

# Chitinase 3 Like-1: An Emerging Molecule Involved in Diabetes and Diabetic Complications

Michelino Di Rosa Lucia Malaguarnera

Department of Biomedical and Biotechnological Sciences, School of Medicine,  
University of Catania, Catania, Italy

## Key Words

Diabetes mellitus · Chitinase 3 like-1 · Endothelial dysfunction · Macrophage activation · Inflammation · Diabetic complications

## Abstract

Chitinase 3 like-1 (CHI3L1) is a chitinase-like protein member of family 18 chitinases, expressed in innate immune cells and involved in endothelial dysfunction and tissue remodelling. Since CHI3L1 is highly expressed in a variety of inflammatory diseases of infectious and non-infectious aetiology, it is recognised as a non-invasive prognostic biomarker for inflammation. A variety of studies revealing the increase in CHI3L1 levels in obesity, insulin resistance and in pathological conditions, such as atherosclerosis, coronary artery disease, acute ischaemic stroke, nephropathy, diabetic retinopathy and osteolytic processes, have suggested that CHI3L1 may also play a critical role in the evolution and complication of diabetes mellitus (DM). In this review we highlight the impact of CHI3L1 expression in DM and its contribution to the complication of this disease.

© 2016 S. Karger AG, Basel

## Introduction

Diabetes mellitus (DM) is distinguished by metabolic defects resulting from dysfunction in the glucose-handling machinery of the body. It arises in two major forms. Type 1 DM (T1DM), which is the most common form of DM in children, may have an autoimmune or post-infectious aetiology and is caused by a decrease in insulin, which regulates glucose metabolism. Type 2 DM (T2DM) typically involves insulin resistance (IR) at the cellular level and comprises the majority of adult-onset DM. T2DM may rarely occur in children. The metabolic defects of DM are common in all forms of the disease. The reduction of insulin activity causes persistent hyperglycaemia, which leads to osmotic and oxidative stress and results in injury to the nerves, kidneys, eyes, liver, articular cartilage and other tissues. The pathophysiology of DM involves genetics, epigenetics and environmental factors, together with immune disturbance [1–3]. The absence of physical activity in combination with excess calorie intake leads to obesity and autoimmunity onset in developed societies [3]. Obesity is reaching epidemic proportions and is one of the most important chronic diseases leading to an increased risk for several metabolic complications. The dietary habits in Western societies and a high body mass index (BMI) constitute risk factors for autoimmune diseases [4]. Adipose tissue represents a

complex and highly active metabolic and endocrine organ [5] and plays a key role in the pathogenesis of IR through several released molecules that can affect different steps in insulin action. IR is the most common metabolic alteration associated with obesity, which in turn generates other metabolic diseases as well as cardiovascular complications [6]. Obesity induces a state of chronic low-grade inflammation because of the circulating levels of markers of inflammation, including both pro-inflammatory cytokines and acute-phase proteins [7]. In response to environmental stresses, macrophages, endothelial cells and adipocytes release inflammatory cytokines, including the interleukins (IL)-1, IL-6, tumour necrosis factor- $\alpha$  (TNF- $\alpha$ ), and chemokines that, in turn, induce the production of acute-phase proteins, such as C-reactive protein (CRP) and amyloid deposition in the pancreas [8]. The cellular stresses, such as oxidative stress, ER stress, lipotoxicity and glucotoxicity, exacerbate IR, islet  $\beta$ -cell dysfunction and induce the inflammatory response that is related to this pathophysiological state. In this context, increasing evidence indicates that chitinase 3-like 1 (CHI3L1), a member of the chitinases family, is also involved also in the pathogenesis of DM [9, 10]. Exaggerated production of CHI3L1 could cause unexpected pathogenic effects in human tissues, directly initiating and perpetuating chronic inflammation. This review considers the relationship between CHI3L1 and DM and its contribution in the complications of this disease.

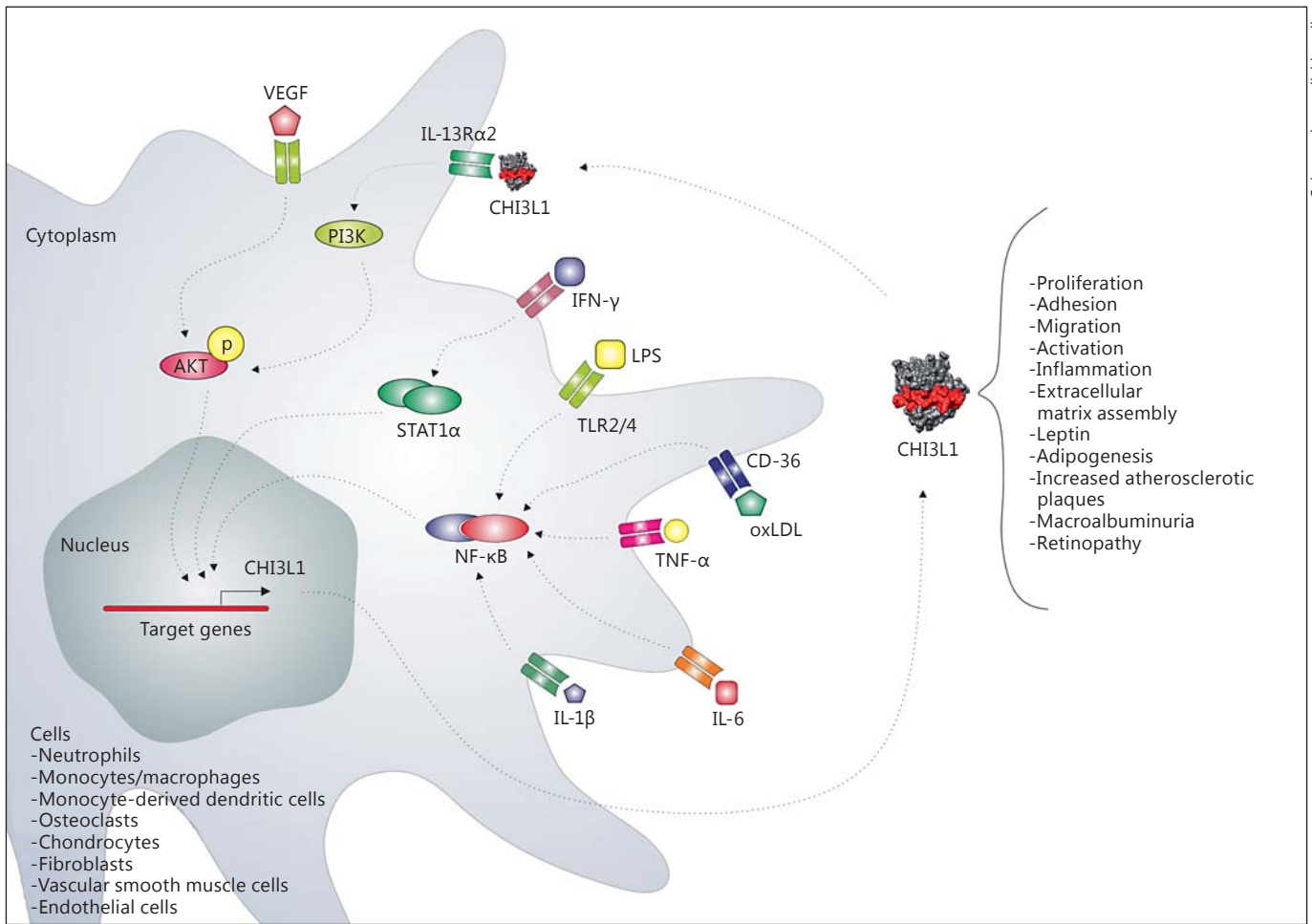
### CHI3L1 Biological Activities

CHI3L1 protein – also called YLK-40, based on its three N-terminal amino acids: tyrosine (Y), lysine (K) and leucine (L) – is a heparin-, chitin- and collagen-binding glycoprotein that was originally discovered in mouse breast cancer cells [11]. CHI3L1 is a member of the protein family of chi-lectins (chitinase-like lectins) [12]. The members of this protein family are structurally related to the 18 glycohydrolase family. On the basis of a substitution of the catalytic essential glutamic acid residue in the active site of this protein, CHI3L1 has no enzymatic activity [13]. Biological activities of CHI3L1 embrace the regulation of cell proliferation, adhesion, migration and activation. CHI3L1 is produced by a variety of cells, including neutrophils, monocytes/macrophages, monocyte-derived dendritic cells, osteoclasts, chondrocytes, fibroblasts, vascular smooth muscle cells and endothelial cells [12, 14–16]. Local inflamed tissues, including adipose tissues in T2DM, produce CHI3L1 locally [17]. In addition,

CHI3L1 stimulating the production of inflammatory mediators (e.g. CCL2, CXCL2, MMP-9) [17] acts as a pro-inflammatory biomarker [18]. Induction of CHI3L1 has been reported in patients suffering from a surprisingly vast array of diseases, including a number of autoimmune disorders [11]. Circulating levels of CHI3L1 are increased in patients with T2DM and positively correlate with IR [19]. In addition, elevated plasma levels of CHI3L1 have been found in liver fibrosis [20], rheumatoid arthritis, atherosclerosis, coronary artery disease, Alzheimer's disease and inflammation-related illnesses in humans [18, 21, 22]. High CHI3L1 levels are predictors of overall and cardiovascular mortality [23]. The expression of CHI3L1 has been shown to be regulated by various pro-inflammatory cytokines, including IL-6, interferon- $\gamma$  (IFN- $\gamma$ ), IL-1 $\beta$  and TNF- $\alpha$  [24]. Therefore, CHI3L1 can be regarded as an acute-phase reactant associated with disease severity and mortality in a variety of infections. Additionally, CHI3L1 stimulates the growth of fibroblast cells [23], promotes proliferation and antagonizes catabolic or degradative processes during the inflammatory response of connective tissues [25], activates the protein kinase B (AKT) and phosphoinositide-3 kinase (PI3K) signalling pathway, and exerts anti-apoptotic [26] and angiogenic function [27] (fig. 1). CHI3L1 plays a crucial role not only during innate but also in acquired immunity [28, 29]. Cellular receptors mediating its biological effects have not been identified; however, its regulation by IL-6 and TNF- $\alpha$  requires sustained activation of the nuclear factor-kappa B (NF- $\kappa$ B), and can bind to collagen types I–III [30]. The ability of CHI3L1 to regulate cell proliferation, adhesion, migration and activation, as well as to regulate extracellular matrix assembly, correlates well with an elevated level of CHI3L1 in the sites of chronic inflammation.

### The Immune System in the Regulation of the DM Metabolism

The islets from patients with DM exhibit the typical histology of inflamed tissue with infiltration of immune cells,  $\beta$ -cell apoptosis containing islet amyloid polypeptide and decreased insulin staining [31]. Elevated glucose levels are perceived by the inflammasomes [32], which, acting as innate immune sensors, detect metabolic danger signals. The inflammasomes activate caspase-1 that cleaves pro-IL-1 $\beta$  into active IL-1 $\beta$ , which, in turn, binds to its membrane receptor and activates the transcription factor NF- $\kappa$ B, resulting in the production of a wide array



**Fig. 1.** The association between CHI3L1 and IR may occur through PI3K, which mediates insulin-stimulated glucose uptake. CHI3L1 binding to a receptor (i.e. IL-13R $\alpha$ 2) is PI3K mediated followed by the phosphorylation of AKT, which leads to CHI3L1 transcription and the ability of CHI3L1 to regulate cell proliferation, adhesion,

migration and activation, as well as the regulation of extracellular matrix assembly. The induction and continued secretion of CHI3L1 requires sustained activation of NF- $\kappa$ B by pro-inflammatory cytokines (IL-1 $\beta$ , TNF- $\alpha$ , IL-6) or inflammation mediators (LPS, oxLDL), which in turn enhance CHI3L1 expression.

of cytokines and chemokines, followed by the recruitment of immune cells, including macrophages. While the inflammatory process contributes to defective insulin secretion, the immune system contributes to  $\beta$ -cell adaptation during states of increased insulin demand, which stimulates glucagon-like peptide-1 (GLP-1) secretion from intestinal L-cells and from the pancreatic  $\alpha$ -cells [33]. IL-6 particularly mediates cross-talk between insulin-sensitive tissues, L-cells and islets to adapt to variations in insulin demand by increasing L-cell GLP-1 secretion and reprogramming  $\alpha$ -cells to process the proglucagon to GLP-1. This endocrine network corroborates the idea of a role for the immune system in the regulation of

the metabolism and the importance of the collaborations between  $\alpha$ - and  $\beta$ -cells [33]. Therefore, inflammatory mediators, such as IL-1 $\beta$ , IL-6, Toll-like receptors (TLRs), macrophages and chemokines, are elevated in pancreatic islets from patients with DM [34] (fig. 1). Increased glucose concentrations enhance free fatty acid (FFA)-induced expression of inflammatory mediators in both human and mouse islets, suggesting that these responses are probably through an IL-1R/TLR-dependent pathway [34]. FFAs stimulate adipose tissue inflammation through the TLR2 and TLR4 pathways, resulting in IR. The activation of TLR4 in pancreatic  $\beta$ -cells affects insulin secretion in response to glucose in both humans and rodents [35].

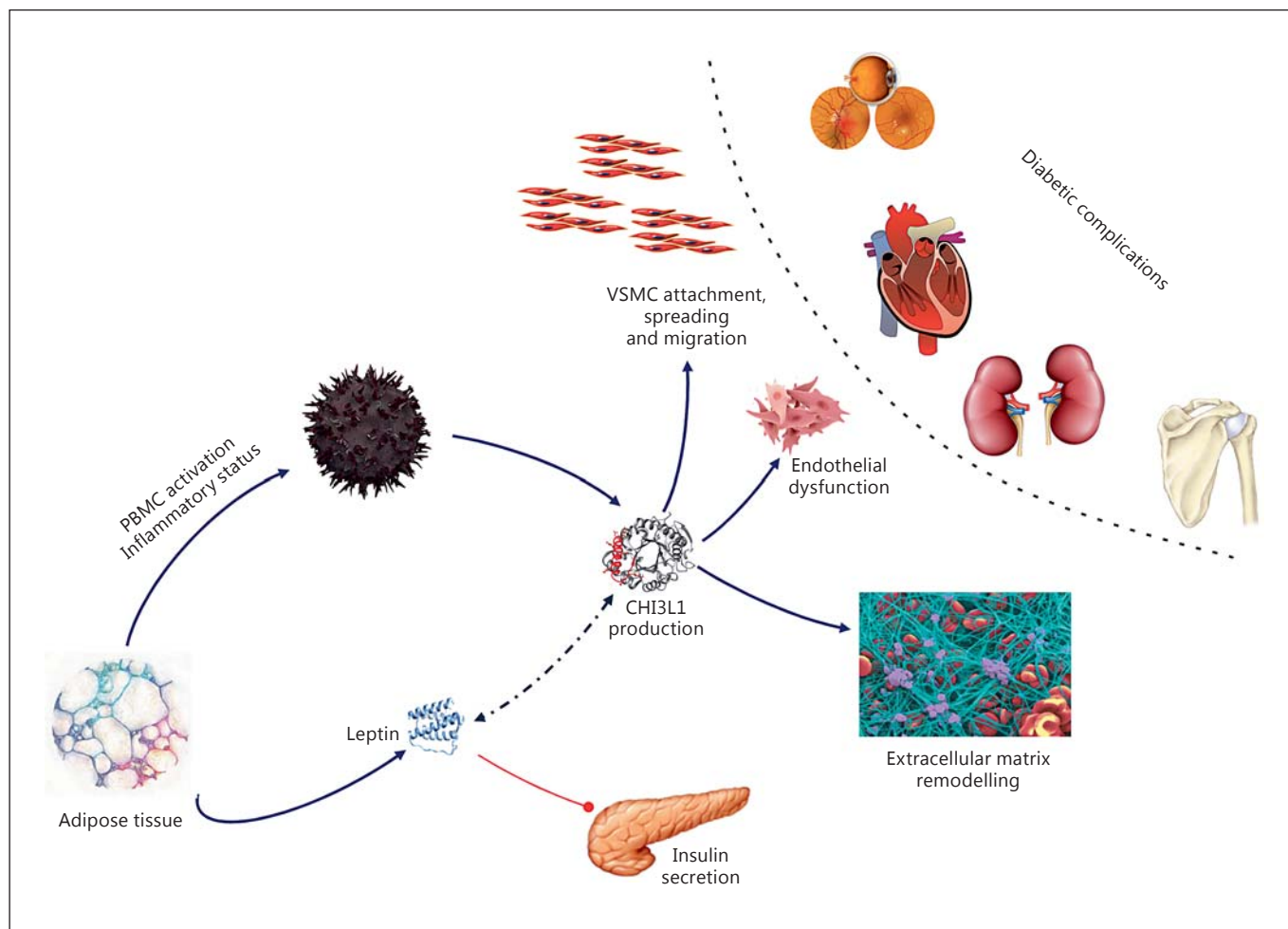
Glucose has been shown to induce TLR2 and TLR4 expression via PKC- $\alpha$  and PKC- $\delta$ , respectively, by stimulating NADPH oxidase in human monocytes [36]. The state of IR may be enabled by the stimulation of TLR-signalling, as suggested by the findings that TLR2- or TLR4-deficient mice are protected against diet-induced IR compared to wild-type animals [37, 38]. In addition, the *in vitro* administration of PAMPs specific for TLR2 or TLR4 stimulates an increase in markers of IR [39, 40]. TLR4 is the major intracellular signalling complex for lipopolysaccharide (LPS) [41] (fig. 1). Circulating levels of LPS correlate with insulin levels, glucose and the homeostatic model assessment of IR (HOMA-IR) [42]. Bolus injections of LPS into healthy human volunteers cause a decrease in the FSIGT (frequently sampled intravenous glucose tolerance) measured insulin sensitivity index [43], indicating that LPS may play a direct role in the progression of IR. Additionally, fetuin-A (FetA), acting as an endogenous ligand for TLR4, plays a crucial role in regulating insulin sensitivity via TLR4 signalling [44]. In FetA knockdown mice, IR caused by a high-fat diet (HFD) resulted in the downregulation of TLR4-mediated inflammatory signalling in adipose tissue, whereas the selective administration of FetA induced inflammatory signalling and IR. Furthermore, it was observed that FFA-induced pro-inflammatory cytokine expression in adipocytes occurred only in the presence of both FetA and TLR4; removing either of them prevented FFA-induced IR [44]. So far, receptors that mediate the effects CHI3L1 have not been defined, and only a potential link between TLR4- and CHI3L1-signalling pathways has been reported [45, 46] (fig. 1).

Nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs), are pattern recognition receptors (PRRs) similar to TLRs that propagate inflammatory signals in response to peptidoglycan [47]. Therefore, NLRs also play a role in inflammation and IR. This idea was corroborated by the finding that elevated levels of NODs were positively correlated to poor glycaemic control and IR [48]. Remarkably, NOD1/2 double-knockout mice were shown to be protected from HFD-induced IR. In addition, the treatment of 3T3-L1 adipocytes with DAP impaired insulin signalling and glucose uptake, which was reverted by NOD1-specific small interfering RNAs (siRNAs) [49]. Similarly, addition of the NOD2 ligand MDP suppressed basal and insulin-stimulated glucose uptake in L6-GLUT4-myc myotubes [50]. In addition, the elevated expression of NOD1/2 induces the expression of the downstream signalling mediators RIPK2 and NF- $\kappa$ B in monocytes from patients with T2DM [48]. The

stimulation of NOD1/2 by their specific bacterial ligands causes the recruitment of RIPK2, a caspase recruitment domain-containing kinase, which then directly binds to I $\kappa$ B kinase (IKK- $\gamma$ ) to activate NF- $\kappa$ B for the pro-inflammatory response [51] (fig. 1). Moreover, RIPK2 participates in innate immunity specifically by mediating NOD1/2 signalling, but not TLR-mediated immune responses [52]. Another intriguing observation from the study by Shiny et al. [48] was the observation that in monocytes from healthy subjects high glucose enhanced NOD1/2 mRNA expression. Glucose showed a synergistic effect with NOD ligands by augmenting ligand-mediated expression of NOD1/2. Glucose-mediated cellular triggers could be the denominators that differentially regulate PRRs and even contribute to the cross-talk among the different immunomodulators involved in the pro-inflammatory response. Further research is required to determine a possible relationship between PRRs and CHI3L1 overexpression and their role in DM and its complications.

### CHI3L1, Obesity and IR

Obesity has profound effects on immunity and inflammation as a critical contributor to the pathogenesis of these metabolic disorders [53, 54]. A high BMI is associated with increased levels of pro-inflammatory cytokines [54]. Different factors are involved in the obesity-related comorbidities, but chronic low-grade inflammation is an emerging responsible element [55]. During obesity, cells from the innate and adaptive immune system infiltrate insulin-sensitive tissues, starting the inflammatory response [56]. Obesity-associated low-grade inflammation results in the persistent stimulation of the immune system mainly characterized by the infiltration of adipose tissue with macrophages [57] that represent the primary source of numerous circulating inflammatory molecules detected in the obese state, and which are fundamental in the development of IR. Consequently, IR results from a combination of altered functions of insulin target cells and the accumulation of macrophages [58]. Thus, both obesity and low-grade inflammation have been linked to the development of IR and T2DM [59]. Plasma levels of acute-phase proteins, white blood cells and pro-inflammatory cytokines are elevated in obese and T2DM patients, whereas these decrease after weight loss [60]. Interestingly, the gene expression profile of peripheral blood mononuclear cells (PBMCs) reflects the visceral fat volume and may be representative of the inflammatory



**Fig. 2.** CHI3L1 expression and diabetic complications.

status in obesity [61]. PBMCs are of great interest and appropriate for research into mechanisms of immune dysfunction in obesity since they are easily available and reflect the responses of dietary modifications, oxidative stress, endogenous lipid mediators and drugs at the gene expression level [61–64]. PBMCs from obese individuals produce elevated levels of TNF- $\alpha$ , IFN- $\gamma$  and IL-2, whereas the production of the anti-inflammatory cytokine IL-10 is reduced [65]. New factors secreted by adipose tissue promoting inflammatory responses and metabolic dysfunction that have recently been identified include CHI3L1, chemerin, LCN-2 (lipocalin-2) and OPN (osteopontin) [66, 67]. These factors are highly diverse in structure and function. So, whereas OPN and CHI3L1 are directly related to increased inflammation and IR via impaired extracellular matrix remodelling [12, 59], LCN-2 acts as an antagonist to the effect of inflammatory mole-

cules [68]. A recent investigation showed that CHI3L1 is strongly upregulated in obesity independently of the glycaemic state and is associated with different circulating inflammatory markers [66]. CHI3L1 is regulated by an HFD and simultaneously plays an important role in the pathogenesis of asthma and obesity [69]. Elevated CHI3L1 levels are also detectable in obese youth and represent a marker of IR even in childhood [70]. In obesity there is also an increased expression of several chemokine genes in adipose tissue. One of them is MCP-1 (monocyte chemoattractant protein-1), which is predominantly produced in the extracellular space where it primarily acts as a local factor. In addition, MCP-1 influences the function of adipocytes, promotes chemotaxis and the migration of monocytes into the sub-endothelial space, and is an important link between the inflammatory response in adipose tissue and IR [71]. Reportedly, elevated gene expres-

sion levels of CHI3L1 in PBMCs contribute to the worsening of the inflammatory response in obesity. Elevated levels of CHI3L1 have also been reported in the obese prepubertal paediatric population [72], and a significant difference in CHI3L1 levels was found between IR and non-IR subjects [70]. Higher CHI3L1 levels were associated with greater adiposity [73], and a positive correlation between leptin and CHI3L1 levels has been detected in obese subjects, either diabetic or not [73]. Leptin inhibits insulin secretion [74] and, as an adipokine, may represent the missing link between IR and obesity [74] (fig. 2). Systemic low-grade inflammation is related to both circulating and adipose tissue leptin levels in obese women [75]. In addition, CHI3L1 levels were associated with markers of IR, even after adjustment for obesity or inflammation markers [76]. According to different reports, an independent association between CHI3L1 and levels of triglycerides as well as no esterified fatty acids exists [77]. Overall, this evidence strongly confirmed that CHI3L1 is involved in modulating obesity and IR. Therefore, low-grade inflammation and endothelial dysfunction would explain the elevated CHI3L1 levels in diabetic patients. CHI3L1 may be essential in obesity since the association between CHI3L1 and IR may occur through the enzyme PI3K that mediates insulin-stimulated glucose uptake [78]. A crucial signal in the cellular response to CHI3L1 binding to its putative receptor is PI3K-mediated phosphorylation of AKT, which modulates the production of matrix metalloproteinases (MMPs) and chemokines [26]. Moreover, CHI3L1 contributes to the trapping of macrophages into adipose tissue by the inhibition of type I collagenolysis by MMP-1 [26]. Increased CHI3L1 secretion and delivery by its potential receptor might explain its effects on insulin metabolism. The correlation between CHI3L1 and IR could be based on the macrophage infiltration in the adipose tissue. Moreover, it is conceivable that CHI3L1 may contribute to the response to hypoxia in regions of fat depots, as the tissue mass increases during the progressive development of obesity.

### CHI3L1 and Diabetes

Subclinical systemic inflammation and changes of a wide variety of inflammatory markers together with the alterations of the metabolic syndrome, including IR, have been found in DM [79, 80]. Rathcke and Vestergaard [81] first reported that CHI3L1 levels increased significantly in T2DM patients. Additionally, they found a correlation between the elevated levels of CHI3L1 and features of

dyslipidaemia to be up to 2-fold higher in T2DM patients. Interestingly, the higher plasma CHI3L1 levels in T2DM patients with HOMA-IR suggested that CHI3L1 is associated with IR rather than impaired insulin secretion. Later, other investigations confirmed that T2DM patients have elevated plasma CHI3L1 compared with healthy control subjects [76]. In multivariate regression analysis adjusted for age, sex, fitness, and either plasma TNF- $\alpha$  or fasting plasma glucose, they reported significant associations between plasma CHI3L1, fasting plasma glucose and plasma levels of IL-6, but no associations with parameters of obesity [76]. Successively, elevated CHI3L1 levels were also detected in T1DM patients [82]. The evaluation of this enzyme in relation to other conventional inflammation markers showed that CHI3L1 levels reflect inflammation independently of hsCRP, which is an acute-phase reactant usually elevated in patients with T2DM and positively correlated with increased components of the metabolic syndrome [83], as demonstrated both in T2DM and in chronic heart failure [84]. Some studies showed a correlation between CHI3L1 and glycaemic parameters such as HbA<sub>1c</sub>, albuminuria [82] and fasting glucose [76]. In concordance with the previous studies performed in T1DM [82, 85], significantly higher concentrations of CHI3L1 and adiponectin were found in T1DM patients from a Mediterranean area with a longer disease evolution but a low atherosclerotic background [86]. A study investigating CHI3L1 in relation to mortality in a population with T2DM demonstrated that high levels CHI3L1 predict all-cause mortality and cardiovascular mortality in these patients [76]. Furthermore, enhanced CHI3L1 levels predict cardiovascular mortality in individuals without known DM or coronary heart disease after adjustment for known cardiovascular risk factors and markers [87, 88]. Interestingly, several studies have documented that single nucleotide polymorphisms of CHI3L1 (rs10399931, -329 G/A, and rs4950928, -131 C/G) are associated with inter-individual CHI3L1 levels and contribute to the susceptibility of different pathological conditions [89–91]. Although a study described an association between a promoter polymorphism of CHI3L1 and levels of low-density lipoprotein (LDL) [92], no consistent association was found between single nucleotide polymorphisms of CHI3L1 and T2DM, IR and dysregulated glucose homeostasis in a total of 9,438 Danish individuals examined [93]. In view of the aforementioned reports indicating that the increased levels of CHI3L1 in patients with DM correlated with IR, levels of non-esterified fatty acids and triglycerides [94], it is reasonable to assume that CHI3L1 activity may represent a crucial me-

diator playing a detrimental role in mechanisms such as endothelial dysfunction, vascular inflammation and the generation of ROS, which are critical for the progression of diabetes and its complications.

### **CHI3L1 Atherosclerosis and Cardiovascular Complications**

Subclinical inflammation is also involved in the pathogenesis of all stages of atherosclerosis [86], where the induction of acute-phase reactants, proinflammatory cytokines and cell adhesion molecules are associated with the development of myocardial infarction, stroke and peripheral vascular disease, and consequently with cardiovascular mortality [95]. Dysfunction of the endothelium is well known in DM and is the earliest event in atherogenesis, playing a pivotal role in all phases of the atherosclerotic process (from the initiation of the fatty streak to plaque rupture), and is largely responsible for the development of ischaemic heart disease and thrombotic strokes [96]. In human endothelial cells, diabetes influences the modulation of plasminogen activator synthesis by insulin-like growth factor-1 (IGF-1), epidermal growth factor or acidic fibroblast growth factor [97]. The development of IR through a low-grade inflammatory state is influenced by multiple genes that, in turn, could affect inflammatory biomarker levels [98, 99]. Based on recent evidence it is possible that the elevated CHI3L1 levels in DM patients are, at least in part, responsible for the endothelial dysfunction and the later micro- and macrovascular complications of DM. The protein expression of CHI3L1 is increased in human smooth muscle cells of atherosclerotic plaques [100] and serum levels of CHI3L1 correlate with arterial wall stiffness – another measure of endothelial dysfunction [101] (fig. 2). In the process of atherosclerotic plaque formation, where smooth muscle cells migrate through the intima in response to exogenous signals, CHI3L1 promotes vascular smooth muscle cell attachment, spreading and migration. This corroborates the idea that CHI3L1 has a role in the process of atherosclerotic plaque formation in response to exogenous signals, and contributes to the process of restenosis and neointima formation. CHI3L1 also modulates vascular endothelial cell morphology by promoting the formation of branching tubules, indicating that CHI3L1 has a role in angiogenesis by stimulating the migration and reorganization of vascular smooth muscle cells [102]. In addition, in atherosclerotic plaques immunohistochemical analysis demonstrated protein expression of CHI3L1 in human

smooth muscle cells [103]. The association between CHI3L1 and atherosclerosis has been related to macrophage activation, as also suggested by the demonstration that the inflammatory cytokine TNF- $\alpha$  enhances CHI3L1 expression in macrophages, and confirmed by the finding that TLR2 and TLR4 activation (i.e. by LPS) further increases the release of CHI3L1 in TNF- $\alpha$ -activated THP-1 monocytes [104] (fig. 1). Additionally, in patients with carotid atherosclerosis, CHI3L1 enhances both gelatinolytic and MMP-9 activity in PBMCs and in THP-1 monocytes via p38 mitogen-activated protein kinase (MAPK) activation [104]. Further studies showing the association between CHI3L1 and ‘pro-atherogenic’ chemokines such as MCP-1 and IL-8 were crucial in understanding the role of CHI3L1 in relation to monocyte/macrophage differentiation, especially in IR and the atherosclerotic process [105, 106]. It is well known that macrophages incorporate oxidized LDL (oxLDL) via the CD36 scavenger receptor pathway, thereby becoming ‘foam cells’, which are the hallmark of the early fatty streak lesion. Glucose-induced upregulation of CD36 in microvascular smooth muscle cells, followed by increased molecular markers of oxidative stress/damage, as well the increased concentrations of stress response proteins initiates vascular endothelial dysfunction [107, 108]. Elevated levels of CHI3L1 have been detected in the supernatant of macrophages after treatment with oxLDL [109], confirming a role of CHI3L1 in the differentiation of monocytes to ‘foam cells’ during formation of the atherosclerotic plaque. The elevated serum levels of CHI3L1 in patients with T2DM correlated positively with dyslipidaemia [104]. Previously, it was showed that CHI3L1 expression is highly upregulated in distinct subsets of macrophages in the atherosclerotic plaque that is characterized by the infiltration of monocytes into the sub-endothelial space of the vessel wall and the subsequent lipid accumulation of the activated macrophages [110]. Macrophages infiltrating deeply into the lesion especially show high CHI3L1 expression and, in particular, the highest expression was found in macrophages in the early lesion of atherosclerosis [104]. Monocytes from diabetic patients have significantly higher levels of CD36, CD14 and CD18 expression [111, 112]. CD14 is a key molecule in the innate immune response, where activation of membrane receptors initiates the secretion of pro-inflammatory cytokines and leads to clustering with other receptors involved in atherogenesis, such as CD11b/CD18 and scavenger receptor CD36 [113]. The differentiation and maturation of CD14+ monocytes to CD16+ macrophages is attended by the expression of CHI3L1 [114] (fig. 2). Remarkably, recent ev-

idence indicates a correlation between CHI3L1 and adhesion molecules [115]. In proximity of the CHI3L1 encoding gene, locus 1q32.2 influences the plasma levels of ICAM-1 [116], an adhesion molecule involved in endothelial dysfunction and the development of atherosclerosis. The production of the adhesion molecules VCAM-1, ICAM-1 and E-selectin, which all play a role in endothelial dysfunction preceding atherosclerosis, are stimulated by cytokine mediators such as IL-6 [117]. VCAM-1 and E-selectin also correlate with IR [118]. Patients with T2DM have elevated levels of CRP, IL-6 and the cell adhesion molecules ICAM-1, VCAM-1 and E-selectin [119]. Therefore, elevated levels of CHI3L1 in patients with DM play a greater role in the process of atherosclerosis.

Vascular endothelial growth factor (VEGF) in response to hypoxia is principally regulated by HIF-1 $\alpha$  (hypoxia-inducible factor-1 $\alpha$ ) and plays a key role in angiogenesis and endothelial function [104]. Plasma levels of CHI3L1 correlated positively with sFlt-1/VEGF, suggesting that inhibition of VEGF by circulating sFlt-1 may play a role in the upregulation of CHI3L1 [120]. It is possible that the levels of VEGF may be rate limiting for CHI3L1 regulation, possibly constituting a negative feedback loop. Raised serum CHI3L1 levels have been found in patients with stable CAD, providing prognostic information in these patients [121]. Furthermore, high CHI3L1 levels have been detected during acute myocardial infarction [104] and in patients with carotid plaques, and, in particular, the highest levels were found in patients with symptomatic lesions. Notably, the association between CHI3L1 and symptomatic disease was also found in patients with the most recent symptoms [104]. This observation strongly indicated that CHI3L1 might play a role in plaque stability. Platelets are important mediators of inflammation, at least partly related to their ability to induce monocyte activation [122]. An inflammatory interaction between platelets and monocytes also includes the ability of platelets to promote CHI3L1 expression, as demonstrated by the finding that the  $\beta$ -receptor agonist isoproterenol activated platelets markedly increasing CHI3L1 expression in THP-1 cells [104]. Patients with atherosclerotic disorders, and in particular those with unstable disease, are characterized by enhanced  $\beta$ -adrenergic receptor stimulation and platelet activation [104], which promotes CHI3L1 release within the lesion, further linking CHI3L1 to the unstable patient phenotype. Therefore, CHI3L1 might also be a marker of plaque instability, potentially reflecting macrophage activation and matrix degradation within the atherosclerotic lesion. Overall, the findings regarding tissue distribution and function of

CHI3L1 and the concept that CHI3L1 is an inflammatory marker strongly indicate that CHI3L1 plays more than a crucial part in atherosclerotic conditions, such as cardiovascular diseases, and in the pathogenesis of diabetic complications. Future studies should further investigate the potential pro-atherogenic effects of CHI3L1 as well as the utility of CHI3L1 as a risk predictor in patients with atherosclerosis.

### CHI3L1 and Diabetic Nephropathy

Diabetic nephropathy is one of the most important complications as a significant cause of end-stage renal disease with high morbidity and mortality. Although clear evidence indicates that the pathogenesis of diabetic nephropathy is multifactorial, inflammation is a key pathophysiological mechanism [123]. The mediators related to the dysfunction and inflammatory pathways are elevated in patients with DM as well as with microvascular complications, and are important in the early stages of diabetic kidney disease. An increasing urinary albumin excretion rate reflects vascular damage in the kidneys as part of systemic endothelial dysfunction [124]. Both micro- and macroalbuminuria are accompanied by increased levels of a variety of markers of endothelial dysfunction [125]. Persistent micro- and macroalbuminuria are well-known predictors of diabetic nephropathy leading to progressive renal insufficiency and end-stage renal disease, and are associated with an increased risk of cardiovascular disease in patients with both type 1 and type 2 diabetes. As a low-molecular-weight protein, CHI3L1 is excreted by the kidney [126], filtered into the early tubular fluid and catabolized in the proximal tubules [127]. Other studies have shown that the urinary excretion of CHI3L1 increases significantly in macroalbuminuric diabetic patients. Nevertheless, it remains unclear whether the cause of enhanced levels of urinary CHI3L1 in macroalbuminuria arises from the tubular damage that causes the reduced tubular reabsorption of filtered CHI3L1 or from increased renal production of CHI3L1, which, in turn, is induced by localized inflammation. Chronic low-grade inflammation is associated with the occurrence and progression of albuminuria [128]. An independent association between high levels of CHI3L1 and albuminuria, which represents an early marker of vascular complications, was detected both in type 1 and type 2 DM patients [81, 126]. These data suggest that CHI3L1 may be considered a potential indicator for risk assessment at least in adults. Since the kidneys excrete CHI3L1, it was not sur-



prising to find a significant correlation between CHI3L1 and the UACR (urinary albumin/creatinine ratio) [84]. The elevated CHI3L1 levels were found to be independently associated with the level of microalbuminuria after adjustment for UACR, age and other significant covariates, and the presence of retinopathy and intermittent claudication. It has been reported that CRP, as a marker of low-grade inflammation, was a determinant of the development of elevated excretion of urinary albumin [129]. However, it was previously demonstrated that both plasma and urine CHI3L1 are not significantly correlated with CRP [84]. In contrast to CRP, which is primarily produced by hepatocytes in response to IL-6 [130], CHI3L1 is produced locally at sites of inflammation [10]. Thus, CHI3L1 has a role as a site-specific inflammatory marker rather than as a systemic inflammatory marker such as CRP. Of note, the CHI3L1 level fulfils prognostic importance in normoalbuminuric patients with preserved kidney function in line with the concept that CHI3L1 acts as an early marker of diabetic nephropathy. Additional evidence demonstrating that CHI3L1 plays an important role in the development of early diabetic nephropathy with conserved eGFR suggests that plasma CHI3L1 measurement might become a useful and non-invasive tool for early incipient diabetic nephropathy as well as for the evaluation of the renal involvement of T2DM patients. Nevertheless, to support the role of CHI3L1 as a pro-inflammatory marker in the early development of diabetic nephropathy, further investigation is needed to determine the correlations with other inflammatory markers, such as TNF- $\alpha$ , interleukins and chemokines.

### Diabetic Retinopathy

Diabetic retinopathy (DR) is a microvascular complication of diabetes. Clinically characterized by retinal vascular microaneurysm and blot haemorrhages, DR can be categorized into early non-proliferative DR (mild NPDR), moderate and severe, or pre-proliferative DR and proliferative DR (PDR) [131]. The middle stages include moderate, severe and very severe non-proliferative diabetic retinopathies, generally with hard exudates and maculopathy. In this stage, venous changes, retinal capillary loss, retinal ischaemia, cotton wool or soft exudates, dot, blot spots and extensive intraretinal haemorrhages are evident. The PDR is the disease at the advanced stage characterized by neovascularization, preretinal and vitreous haemorrhages, fibrovascular proliferation and retinal de-

tachments. PDR is one of the major causes of vision loss in subjects with diabetes [131]. Subjects with T1DM have a higher prevalence of PDR than those with T2DM and key risk factors for the development of PDR include hyperglycaemia, diabetes duration and hypertension [131]. Altered epigenetic patterns may contribute to disease development and differential DNA methylation has been found in subjects with T1DM and T2DM compared with non-diabetic controls [132, 133]. Currently, it is supposed that transient peaks of hyperglycaemia might be an independent risk factor for the progression of retinopathy and that hyperglycaemic peaks may cause persistent epigenetic changes in spite of normoglycaemia [134]. Epigenetic modifications may also influence the development of vascular complications in diabetic subjects and recent evidence demonstrates that differential DNA methylation can be associated with DR [135]. Recently, it was found that serum levels of CHI3L1 were elevated in subjects with diabetic retinopathy [20]. Additionally, a decreased DNA methylation of CHI3L1 was observed in a subject affected by PDR, which may contribute to the increased expression of this gene [136]. Previous data showed that there was statistical significance in the levels of plasma concentration of CHI3L1 among two groups of patients with T2DM and different forms of DR [137]. The levels of CHI3L1 in peripheral blood samples have been found to also be elevated in the group with mild NPDR [137]. Moreover, correlation analysis revealed that the outer diameter retinal blood vessels positively correlated with CHI3L1 levels in all zones and in both eyes, and negatively correlated with the number of retinal vessels [137] (fig. 2).

### CHI3L1 and Skeletal Fragility

The clinical literature reporting the influence of DM on osteoarthritis (OA) and its therapeutic consequences suggests that DM may increase the risk of the development and severity of OA. Articular cartilage defects cause pain, reduced joint function and significant disability [138]. OA and DM frequently co-exist simply by chance due to their high prevalence and shared risk factors. Nearly half of patients with DM have some form of arthritis. The development of OA may also complicate DM. Increasing evidence indicates that OA adds to the burden of cardiovascular disease, which is higher than average in DM patients [139]. CHI3L1 has been linked to joint injury [140] and significantly elevated levels of CHI3L1 protein have been detected in serum and synovial fluid

from OA patients [25] (fig. 2). In synovial cells CHI3L1 mediates a mitogenic effect through the initiation of MAPK and PI3K signalling pathways by phosphorylation of ERK1/ERK2 (extracellular signal-regulated kinase-1 and -2) and AKT, respectively. Both pathways are required for the cells to complete mitosis and the activation of these pathways stimulates the growth of connective tissue cells [141]. In chondrocytes, CHI3L1 reduces the activation of p38 and SAPK/JNK MAPKs, which counteracts the inflammatory responses to TNF- $\alpha$  and IL-1. This leads to a reduced production of MMPs and IL-8. The modulation of p38 and SAPK/JNK by CHI3L1 is mediated through PI3K [25], and the induction and continued secretion of CHI3L1 requires sustained activation of NF- $\kappa$ B [24]. It has been reported that CHI3L1 interferes with the synthesis of hyaluronan, one of the most widespread and abundant glycosaminoglycan in humans, but no degenerative activity of CHI3L1 against hyaluronan has yet been demonstrated [142]. CHI3L1 is a growth factor for fibroblasts, chondrocytes and human synovial cells [143], and the promotion of growth and proliferation occurs in a dose-dependent manner in a concentration range similar to the effective dose of IGF-1. The regulation of CHI3L1 is scantily evaluated. The secretion of CHI3L1 by human chondrocytes in vitro is not influenced by IGF-1, while transforming growth factor- $\beta$  reduces the release of CHI3L1 to barely detectable levels [22]. CHI3L1 and IGF-1 work in a synergistic fashion when present in suboptimal concentrations [25]. CHI3L1 was expressed in an osteoarthritic rat cartilage model [140]. These findings suggested that patients with elevated serum levels of CHI3L1 may have a more increased osteolytic activity and a faster progression of the disease. Indeed, silencing CHI3L1 with siRNA resulted in a significant decrease in bone resorption activity and transfection with CHI3L1 siRNA decreased the levels of the pro-differentiation marker MMP9 [16]. The plethora of evidence showing that CHI3L1 stimulates the proliferation of connective tissue cells and modulates expression levels of chemokines and metalloproteases in inflammatory fibroblasts, and that this enhances the chemotaxis of endothelial cells [24], strongly indicates that CHI3L1 plays a crucial role in stromal cells not only in inflammatory conditions. CHI3L1 is expressed in osteophyte and diseased human osteoarthritic cartilage, but not in non-diseased cartilage, and its distribution within the tissue changes as the disease progresses [14]. The number of chondrocytes with a positive staining for both CHI3L1 and CHIT1 was weak or absent in normal cartilage, while the expression for CHI3L1 was very strong in osteoarthritic cartilage with anterior cruci-

ate ligament transection. Their production is closely related to an inflammatory process and pro-inflammatory cytokines [144]. Immunohistochemical analysis confirmed that CHI3L1 staining was found in chondrocytes of osteoarthritic cartilage mainly in the superficial and middle zone of the cartilage rather than the deep zone. There was a tendency for a high number of positive chondrocytes in areas of the femoral condyles with a considerable biomechanical load. CHI3L1 presents CBM (carbohydrate-binding motif), followed by the ability to bind carbohydrates, in particular glycosaminoglycan. CHI3L1 could activate pro-inflammatory cytokines through its CBM. This ability might explain the altered levels of these two molecules during a chronic inflammatory process such as OA. Recent articles confirm that the gelatinases influence OA onset and progression, regulating subchondral bone remodelling. In particular, a predominant role of MMP-9 has emerged during last year. Among various MMPs, the total MMP-9 level is positively correlated with the total MMP-13 level in OA [145], and it has been hypothesized that this gelatinase might be involved in the activation of pro-MMP-13 through yet unknown mechanisms. Notably, MMP-13 has long been considered the major enzyme involved in cartilage erosion during OA, thus MMP-9 might play a role, at least co-operatively, in joint degradation. Therefore, it is reasonable that the chitinase may be involved in the resorption of articular cartilage in OA through different pathways. The present finding indicates that CHI3L1 could play an important role in cartilage remodelling/degradation of osteoarthritic joints. CHI3L1 may emerge as a useful marker for OA and tissue degeneration.

## Conclusion

CHI3L1 has emerged as a new molecule that may contribute to numerous aspects and phases of diabetes and its complications, including the impaired endothelial dysfunction wound healing and cartilage remodelling/degradation observed in DM (fig. 2). It has become clear that this enzyme may provide valuable information within a clinical setting, potentially acting as a screening tool for high-risk patients and becoming an early predictive diagnostic tool, while informing the treatment decision-making process. Many of the results obtained so far on the involvement of endothelial dysfunction and cartilage remodelling/degradation in diabetic complications have been the result of large screening studies, and have been validated in in vitro cell systems and in in vivo settings.

CHI3L1 accumulating in diabetic tissues could affect biomechanics. Most importantly, we still lack critical knowledge of the pathways involved in CHI3L1 expression. Since CHI3L1 exerts different functions according to the tissues and conditions where it is expressed, we are only seeing a small fraction of the 'big picture'. Maintaining

CHI3L1 equilibrium is mandatory to prevent or delay DM complications. The gaining of more understanding into the mechanisms by which CHI3L1 is regulated will provide new tools with which to avoid risk factors eliciting multifactorial diseases such as DM and its complications.

## References

- Reddy MA, Zhang E, Natarajan R: Epigenetic mechanisms in diabetic complications and metabolic memory. *Diabetologia* 2015;58:443–455.
- Gooding JR, Jensen MV, Newgard CB: Metabolomics applied to the pancreatic islet. *Arch Biochem Biophys* 2016;589:120–130.
- McPhee JB, Schertzer JD: Immunometabolism of obesity and diabetes: microbiota link compartmentalized immunity in the gut to metabolic tissue inflammation. *Clin Sci (Lond)* 2015;129:1083–1096.
- Islam ST, Srinivasan S, Craig ME: Environmental determinants of type 1 diabetes: a role for overweight and insulin resistance. *J Paediatr Child Health* 2014;50:874–879.
- Lopategi A, López-Vicario C, Alcaraz-Quiles J, García-Alonso V, Rius B, Titos E, Clària J: Role of bioactive lipid mediators in obese adipose tissue inflammation and endocrine dysfunction. *Mol Cell Endocrinol* 2016;419:44–59.
- Tinkov AA, Sinitskii AI, Popova EV, Nemesheina ON, Gatiatulina ER, Skalnaya MG, Skalny AV, Nikonov AA: Alteration of local adipose tissue trace element homeostasis as a possible mechanism of obesity-related insulin resistance. *Med Hypotheses* 2015;85:343–347.
- Cao H: Adipocytokines in obesity and metabolic disease. *J Endocrinol* 2014;220:T47–T59.
- Rabinovitch A, Suarez-Pinzon WL: Cytokines and their roles in pancreatic islet  $\beta$ -cell destruction and insulin-dependent diabetes mellitus. *Biochem Pharmacol* 1998;55:1139–1149.
- Kyrgios I, Galli-Tsinopoulou A, Stylianou C, Papakonstantinou E, Arvanitidou M, Haidich AB: Elevated circulating levels of the serum acute-phase protein YKL-40 (chitinase 3-like protein 1) are a marker of obesity and insulin resistance in prepubertal children. *Metabolism* 2012;61:562–568.
- Żurawska-Plaksej E, Ługowska A, Hetmańczyk K, Knapik-Kordecka M, Piwowar A: Neutrophils as a source of chitinases and chitinase-like proteins in type 2 diabetes. *PLoS One* 2015;10:e0141730.
- Coffman FD: Chitinase 3-Like-1 (CHI3L1): a putative disease marker at the interface of proteomics and glycomics. *Crit Rev Clin Lab Sci* 2008;45:531–562.
- Johansen JS: Studies on serum YKL-40 as a biomarker in diseases with inflammation, tissue remodelling, fibrosis and cancer. *Dan Med Bull* 2006;53:172–209.
- Rehli M, Krause SW, Andreesen R: Molecular characterization of the gene for human cartilage gp-39 (CHI3L1), a member of the chitinase protein family and marker for late stages of macrophage differentiation. *Genomics* 1997;43:221–225.
- Kawada M, Hachiya Y, Arihiro A, Mizoguchi E: Role of mammalian chitinases in inflammatory conditions. *Keio J Med* 2007;56:21–27.
- Rehli M, Niller HH, Ammon C, Langmann S, Schwarzfischer L, Andreesen R, Krause SW: Transcriptional regulation of CHI3L1, a marker gene for late stages of macrophage differentiation. *J Biol Chem* 2003;278:44058–44067.
- Di Rosa M, Tibullo D, Vecchio M, Nunnari G, Saccone S, Di Raimondo F, Malaguarnera L: Determination of chitinases family during osteoclastogenesis. *Bone* 2014;61:55–63.
- Libreros S, Garcia-Areas R, Shibata Y, Carrio R, Torroella-Kouri M, Iragavarapu-Charyulu V: Induction of proinflammatory mediators by CHI3L1 is reduced by chitin treatment: decreased tumor metastasis in a breast cancer model. *Int J Cancer* 2012;131:377–386.
- Choi J, Lee HW, Suk K: Plasma level of chitinase 3-like 1 protein increases in patients with early Alzheimer's disease. *J Neurol* 2011;258:2181–2185.
- Lee JH, Kim SS, Kim IJ, Song SH, Kim YK, In Kim J, Jeon YK, Kim BH, Kwak IS: Clinical implication of plasma and urine YKL-40, as a proinflammatory biomarker, on early stage of nephropathy in type 2 diabetic patients. *J Diabetes Complications* 2012;26:308–312.
- Lebensztejn DM, Skiba E, Werpachowska I, Sobaniec-Lotowska ME, Kaczmarek M: Serum level of YKL-40 does not predict advanced liver fibrosis in children with chronic hepatitis B. *Adv Med Sci* 2007;52:120–124.
- Vind I, Johansen JS, Price PA, Munkholm P: Serum YKL-40, a potential new marker of disease activity in patients with inflammatory bowel disease. *Scand J Gastroenterol* 2003;38:599–605.
- Wang Y, Ripa RS, Johansen JS, Gabrielsen A, Steinbruchel DA, Friis T, Bindlev L, Haack-Sørensen M, Jørgensen E, Kastrup J: YKL-40 a new biomarker in patients with acute coronary syndrome or stable coronary artery disease. *Scand Cardiovasc J* 2008;42:295–302.
- Rathcke CN, Raymond I, Kistorp C, Hildebrandt P, Faber J, Vestergaard H: Low grade inflammation as measured by levels of YKL-40: association with an increased overall and cardiovascular mortality rate in an elderly population. *Int J Cardiol* 2010;143:35–42.
- Ling H, Recklies AD: The chitinase 3-like protein human cartilage glycoprotein 39 inhibits cellular responses to the inflammatory cytokines interleukin-1 and tumour necrosis factor-alpha. *Biochem J* 2004;380:651–659.
- Recklies AD, White C, Ling H: The chitinase 3-like protein human cartilage glycoprotein 39 (HC-gp39) stimulates proliferation of human connective tissue cells and activates both extracellular signal-regulated kinase and protein kinase B-mediated signalling pathways. *Biochem J* 2002;365:119–126.
- Kim MN, Lee KE, Hong JY, Heo WI, Kim KW, Kim KE, Sohn MH: Involvement of the MAPK and PI3K pathways in chitinase 3-like 1-regulated hyperoxia-induced airway epithelial cell death. *Biochem Biophys Res Commun* 2012;421:790–796.
- Ngernyuan N, Francescone RA, Jearanai-koon P, Daduang J, Supoken A, Yan W, Shao R, Limpaboon T: Chitinase 3 like 1 is associated with tumor angiogenesis in cervical cancer. *Int J Biochem Cell Biol* 2014;51:45–52.
- Di Rosa M, Tibullo D, Saccone S, Distefano G, Basile MS, Di Raimondo F, Malaguarnera L: CHI3L1 nuclear localization in monocyte derived dendritic cells. *Immunobiology* 2016;22:347–356.
- Di Rosa M, Malaguarnera G, De Gregorio C, Drago F, Malaguarnera L: Evaluation of CHI3L-1 and CHIT-1 expression in differentiated and polarized macrophages. *Inflammation* 2013;36:482–492.
- Bigg HF, Wait R, Rowan AD, Cawston TE: The mammalian chitinase-like lectin, YKL-40, binds specifically to type I collagen and modulates the rate of type I collagen fibril formation. *J Biol Chem* 2006;281:21082–21095.
- Ehnes JA, Perren A, Eppler E, Ribaux P, Pospisilik JA, Maor-Cahn R, Gueripel X, Ellingsgaard H, Schneider MK, Biollaz G, Fontana A, Reinecke M, Homo-Delarche F, Donath MY: Increased number of islet-associated macrophages in type 2 diabetes. *Diabetes* 2007;56:2356–2370.

- 32 Zhou R, Tardivel A, Thorens B, Choi I, Tschopp J: Thioredoxin-interacting protein links oxidative stress to inflammasome activation. *Nat Immunol* 2010;11:136–140.
- 33 Ellingsgaard H, Hauselmann I, Schuler B, Habib AM, Baggio LL, Meier DT, Eppler E, Bouzakri K, Wueest S, Muller YD, Hansen AM, Reinecke M, Konrad D, Gassmann M, Reimann F, Halban PA, Gromada J, Drucker DJ, Gribble FM, Ehses JA, Donath MY: Interleukin-6 enhances insulin secretion by increasing glucagon-like peptide-1 secretion from L cells and alpha cells. *Nat Med* 2011;17:1481–1489.
- 34 Böni-Schnetzler M, Boller S, Debray S, Bouzakri K, Meier DT, Prazak R, Kerr-Conte J, Pattou F, Ehses JA, Schuit FC, Donath MY: Free fatty acids induce a proinflammatory response in islets via the abundantly expressed interleukin-1 receptor I. *Endocrinology* 2009;150:5218–5229.
- 35 Garay-Malpartida HM, Mourão RF, Mantovani M, Santos IA, Sagayar MC, Goldberg AC: Toll-like receptor 4 (TLR4) expression in human and murine pancreatic beta-cells affects cell viability and insulin homeostasis. *BMC Immunol* 2011;12:18.
- 36 Dasu M, Devaraj S, Zhao L, Hwang D, Jialal I: High glucose induces toll-like receptor expression in human monocytes: mechanism of activation. *Diabetes* 2008;57:3090–3098.
- 37 Ehses JA, Meier DT, Wueest S, Rytka J, Boller S, Wielinga PY, Schraenen A, Lemaire K, Debray S, van Lommel L, Pospisilik JA, Tschopp O, Schultze SM, Malipiero U, Esterbauer H, Ellingsgaard H, Rützi S, Schuit FC, Lutz TA, Böni-Schnetzler M, Konrad D, Donath MY: Toll-like receptor 2-deficient mice are protected from insulin resistance and beta cell dysfunction induced by a high-fat diet. *Diabetologia* 2010;53:1795–1806.
- 38 Poggi M, Bastelica D, Gual P, Iglesias MA, Gremeaux T, Knauf C, Peiretti F, Verdier M, Juhán-Vague I, Tanti JF, Burcelin R, Alessi MC: C3H/HeJ mice carrying a toll-like receptor 4 mutation are protected against the development of insulin resistance in white adipose tissue in response to a high-fat diet. *Diabetologia* 2007;50:1267–1276.
- 39 Tsukumo DM, Carvalho-Filho MA, Carvalheira JB, Prada PO, Hirabara SM, Schenka AA, Araújo EP, Vassallo J, Curi R, Velloso LA, Saad MJ: Loss-of-function mutation in Toll-like receptor 4 prevents diet-induced obesity and insulin resistance. *Diabetes* 2007;56:1986–1998.
- 40 Senn JJ: Toll-like receptor-2 is essential for the development of palmitate induced insulin resistance in myotubes. *J Biol Chem* 2006;281:26865–26875.
- 41 Alexander C, Rietschel ET: Bacterial lipopolysaccharides and innate immunity. *J Endotoxin Res* 2001;7:167–202.
- 42 Al-Attas OS, Al-Daghri NM, Al-Rubeaan K, da Silva NF, Sabico SL, Kumar S, McTernan PG, Harte AL: Changes in endotoxin levels in T2DM subjects on anti-diabetic therapies. *Cardiovasc Diabetol* 2009;8:20.
- 43 Mehta NN, McGillicuddy FC, Anderson PD, Hinkle CC, Shah R, Pruscino L, Tabita-Martinez J, Sellers KF, Rickels MR, Reilly MP: Experimental endotoxemia induces adipose inflammation and insulin resistance in humans. *Diabetes* 2010;59:172–181.
- 44 Pal D, Dasgupta S, Kundu R, Maitra S, Das G, Mukhopadhyay S, Ray S, Majumdar SS, Bhattacharya S: Fetuin-A acts as an endogenous ligand of TLR4 to promote lipid-induced insulin resistance. *Nat Med* 2012;18:1279–1285.
- 45 Kamba A, Lee IA, Mizoguchi E: Potential association between TLR4 and chitinase 3-like 1 (CHI3L1/YKL-40) signaling on colonic epithelial cells in inflammatory bowel disease and colitis-associated cancer. *Curr Mol Med* 2013;13:1110–1121.
- 46 Haglund L, Bernier SM, Onnerfjord P, Recklies AD: Proteomic analysis of the LPS-induced stress response in rat chondrocytes reveals induction of innate immune response components in articular cartilage. *Matrix Biol* 2008;27:107–118.
- 47 Müller-Anstett MA, Müller P, Albrecht T, Nega M, Wagener J, Gao Q, Kaesler S, Schaller M, Biedermann T, Götz F: Staphylococcal peptidoglycan co-localizes with Nod2 and TLR2 and activates innate immune response via both receptors in primary murine keratinocytes. *PLoS One* 2010;5:e13153.
- 48 Shiny A, Regin B, Balachandar V, Gokulakrishnan K, Mohan V, Babu S, Balasubramanyam M: Convergence of innate immunity and IR as evidenced by increased nucleotide oligomerization domain (NOD) expression and signaling in monocytes from patients with type 2 diabetes. *Cytokine* 2013;64:564–570.
- 49 Zhao L, Hu P, Zhou Y, Purohit J, Hwang D: Nod1 activation induces proinflammatory gene expression and insulin resistance in 3T3-L1 adipocytes. *Am J Physiol* 2011;301:587–598.
- 50 Tamrakar A, Schertzer J, Chiu T, Foley K, Bilan P, Philpott D, et al: Nod2 activation induces muscle cell-autonomous innate immune responses and insulin resistance. *Endocrinology* 2010;151:5624–5637.
- 51 Abbott DW, Andrew W, Asara MJ, Cantley CL: The Crohns disease protein, Nod2, requires Rip2 in order to induce ubiquitinylation of a novel site on nemo. *Curr Biol* 2004;14:2217–2227.
- 52 Park JH, Kim YG, McDonald C, Kanneganti TD, Hasegawa M, Body-Malapel M, Inohara N, Núñez G: RICK/RIP2 mediates innate immune responses induced through Nod1 and Nod2 but not TLRs. *J Immunol* 2007;178:2380–2386.
- 53 Esser N, Legrand-Poels S, Piette J, Scheen AJ, Paquot N: Inflammation as a link between obesity, metabolic syndrome and type 2 diabetes. *Diabetes Res Clin Pract* 2014;105:141–150.
- 54 Curat CA, Wegner V, Sengenès C, Miranville A, Tonus C, Busse R, Bouloumié A: Macrophages in human visceral adipose tissue: increased accumulation in obesity and a source of resistin and visfatin. *Diabetologia* 2006;49:744–747.
- 55 Johnson AM, Olefsky JM: The origins and drivers of insulin resistance. *Cell* 2013;152:673–684.
- 56 Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr: Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 2003;112:1796–1808.
- 57 Lackey DE, Olefsky JM: Regulation of metabolism by the innate immune system. *Nat Rev Endocrinol* 2016;12:15–28.
- 58 Olefsky JM, Glass CK: Macrophages, inflammation, and insulin resistance. *Annu Rev Physiol* 2010;72:219–246.
- 59 Kohlgruber A, Lynch L: Adipose tissue inflammation in the pathogenesis of type 2 diabetes. *Curr Diab Rep* 2015;15:92.
- 60 Lasselín J, Magne E, Beau C, Ledagueñel P, Dexpert S, Aubert A, Layé S, Capuron L: Adipose inflammation in obesity: relationship with circulating levels of inflammatory markers and association with surgery-induced weight loss. *J Clin Endocrinol Metab* 2014;99:E53–E61.
- 61 Bories G, Caiazzo R, Derudas B, Copin C, Raverdy V, Pigeyre M, Pattou F, Staels B, Chinnetti-Gbaguidi G: Impaired alternative macrophage differentiation of peripheral blood mononuclear cells from obese subjects. *Diab Vasc Dis Res* 2012;9:189–195.
- 62 Buđak L, Łabuzek K, Buđak RJ, Machnik G, Bołdys A, Okopień B: Exenatide (a GLP-1 agonist) improves the antioxidative potential of in vitro cultured human monocytes/macrophages. *Naunyn-Schmiedeberg's Arch Pharmacol* 2015;388:905–919.
- 63 Henriksbo BD, Lau TC, Cavallari JF, Denou E, Chi W, Lally JS, Crane JD, Duggan BM, Foley KP, Fullerton MD, Tarnopolsky MA, Steinberg GR, Schertzer JD: Fluvastatin causes NLRP3 inflammasome-mediated adipose insulin resistance. *Diabetes* 2014;63:3742–3747.
- 64 Pontis S, Ribeiro A, Sasso O, Piomelli D: Macrophage-derived lipid agonists of PPAR-α as intrinsic controllers of inflammation. *Crit Rev Biochem Mol Biol* 2016;51:7–14.
- 65 Dicker D, Salook MA, Marcovicic D, Djaldetti M, Bessler H: Role of peripheral blood mononuclear cells in the predisposition of obese individuals to inflammation and infection. *Obes Facts* 2013;6:146–151.
- 66 Catalán V, Gómez-Ambrosi J, Rodríguez A, Ramírez B, Valentí V, Moncada R, Silva C, Salvador J, Frühbeck G: Peripheral mononuclear blood cells contribute to the obesity-associated inflammatory state independently of glycemic status: involvement of the novel pro-inflammatory adipokines chemerin, chitinase-3-like protein 1, lipocalin-2 and osteopontin. *Genes Nutr* 2015;10:460.

- 67 Lancha A, Rodríguez A, Catalán V, Becerril S, Sáinz N, Ramírez B, Burrell MA, Salvador J, Frühbeck G, Gómez-Ambrosi J: Osteopontin deletion prevents the development of obesity and hepatic steatosis via impaired adipose tissue matrix remodeling and reduced inflammation and fibrosis in adipose tissue and liver in mice. *PLoS One* 2014;9:e98398.
- 68 Zhang J, Wu Y, Zhang Y, Leroith D, Bernlohr DA, Chen X: The role of lipocalin 2 in the regulation of inflammation in adipocytes and macrophages. *Mol Endocrinol* 2008;22:1416–1426.
- 69 Ahangari F, Sood A, Ma B, Takyar S, Schuyler M, Qualls C, Dela Cruz CS, Chupp GL, Lee CG, Elias JA: Chitinase 3-like-1 regulates both visceral fat accumulation and asthma-like Th2 inflammation. *Am J Respir Crit Care Med* 2015;191:746–757.
- 70 Kyrgios I, Galli-Tsinopoulou A, Stylianou C, Papakonstantinou E, Arvanitidou M, Haidich AB: Elevated circulating levels of the serum acute-phase protein YKL-40 (chitinase 3-like protein 1) are a marker of obesity and insulin resistance in prepubertal children. *Metabolism* 2012;61:562–568.
- 71 Dahlman I, Kaaman M, Olsson T, Tan GD, Bickerton AS, Wählén K, Andersson J, Nordström EA, Blomqvist L, Sjögren A, Forsgren M, Attersand A, Arner P: A unique role of monocyte chemoattractant protein 1 among chemokines in adipose tissue of obese subjects. *J Clin Endocrinol Metab* 2005;90:5834–5840.
- 72 Kyrgios I, Galli-Tsinopoulou A, Stylianou C: Ghrelin-leptin network influences serum chitinase 3-like protein 1 (YKL-40) levels in obese prepubertal children. *Regul Pept* 2013;183:69–73.
- 73 El-Mesallamy HO, Mostafa AM, Amin AI, El Demerdash E: The interplay of YKL-40 and leptin in type 2 diabetic obese patients. *Diabetes Res Clin Pract* 2011;93:e113–e116.
- 74 Marroquí L, Gonzalez A, Neco P, Caballero-Garrido E, Vieira E, Ripoll C, Nadal A, Quesada I: Role of leptin in the pancreatic  $\beta$ -cell: effects and signaling pathways. *J Mol Endocrinol* 2012;49:R9–R17.
- 75 Nayak M, Eekhoff ME, Peinhaupt M, Heinemann A, Desoye G, van Poppel MN: Cytokines and their association with insulin resistance in obese pregnant women with different levels of physical activity. *Cytokine* 2015;77:72–78.
- 76 Nielsen AR, Erikstrup C, Johansen JS, Fischer CP, Plomgaard P, Krogh-Madsen R, Taudorf S, Lindegaard B, Pedersen BK: Plasma YKL-40: a BMI-independent marker of type 2 diabetes. *Diabetes* 2008;57:3078–3082.
- 77 Rathcke CN, Johansen JS, Vestergaard H: YKL-40, a biomarker of inflammation, is elevated in patients with type 2 diabetes and is related to insulin resistance. *Inflamm Res* 2006;55:53–59.
- 78 Abate N: Obesity and cardiovascular disease: pathogenetic role of the metabolic syndrome and therapeutic implications. *J Diabetes Complications* 2000;14:154–174.
- 79 Downs CA, Faulkner MS: Toxic stress, inflammation and symptomatology of chronic complications in diabetes. *World J Diabetes* 2015;6:554–565.
- 80 Cabrera SM, Henschel AM, Hessner MJ: Innate inflammation in type 1 diabetes. *Transl Res* 2016;167:214–227.
- 81 Rathcke CN, Vestergaard H: YKL-40, a new inflammatory marker with relation to insulin resistance and with a role in endothelial dysfunction and atherosclerosis. *Inflamm Res* 2006;55:221–227.
- 82 Rathcke CN, Persson F, Tarnow L, Rossing P, Vestergaard H: YKL-40, a marker of inflammation and endothelial dysfunction, is elevated in patients with type 1 diabetes and increases with levels of albuminuria. *Diabetes Care* 2009;32:323–328.
- 83 Ford ES: Body mass index, diabetes, and C-reactive protein among US adults. *Diabetes Care* 1999;22:1971–1977.
- 84 Rathcke CN, Kistorp C, Raymond I, Hildebrandt P, Gustafsson F, Lip GY, Faber J, Vestergaard H: Plasma YKL-40 levels are elevated in patients with chronic heart failure. *Scand Cardiovasc J* 2010;44:92–99.
- 85 Sakamoto F, Katakami N, Kaneto H, Yasuda T, Takahara M, Miyashita K, Kuroda A, Matsuhisa M, Kosugi K, Shimomura I: Association of serum YKL-40 levels with urinary albumin excretion rate in young Japanese patients with type 1 diabetes mellitus. *Endocr J* 2013;60:73–79.
- 86 Aguilera E, Serra-Planas E, Granada ML, Pellicer S, Reverter JL, Alonso N, Soldevila B, Mauricio D, Puig-Domingo M: Relationship of YKL-40 and adiponectin and subclinical atherosclerosis in asymptomatic patients with type 1 diabetes mellitus from a European Mediterranean population. *Cardiovasc Diabetol* 2015;14:121.
- 87 Persson F, Rathcke CN, Gall MA, Parving HH, Vestergaard H, Rossing P: High YKL-40 levels predict mortality in patients with type 2 diabetes. *Diabetes Res Clin Pract* 2012;96:84–89.
- 88 Rathcke CN, Raymond I, Kistorp C, Hildebrandt P, Faber J, Vestergaard H: Low grade inflammation as measured by levels of YKL-40: association with an increased overall and cardiovascular mortality rate in an elderly population. *Int J Cardiol* 2010;143:35–42.
- 89 Kruit A, Grutters JC, Ruven HJ, van Moorsel CC, van den Bosch JM: A CHI3L1 gene polymorphism is associated with serum levels of YKL-40, a novel sarcoidosis marker. *Respir Med* 2007;101:1563–1571.
- 90 Dixon AL, Liang L, Moffatt MF, Chen W, Heath S, Wong KC, Taylor J, Burnett E, Gut I, Farrall M, Lathrop GM, Abecasis GR, Cookson WO: A genome-wide association study of global gene expression. *Nat Genet* 2007;39:1202–1207.
- 91 Ober C, Tan Z, Sun Y, Possick JD, Pan L, Nicolae R, Radford S, Parry RR, Heinzmann A, Deichmann KA, Lester LA, Gern JE, Lemanske RF Jr, Nicolae DL, Elias JA, Chupp GL: Effect of variation in CHI3L1 on serum YKL-40 level, risk of asthma, and lung function. *N Engl J Med* 2008;358:1682–1691.
- 92 Lee H, Kang R, Jee SH, Yoon Y: A promoter polymorphism -2122C>T of *CHI3L1* is associated with serum low density lipoprotein cholesterol level in Korean subjects. *Clin Biochem* 2010;43:1195–1200.
- 93 Rathcke CN, Holmkvist J, Jørgensen T, Borch-Johnsen K, Hansen T, Pedersen OB, Vestergaard H: Variation in CHI3LI in relation to type 2 diabetes and related quantitative traits. *PLoS One* 2009;4:e5469.
- 94 Batinic K, Höbaus C, Grujicic M, Steffan A, Jelic F, Lorant D, Hörtenhuber T, Hoellerl F, Brix JM, Scherthner G, Koppensteiner R, Scherthner GH: YKL-40 is elevated in patients with peripheral arterial disease and diabetes or pre-diabetes. *Atherosclerosis* 2012;222:557–563.
- 95 Hernandez-Mijares A, Rocha M, Rovira-Llopis S, Bañuls C, Bellod L, de Pablo C, Alvarez A, Roldan-Torres I, Sola-Izquierdo E, Victor VM: Human leukocyte/endothelial cell interactions and mitochondrial dysfunction in type 2 diabetic patients and their association with silent myocardial ischemia. *Diabetes Care* 2013;36:1695–1702.
- 96 Stratmann B, Tschöepe D: Atherogenesis and atherothrombosis – focus on diabetes mellitus. *Best Pract Res Clin Endocrinol Metab* 2009;23:291–303.
- 97 Zhang C, Zhang L, Chen S, Feng B, Lu X, Bai Y, Liang G, Tan Y, Shao M, Skibba M, Jin L, Li X, Chakrabarti S, Cai L: The prevention of diabetic cardiomyopathy by non-mitogenic acidic fibroblast growth factor is probably mediated by the suppression of oxidative stress and damage. *PLoS One* 2013;8:e82287.
- 98 Catalán V, Gómez-Ambrosi J, Rodríguez A, Ramírez B, Silva C, Rotellar F, Gil MJ, Cienfuegos JA, Salvador J, Frühbeck G: Increased adipose tissue expression of lipocalin-2 in obesity is related to inflammation and matrix metalloproteinase-2 and metalloproteinase-9 activities in humans. *J Mol Med (Berl)* 2009;87:803–813.
- 99 Permana PA, Menge C, Reaven PD: Macrophage-secreted factors induce adipocyte inflammation and insulin resistance. *Biochem Biophys Res Commun* 2006;341:507–514.
- 100 Nøjgaard C, Høst NB, Christensen IJ, Poulsen SH, Egstrup K, Price PA, Johansen JS: Serum levels of YKL-40 increases in patients with acute myocardial infarction. *Coron Artery Dis* 2008;19:257–263.
- 101 Ma WH, Wang XL, Du YM, Wang YB, Zhang Y, Wei DE, Guo LL, Bu PL: Association between human cartilage glycoprotein 39 (YKL-40) and arterial stiffness in essential hypertension. *BMC Cardiovasc Disord* 2012;12:35.

- 102 Bilim O, Takeishi Y, Kitahara T, Ishino M, Sasaki T, Suzuki S, Shishido T, Kubota I: Serum YKL-40 predicts adverse clinical outcomes in patients with chronic heart failure. *J Card Fail* 2010;16:873–879.
- 103 Gong Z, Xing S, Zheng F, Xing Q: Increased expression of chitinase 3-like 1 in aorta of patients with atherosclerosis and suppression of atherosclerosis in apolipoprotein E-knockout mice by chitinase 3-like 1 gene silencing. *Mediators Inflamm* 2014;2014:905463.
- 104 Michelsen AE, Rathcke CN, Skjelland M, Holm S, Ranheim T, Krohg-Sørensen K, Klingvall MF, Brosstad F, Oie E, Vestergaard H, Aukrust P, Halvorsen B: Increased YKL-40 expression in patients with carotid atherosclerosis. *Atherosclerosis* 2010;211:589–595.
- 105 Catalán V, Gómez-Ambrosi J, Rodríguez A, Ramírez B, Rotellar F, Valentí V, Silva C, Gil MJ, Salvador J, Frühbeck G: Increased circulating and visceral adipose tissue expression levels of YKL-40 in obesity-associated type 2 diabetes are related to inflammation: impact of conventional weight loss and gastric bypass. *J Clin Endocrinol Metab* 2011;96:200–209.
- 106 Tang H, Shi Z, Xiu Q, Li B, Sun Y: YKL-40-mediated interleukin 8 production may be closely associated with remodeling of bronchial smooth muscle cells. *Am J Respir Crit Care Med* 2012;186:386.
- 107 Farhangkhoe H, Khan ZA, Barbin Y, Chakrabarti S: Glucose induced up-regulation of CD36 mediates oxidative stress and microvascular endothelial cell dysfunction. *Diabetologia* 2005;48:1401–1410.
- 108 Boullier A, Bird DA, Chang MK, Dennis EA, Friedman P, Gillotre-Taylor K, Hörkkö S, Palinski W, Quehenberger O, Shaw P, Steinberg D, Terpstra V, Witztum JL: Scavenger receptors, oxidized LDL, and atherosclerosis. *Ann NY Acad Sci* 2001;947:214–222.
- 109 Thomsen SB, Rathcke CN, Skaaby T, Linneberg A, Vestergaard H: The Association between genetic variations of CHI3L1, levels of the encoded glycoprotein YKL-40 and the lipid profile in a Danish population. *PLoS One* 2012;7:e47094.
- 110 Boot RG, van Achtenberg TA, van Aken BE, Renkema GH, Jacobs MJ, Aerts JM, de Vries CJ: Strong induction of members of the chitinase family of proteins in atherosclerosis: chitotriosidase and human cartilage gp-39 expressed in lesion macrophages. *Arterioscler Thromb Vasc Biol* 1999;19:687–694.
- 111 Fogelstrand L, Hulthe J, Hulthen LM, Wiklund O, Fagerberg B: Monocytic expression of CD14 and CD18, circulating adhesion molecules and inflammatory markers in women with diabetes mellitus and impaired glucose tolerance. *Diabetologia* 2004;47:1948–1952.
- 112 Bernal-Lopez RM, Llorente-Cortes V, López-Carmona D, Mayas DM, Gomez-Huelgas R, Tinahones FJ, Badimon L: Modulation of human monocyte CD36 by type 2 diabetes mellitus and other atherosclerotic risk factors. *Eur J Clin Invest* 2011;41:854–862.
- 113 Pfeiffer A, Böttcher A, Orsó E, Kapinsky M, Nagy P, Bodnár A, Spreitzer I, Liebisch G, Drobniak W, Gempel K, Horn M, Holmer S, Hartung T, Multhoff G, Schütz G, Schindler H, Ulmer AJ, Heine H, Stelter F, Schütt C, Rothe G, Szöllösi J, Damjanovich S, Schmitz G: Lipopolysaccharide and ceramide docking to CD14 provokes ligand-specific receptor clustering in rafts. *Eur J Immunol* 2001;31:3153–3164.
- 114 Baeten D, Boots AM, Steenbakkers PG, Elewaut D, Bos E, Verheijden GF, Berheijden G, Miltenburg AM, Rijnders AW, Veys EM, De Keyser F: Human cartilage gp-39+, CD16+ monocytes in peripheral blood and synovium: correlation with joint destruction in rheumatoid arthritis. *Arthritis Rheum* 2000;43:1233–1243.
- 115 Otterdal K, Janardhanan J, Astrup E, Ueland T, Prakash JA, Lekva T, Abraham OC, Thomas K, Damås JK, Mathews P, Mathai D, Aukrust P, Varghese GM: Increased endothelial and macrophage markers are associated with disease severity and mortality in scrub typhus. *J Infect* 2014;69:462–469.
- 116 Dupuis J, Larson MG, Vasani RS, Massaro JM, Wilson PW, Lipinska I, Corey D, Vita JA, Keaney JF Jr, Benjamin EJ: Genome scan of systemic biomarkers of vascular inflammation in the Framingham Heart Study: evidence for susceptibility loci on 1q. *Atherosclerosis* 2005;182:307–314.
- 117 Rajas O, Ortega-Gómez M, Galván Román JM, Curbelo J, Fernández Jiménez G, Vega Piris L, Rodríguez Salvanes F, Arnalich B, Luquero Bueno S, Díaz López A, de la Fuente H, Suárez C, Ancochea J, Aspa J: The incidence of cardiovascular events after hospitalization due to CAP and their association with different inflammatory markers. *BMC Pulm Med* 2014;14:197.
- 118 Burns SF, Lee S, Bacha F, Tfayli H, Hannon TS, Arslanian SA: Pre-diabetes in overweight youth and early atherogenic risk. *Metabolism* 2014;63:1528–1535.
- 119 Gustafsson S, Lind L, Söderberg S, Zilmer M, Hulthe J, Ingelsson E: Oxidative stress and inflammatory markers in relation to circulating levels of adiponectin. *Obesity (Silver Spring)* 2013;21:1467–1473.
- 120 Jafari B, Elias JA, Mohsenin V: Increased plasma YKL-40/chitinase-3-like-protein-1 is associated with endothelial dysfunction in obstructive sleep apnea. *PLoS One* 2014;9:e98629.
- 121 Kastrup J, Johansen JS, Winkel P, Hansen JF, Hildebrandt P, Jensen GB, Jespersen CM, Kjølner E, Kolmos HJ, Lind I, Nielsen H, Gluud C; CLARICOR Trial Group: High serum YKL-40 concentration is associated with cardiovascular and all-cause mortality in patients with stable coronary artery disease. *Eur Heart J* 2009;30:1066–1072.
- 122 Weyrich AS, Elstad MR, McEver RP, McIntyre TM, Moore KL, Morrissey JH, Prescott SM, Zimmerman GA: Activated platelets signal chemokine synthesis by human monocytes. *J Clin Invest* 1996;97:1525–1534.
- 123 Navarro-González JF, Mora-Fernández C, Muros de Fuentes M, García-Pérez J: Inflammatory molecules and pathways in the pathogenesis of diabetic nephropathy. *Nat Rev Nephrol* 2011;7:327–340.
- 124 Deckert T, Feldt-Rasmussen B, Borch-Johnsen K, Jensen T, Kofoed-Enevoldsen A: Albuminuria reflects widespread vascular damage: the Steno hypothesis. *Diabetologia* 1989;32:219–226.
- 125 Mahdy RA, Nada WM, Hadhoud KM, El-Tarhony SA: The role of vascular endothelial growth factor in the progression of diabetic vascular complications. *Eye (Lond)* 2010;24:1576–1584.
- 126 Żurawska-Plaksej E, Ługowska A, Hetmańczyk K, Knapik-Kordecka M, Adamiec R, Piwowar A: Proteins from the 18 glycosyl hydrolase family are associated with kidney dysfunction in patients with diabetes type 2. *Biomarkers* 2015;20:52–57.
- 127 Matheson A, Willcox MD, Flanagan J, Walsh BJ: Urinary biomarkers involved in type 2 diabetes: a review. *Diabetes Metab Res Rev* 2010;26:150–171.
- 128 Gomes MB, Piccirillo LJ, Nogueira VG, Matos HJ: Acute-phase proteins among patients with type 1 diabetes. *Diabetes Metab* 2003;29:405–411.
- 129 Jager A, van Hinsbergh VW, Kostense PJ, Emeis JJ, Nijpels G, Dekker JM, Heine RJ, Bouter LM, Stehouwer CD: C-reactive protein and soluble vascular cell adhesion molecule-1 are associated with elevated urinary albumin excretion but do not explain its link with cardiovascular risk. *Arterioscler Thromb Vasc Biol* 2002;22:593–598.
- 130 Engler R: Acute-phase proteins in inflammation (in French). *C R Seances Soc Biol Fil* 1995;189:563–578.
- 131 Yau JW, Rogers SL, Kawasaki R, Lamoureux EL, Kowalski JW, Bek T, et al: Global prevalence and major risk factors of diabetic retinopathy. *Diabetes Care* 2012;35:556–564.
- 132 Rakyan VK, Beyan H, Down TA, Hawa MI, Maslau S, Aden D, Daunay A, Busato F, Mein CA, Manfras B, Dias KR, Bell CG, Tost J, Boehm BO, Beck S, Leslie RD: Identification of type 1 diabetes-associated DNA methylation variable positions that precede disease diagnosis. *PLoS Genet* 2011;7:e1002300.

- 133 Yang BT, Dayeh TA, Kirkpatrick CL, Taneera J, Kumar R, Groop L, Wollheim CB, Nitert MD, Ling C: Insulin promoter DNA methylation correlates negatively with insulin gene expression and positively with HbA<sub>1c</sub> levels in human pancreatic islets. *Diabetologia* 2011;54:360–367.
- 134 El-Osta A, Brasacchio D, Yao D, Poci A, Jones PL, Roeder RG, Cooper ME, Brownlee M: Transient high glucose causes persistent epigenetic changes and altered gene expression during subsequent normoglycemia. *J Exp Med* 2008;205:2409–2417.
- 135 Bell CG, Teschendorff AE, Rakyan VK, Maxwell AP, Beck S, Savage DA: Genome-wide DNA methylation analysis for diabetic nephropathy in type 1 diabetes mellitus. *BMC Med Genet* 2010;3:33.
- 136 Agardh E, Lundstig A, Perfiljev A, Volkov P, Freiburghaus T, Lindholm E, Rönn T, Agardh CD, Ling C: Genome-wide analysis of DNA methylation in subjects with type 1 diabetes identifies epigenetic modifications associated with proliferative diabetic retinopathy. *BMC Med* 2015;13:182.
- 137 Cekić S, Cvetković T, Jovanović I, Jovanović P, Pešić M, Stanković Babić G, Milenković S, Risimić D: C-reactive protein and chitinase 3-like protein 1 as biomarkers of spatial redistribution of retinal blood vessels on digital retinal photography in patients with diabetic retinopathy. *Bosn J Basic Med Sci* 2014; 14:177–184.
- 138 Johnstone B, Alini M, Cucchiari M, Dodge GR, Eglin D, Guilak F, Madry H, Mata A, Mauck RL, Semino CE, Stoddart MJ: Tissue engineering for articular cartilage repair – the state of the art. *Eur Cell Mater* 2013;25: 248–267.
- 139 Adriaanse MC, Drewes HW, van der Heide I, Struijs JN, Baan CA: The impact of comorbid chronic conditions on quality of life in type 2 diabetes patients. *Qual Life Res* 2016; 25:175–182.
- 140 Di Rosa M, Szychlinska MA, Tibullo D, Malaguarnera L, Musumeci G: Expression of CHI3L1 and CHIT1 in osteoarthritic rat cartilage model: a morphological study. *Eur J Histochem* 2014;58:2423.
- 141 Areshkov PO, Avdieiev SS, Balynska OV, Leroith D, Kavsan VM: Two closely related human members of chitinase-like family, CHI3L1 and CHI3L2, activate ERK1/2 in 293 and U373 cells but have the different influence on cell proliferation. *Int J Biol Sci* 2012;8:39–48.
- 142 Johansen JS, Olee T, Price PA, Hashimoto S, Ochs RL, Lotz M: Regulation of YKL-40 production by human articular chondrocytes. *Arthritis Rheum* 2001;44:826–837.
- 143 De Ceuninck F, Gauffillier S, Bonnaud A, Sabatini M, Lesur C, Pastoureau P: YKL-40 (cartilage gp-39) induces proliferative events in cultured chondrocytes and synoviocytes and increases glycosaminoglycan synthesis in chondrocytes. *Biochem Biophys Res Commun* 2001;285:926–931.
- 144 Chen CC, Llado V, Eurich K, Tran HT, Mizoguchi E: Carbohydrate-binding motif in chitinase 3-like 1 (CHI3L1/YKL-40) specifically activates Akt signaling pathway in colonic epithelial cells. *Clin Immunol* 2011; 140:268–275.
- 145 Connor JR, Dodds RA, Emery JG, Kirkpatrick RB, Rosenberg M, Gowen M: Human cartilage glycoprotein 39 (HC gp-39) mRNA expression in adult and fetal chondrocytes, osteoblasts and osteocytes by in-situ hybridization. *Osteoarthritis Cartilage* 2000;8:87–95.