in response to apoptotic stimuli (36,37). Although p53 is seldom mutated in NHL, the p53 pathway has been reported to be attenuated in these tumors (38). Among the targets of p53, miR-34a has been found to be repressed as a consequence of promoter methylation (39). These findings support the notion that YY1 may directly control the expression of genes involved in the apoptotic machinery independently of the p53 protein. However, certain drug-resistant hematopoietic disorders that exhibit wild-type p53 may benefit from targeted therapy (40).

In agreement with our results, the loss of miR-200a expression in primary cutaneous melanoma is considered as an additional indicator of aggressiveness, as it was associated with increased thickness and disease progression (41).

Low levels of miR-200a have also been observed in mucosaassociated lymphoid tissue lymphoma (MALT) lymphoma of the conjunctiva. It was also found that cyclin E2 is a target of miR-200a (42). Accordingly, the putative role of YY1 in tumorigenesis is sustained by its interaction with cell cycle regulation. The downregulated expression of miR-363 can precisely differentiate the high-grade lymphomas into ABC or GCB subtypes, and its expression has been shown to correlate with the clinical outcome (43). Similarly, the downregulation of miR-363 has also been detected in ovarian cancer and has been shown to be associated with the drug-resistance phenomenon (44).

Although further validation studies are warranted, the results of the present study indicate that the negative correlation between miR-200a and miR-363 and YY1 expression, which is observed in BL, may provide further insight into the pathogenesis of BL and may have implications on treatment strategies. Furthermore, our data suggest that miRNA expression profiling provides a useful tool for a better understanding of cancer development and progression and for the identification of molecular predictors of response and outcome.

Acknowledgements

This study was supported in part by Lega Italiana per la Lotta contro i Tumori and by the Jonsson Cancer Research Center.

References

- 1. Baek D, Villén J, Shin C, Camargo FD, Gygi SP and Bartel DP: The impact of microRNAs on protein output. Nature 455: 64-71, 2008.
- 2. Bartel DP: MicroRNAs: Target recognition and regulatory functions. Cell 136: 215-233, 2009.
- Johnson SM, Grosshans H, Shingara J, Byrom M, Jarvis R, Cheng A, Labourier E, Reinert KL, Brown D and Slack FJ: RAS is regulated by the let-7 microRNA family. Cell 120: 635-647, 2005.
- Iorio MV and Croce CM: MicroRNA dysregulation in cancer: Diagnostics, monitoring and therapeutics. A comprehensive review. EMBO Mol Med 4: 143-159, 2012.
- 5. Di Leva G, Garofalo M and Croce CM: MicroRNAs in cancer. Annu Rev Pathol 9: 287-314, 2014.
- Jaffe ES: The 2008 WHO classification of lymphomas: Implications for clinical practice and translational research. Hematology Am Soc Hematol Educ Program 2009: 523-531, 2009.
- Yunis JJ, Frizzera G, Oken MM, McKenna J, Theologides A and Arnesen M: Multiple recurrent genomic defects in follicular lymphoma. A possible model for cancer. N Engl J Med 316: 79-84, 1987.
- Ansell SM: Non-Hodgkin Lymphoma: Diagnosis and Treatment. Mayo Clin Proc 90: 1152-1163, 2015.
- 9. Mehta-Shah N and Younes A: Novel targeted therapies in diffuse large B-cell lymphoma. Semin Hematol 52: 126-137, 2015.
- Vega MI, Jazirehi AR, Huerta-Yepez S and Bonavida B: Rituximab-induced inhibition of YY1 and Bcl-xL expression in Ramos non-Hodgkin's lymphoma cell line via inhibition of NF-kappa B activity: role of YY1 and Bcl-xL in Fas resistance and chemoresistance, respectively. J Immunol 175: 2174-2183, 2005.

- 11. Gordon S, Akopyan G, Garban H and Bonavida B: Transcription factor YY1: Structure, function, and therapeutic implications in cancer biology. Oncogene 25: 1125-1142, 2006.
- 12. Castellano G, Torrisi E, Ligresti G, Malaponte G, Militello L, Russo AE, McCubrey JA, Canevari S and Libra M: The involvement of the transcription factor Yin Yang 1 in cancer development and progression. Cell Cycle 8: 1367-1372, 2009.
- 13. Bonavida B, Huerta-Yepez S, Baritaki S, Vega M, Liu H, Chen H and Berenson J: Overexpression of Yin Yang 1 in the pathogenesis of human hematopoietic malignancies. Crit Rev Oncog 16: 261-267, 2011.
- 14. Castellano G, Torrisi E, Ligresti G, Nicoletti F, Malaponte G, Traval S, McCubrey JA, Canevari S and Libra M: Yin Yang 1 overexpression in diffuse large B-cell lymphoma is associated with B-cell transformation and tumor progression. Cell Cycle 9: 557-563, 2010.
- Maragkakis M, Alexiou P, Papadopoulos GL, Reczko M, Dalamagas T, Giannopoulos G, Goumas G, Koukis E, Kourtis K, Simossis VA, *et al*: Accurate microRNA target prediction correlates with protein repression levels. BMC Bioinformatics 10: 295, 2009.
- John B, Enright AJ, Aravin A, Tuschl T, Sander C and Marks DS: Correction: Human MicroRNA targets. PLoS Biol 3: e264, 2005.
- 17. Krek A, Grün D, Poy MN, Wolf R, Rosenberg L, Epstein EJ, MacMenamin P, da Piedade I, Gunsalus KC, Stoffel M, et al: Combinatorial microRNA target predictions. Nat Genet 37: 495-500, 2005.
- Agarwal V, Bell GW, Nam JW and Bartel DP: Predicting effective microRNA target sites in mammalian mRNAs. Elife 4: e05005, 2015.
- 19. Bueno MJ, Gómez de Cedrón M, Gómez-López G, Pérez de Castro I, Di Lisio L, Montes-Moreno S, Martínez N, Guerrero M, Sánchez-Martínez R, Santos J, et al: Combinatorial effects of microRNAs to suppress the Myc oncogenic pathway. Blood 117: 6255-6266, 2011.
- 20. Di Lisio L, Sánchez-Beato M, Gómez-López G, Rodríguez ME, Montes-Moreno S, Mollejo M, Menárguez J, Martínez MA, Alves FJ, Pisano DG, *et al*: MicroRNA signatures in B-cell lymphomas. Blood Cancer J 2: e57, 2012.
- 21. Troppan K, Wenzl K, Deutsch A, Ling H, Neumeister P and Pichler M: MicroRNAs in diffuse large B-cell lymphoma: implications for pathogenesis, diagnosis, prognosis and therapy. Anticancer Res 34: 557-564, 2014.
- 22. Musilova K and Mraz M: MicroRNAs in B-cell lymphomas: How a complex biology gets more complex. Leukemia 29: 1004-1017, 2015.
- Garofalo M, Leva GD and Croce CM: MicroRNAs as anti-cancer therapy. Curr Pharm Des 20: 5328-5335, 2014.
 Lim EL, Trinh DL, Scott DW, Chu A, Krzywinski M,
- 24. Lim EL, Trinh DL, Scott DW, Chu A, Krzywinski M, Zhao Y, Robertson AG, Mungall AJ, Schein J, Boyle M, *et al*: Comprehensive miRNA sequence analysis reveals survival differences in diffuse large B-cell lymphoma patients. Genome Biol 16: 18, 2015.
- 25. Shi Y, Seto E, Chang LS and Shenk T: Transcriptional repression by YY1, a human GLI-Krüppel-related protein, and relief of repression by adenovirus E1A protein. Cell 67: 377-388, 1991.
- Lee HY, Chaudhary J, Walsh GL, Hong WK and Kurie JM: Suppression of c-Fos gene transcription with malignant transformation of human bronchial epithelial cells. Oncogene 16: 3039-3046, 1998.
- 27. Liao WR, Hsieh RH, Hsu KW, Wu MZ, Tseng MJ, Mai RT, Wu Lee YH and Yeh TS: The CBF1-independent Notch1 signal pathway activates human c-myc expression partially via transcription factor YY1. Carcinogenesis 28: 1867-1876, 2007.
- 28. Allouche A, Nolens G, Tancredi A, Delacroix L, Mardaga J, Fridman V, Winkler R, Boniver J, Delvenne P and Begon DY: The combined immunodetection of AP-2alpha and YY1 transcription factors is associated with ERBB2 gene overexpression in primary breast tumors. Breast Cancer Res 10: R9, 2008.
- Begon DÝ, Delacroix L, Vernimmen D, Jackers P and Winkler R: Yin Yang 1 cooperates with activator protein 2 to stimulate ERBB2 gene expression in mammary cancer cells. J Biol Chem 280: 24428-24434, 2005.
 Wan M, Huang W, Kute TE, Miller LD, Zhang Q, Hatcher H,
- 30. Wan M, Huang W, Kute TE, Miller LD, Zhang Q, Hatcher H, Wang J, Stovall DB, Russell GB, Cao PD, *et al*: Yin Yang 1 plays an essential role in breast cancer and negatively regulates p27. Am J Pathol 180: 2120-2133, 2012.
- Rayess H, Wang MB and Srivatsan ES: Cellular senescence and tumor suppressor gene p16. Int J Cancer 130: 1715-1725, 2012.

- Wu S, Murai S, Kataoka K and Miyagishi M: Cooperative regulation og p73 promoter by Yin Yang 1 and E2F1. Nucleic Acids Symp Ser (Oxf) 51: 347-348, 2007.
- 33. Grönroos E, Terentiev AA, Punga T and Ericsson J: YY1 inhibits the activation of the p53 tumor suppressor in response to genotoxic stress. Proc Natl Acad Sci USA 101: 12165-12170, 2004.
- 34. Migliazza A, Martinotti S, Chen W, Fusco C, Ye BH, Knowles DM, Offit K, Chaganti RS and Dalla-Favera R: Frequent somatic hypermutation of the 5' noncoding region of the BCL6 gene in B-cell lymphoma. Proc Natl Acad Sci USA 92: 12520-12524, 1995.
- 35. Libra M, Torrisi E, Castellano G, Nicoletti F, Malaponte G, Mazzarino MC, Marconi A, Proietti L, Militello L, Bonavida B, *et al*: Computational Evaluation of Yin Yang 1 Transcript Levels in the Spectrum of B-cell Neoplasia. Immunopathol Dis Therap 1: 115-125, 2010.
- 36. Sui G, Affar B, Shi Y, Brignone C, Wall NR, Yin P, Donohoe M, Luke MP, Calvo D, Grossman SR, *et al*: Yin Yang 1 is a negative regulator of p53. Cell 117: 859-872, 2004.
- 37. Affar B, Gay F, Shi Y, Liu H, Huarte M, Wu S, Collins T, Li E and Shi Y: Essential dosage-dependent functions of the transcription factor yin yang 1 in late embryonic development and cell cycle progression. Mol Cell Biol 26: 3565-3581, 2006.
- 38. Maestro R, Gloghini A, Doglioni C, Piccinin S, Vukosavljevic T, Gasparotto D, Carbone A and Boiocchi M: Human non-Hodgkin's lymphomas overexpress a wild-type form of p53 which is a functional transcriptional activator of the cyclin-dependent kinase inhibitor p21. Blood 89: 2523-2528, 1997.

- Chim CS, Wong KY, Qi Y, Loong F, Lam WL, Wong LG, Jin DY, Costello JF and Liang R: Epigenetic inactivation of the miR-34a in hematological malignancies. Carcinogenesis 31: 745-750, 2010.
- 40. McCubrey JA, Abrams SL, Ligresti G, Misaghian N, Wong EW, Steelman LS, Bäsecke J, Troppmair J, Libra M, Nicoletti F, *et al*: Involvement of p53 and Raf/MEK/ERK pathways in hematopoietic drug resistance. Leukemia 22: 2080-2090, 2008.
- 41. van Kempen LC, van den Hurk K, Lazar V, Michiels S, Winnepenninckx V, Stas M, Spatz A and van den Oord JJ: Loss of microRNA-200a and c, and microRNA-203 expression at the invasive front of primary cutaneous melanoma is associated with increased thickness and disease progression. Virchows Arch 461: 441-448, 2012.
- 42. Cai J, Liu X, Cheng J, Li Y, Huang X, Li Y, Ma X, Yu H, Liu H and Wei R: MicroRNA-200 is commonly repressed in conjunctival MALT lymphoma, and targets cyclin E2. Graefes Arch Clin Exp Ophthalmol 250: 523-531, 2012.
- Sandhu SK, Croce CM and Garzon R: Micro-RNA expression and function in lymphomas. Adv Hematol 2011: 347137, 2011.
- 44. Zou J, Yin F, Wang Q, Zhang W and Li L: Analysis of microarray-identified genes and microRNAs associated with drug resistance in ovarian cancer. Int J Clin Exp Pathol 8: 6847-6858, 2015.