



**Dopamine D<sub>3</sub> Receptor is Necessary for Ethanol Consumption:  
An Approach with Buspirone OPEN**

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## Dopamine D<sub>3</sub> receptor is necessary for ethanol consumption: an approach with buspirone

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**Key words:** ethanol, dopamine D<sub>3</sub> receptor, SB277011A, BDNF, ANA-12, buspirone.

**Abstract**

Mesolimbic dopamine (DA) controls drug and alcohol seeking behavior, but the role of specific DA receptor subtypes is unclear. We tested the hypothesis that D<sub>3</sub>R gene deletion or the D<sub>3</sub>R pharmacological blockade inhibits ethanol preference in mice. D<sub>3</sub>R deficient mice (D<sub>3</sub>R<sup>-/-</sup>) and their wild type (WT) littermates, treated or not with the D<sub>3</sub>R antagonists SB277011A and U99194A, were tested in a long-term free choice ethanol-drinking (two-bottle choice) and in a binge-like ethanol drinking paradigm (drinking in the dark, DID). The selectivity of the D<sub>3</sub>R antagonists was further assessed by molecular modeling. Ethanol intake was negligible in D<sub>3</sub>R<sup>-/-</sup> and robust in WT both in the two-bottle choice and DID paradigms. Treatment with D<sub>3</sub>R antagonists inhibited ethanol intake in WT but was ineffective in D<sub>3</sub>R<sup>-/-</sup> mice. Ethanol intake increased the expression of RACK1 and BDNF in both WT and D<sub>3</sub>R<sup>-/-</sup>; in WT there was also a robust overexpression of D<sub>3</sub>R. Thus, increased expression of D<sub>3</sub>R associated with activation of RACK1/BDNF seems to operate as a reinforcing mechanism in voluntary ethanol intake. Indeed, blockade of the BDNF pathway by the TrkB selective antagonist ANA-12 reversed chronic stable ethanol intake and strongly decreased the striatal expression of D<sub>3</sub>R. Finally, we evaluated buspirone, an approved drug for anxiety disorders endowed with D<sub>3</sub>R antagonist activity (confirmed by molecular modeling analysis), that resulted effective in inhibiting ethanol intake. Thus, DA signaling via D<sub>3</sub>R is essential for ethanol-related reward and consumption and may represent a novel therapeutic target for weaning.

## Introduction

The mesolimbic dopamine (DA) pathway mediates the rewarding effects of drugs of abuse (Bowers *et al*, 2010; Ikemoto and Bonci, 2013; Koob, 1992; Robbins and Everitt, 1996; Wise and Bozarth, 1987), including ethanol and opiates (Pierce and Kumaresan, 2006; Wise *et al*, 1987). Both oral self-administration (Weiss *et al*, 1992) and systemic administration of ethanol increase the firing rate of mesolimbic dopaminergic neurons (Gessa *et al*, 1985; Mereu *et al*, 1984) and stimulate extracellular DA release in the striatum and in the nucleus accumbens (Imperato and Di Chiara, 1986; Yoshimoto *et al*, 1992). In a recent meta-analysis on published data sets of *in vivo* microdialysis in rat brain, the acute administrations of ethanol appear to increase the level of monoamines, including DA, globally and independent of the brain sites up to 270% of the basal concentrations (Brand *et al*, 2013). DA exerts its action through five receptor subtypes (D<sub>1-5</sub>R); the D<sub>3</sub> receptor (D<sub>3</sub>R) subtype plays an important role in the modulation of the mesolimbic DA pathway and in the control of drug-seeking behavior (Heidbreder *et al*, 2005; Joyce and Millan, 2005). The D<sub>3</sub>R is located both at pre- and post-synapses, in the ventral striatum (nucleus accumbens and island of Calleja (Bouthenet *et al*, 1991; Murray *et al*, 1994)); in these structures, stimulation of presynaptic D<sub>3</sub>R may modulate DA synthesis and release (Levant, 1997). Several studies have explored the involvement of D<sub>3</sub>R in ethanol-drinking paradigms (Cohen *et al*, 1998; Harrison and Nobrega, 2009; Heidbreder *et al*, 2007; Rice *et al*, 2012; Silvestre *et al*, 1996; Thanos *et al*, 2005), but their precise role remains unclear. Indeed, pharmacological studies generally report that D<sub>3</sub>R blockade decreases ethanol consumption (Heidbreder *et al*, 2007; Rice *et al*, 2012; Silvestre *et al*, 1996; Vengeliene *et al*, 2006); in contrast, genetic manipulation studies did not find a change in ethanol intake following D<sub>3</sub>R gene deletion (McQuade *et al*, 2003).

In the present study we tested the hypothesis that D<sub>3</sub>R gene deletion or the D<sub>3</sub>R pharmacological blockade inhibits the ethanol preference and the voluntary intake in mice.

Mice  $D_3R^{-/-}$  and their wild type (WT) littermates, treated or not with  $D_3R$  selective antagonists, were tested in a long-term free choice ethanol-drinking paradigm (two-bottle choice) (McQuade *et al*, 2003; Wise, 1973) and in a binge-like ethanol drinking paradigm (drinking in the dark, DID). Activation of the RACK1/BDNF/ $D_3R$  pathway (Jeanblanc *et al*, 2006) and activation of DA transmission were assessed at the end of behavioral experiments. The RACK1/BDNF/ $D_3R$  pathway was here considered because  $D_3R$  expression is related to BDNF (Guillin *et al*, 2001; Le Foll *et al*, 2005b) and ethanol exposure is able to increase RACK1 translocation into the nucleus of neurons, which increases expression of BDNF (Jeanblanc *et al*, 2006; McGough *et al*, 2004). Finally, the effect of buspirone was evaluated in the drinking paradigms. Because buspirone is an already approved drug for anxiety disorders, endowed with  $D_3R$  antagonist activity, it may be easier to translate to the clinic practice.

## Materials and Methods

### *Animals*

Mice  $D_3R$  null ( $D_3R^{-/-}$ ) and WT littermates (males, 8–12 weeks old) were individually housed, with free access to chow and water (except in the ethanol drinking procedures), in an air-conditioned room, with a 12-h light–dark cycle. Mice  $D_3R^{-/-}$  were 10<sup>th</sup>–12<sup>th</sup> generation of congenic C57BL/6J mice, generated by a back crossing strategy (Accili *et al*, 1996). All experiments were carried out according to the Directive 2010/63/EU and to the Institutional Animal Care and Use Committee of the Catania University.

### *Two-bottle choice paradigm*

Mice  $D_3R^{-/-}$  (n=30) and WT (n=30) received 24 h free access to tap water and 10% ethanol solution (v/v), contained in 100 ml graduated tubes with stainless steel drinking spouts; the position of tubes was interchanged (left/right) every 24 h, to prevent acquisition of position bias. Ethanol and water intake was measured as daily consumption in grams. The experiments lasted 59 days. For the first 15 days (habituation period) animals received 24 h free access to two tubes containing only tap water (time 0 in Figure 1a). After the habituation period (from 15 to 59 day) 10% ethanol solution was available in one of the bottles.

In the forced alcohol-drinking procedure,  $D_3R^{-/-}$  (n=12) and WT (n=18) received for the first 15 days (habituation period) tap water only (time 0), followed (from 15 to 59 day) by 10% ethanol only.

### *Drinking in the dark paradigm (DID)*

The 4 hour version of the behavioral paradigm was used, as described by Rhodes *et al.* (2005) The procedure started 3 h after lights off in the animal room. Water bottles

were replaced with graduated tubes with stainless steel drinking spouts containing 20% (v/v) ethanol in tap water. This was done in home cages where animals were singly housed. (Rhodes *et al*, 2005) The ethanol tubes remained in place for 2 h. After the 2-h period, intakes were recorded, and the ethanol tubes were replaced with water tubes. This procedure was repeated on days 2 and 3. On day 4, the procedure was again repeated except that the ethanol tubes were left in place for 4 h, and intakes were recorded after 4 h.

### *Drugs and treatments*

Ethanol, U99194A maleate, SB277011A hydrochloride, buspirone hydrochloride, 8-OH-DPAT and ANA-12 were from Sigma (St Louis, MO). All drugs were dissolved in saline and intraperitoneally (i.p.) injected (in a volume of 10 ml/kg), except ANA-12 that was dissolved in 10% dimethyl sulfoxide. U99194A was used at 10 mg/kg (Harrison *et al*, 2009), SB277011A was used at 10 mg/kg (Song *et al*, 2012), buspirone was used in the range 0.1-10 mg/kg (Martin *et al*, 1992), 8-OH-DPAT was used at 1 mg/kg (Martin *et al*, 1992) and ANA-12 was used at 0.5 mg/kg (Cazorla *et al*, 2011).

In the two-bottle choice paradigm, after 30 days of voluntary alcohol drinking procedure,  $D_3R^{-/-}$  and WT were randomly allocated to the 8 experimental groups (n=6/10 per group): WT/vehicle, WT/U99194A, WT/SB277011A WT/buspirone,  $D_3R^{-/-}$ /vehicle,  $D_3R^{-/-}$ /U99194A,  $D_3R^{-/-}$ /SB277011A,  $D_3R^{-/-}$ /buspirone. Animals were i.p. injected once a day, for 14 consecutive days. On day 14, animals were sacrificed 1 hour after the last administration and brain tissues were taken. In another set of experiments, after 30 days of voluntary alcohol drinking procedure, mice were randomly allocated to 5 experimental groups (n=5/7 per group): WT naïve, WT/vehicle, WT/ANA-12,  $D_3R^{-/-}$ /vehicle,  $D_3R^{-/-}$ /ANA-12. Animals were i.p. injected once a day, for 4 consecutive days with the selective Trkb

antagonist ANA-12 at 0.5 mg/kg (Cazorla *et al*, 2011; Vassoler *et al*, 2013). On day 4, animals were sacrificed 1 hour after the last administration and brain tissues were taken.

In the DID paradigm, mice were allocated to 10 experimental groups (n=5/6 per group): WT naïve, D<sub>3</sub>R<sup>-/-</sup> naïve, WT/vehicle, D<sub>3</sub>R<sup>-/-</sup>/vehicle, WT/SB277011A, D<sub>3</sub>R<sup>-/-</sup>/SB277011A, WT/buspironone 0.1 mg/kg, WT/buspironone 1 mg/kg, WT/buspironone 3 mg/kg, WT/buspironone 10 mg/kg. In another set of experiments mice were allocated to 4 experimental groups (n=5/6 per group): WT/vehicle, WT/8-OH-DPAT, D<sub>3</sub>R<sup>-/-</sup>/vehicle, D<sub>3</sub>R<sup>-/-</sup>/8-OH-DPAT and they were tested in the DID paradigm. Animals were i.p. injected 1 hour before the behavioral procedure.

#### *8-OH-DPAT-induced-hypothermia*

Body temperature was measured intrarectally using a lubricated probe inserted ~2 cm and a digital thermometer (CEM advanced thermometer; DT-610B). Mice were moved to the behavioral room and two baseline temperature measurements were taken. After 10 min, animals received an i.p. injection of vehicle or 1 mg/kg 8-OH-DPAT or 3 mg/kg buspirone. The body temperature was recorded every 15 min for a total of 45 min.

#### *Analysis of mRNA expression by real-time quantitative RT-PCR*

Total RNA was isolated by TRIzol (Invitrogen, Carlsbad, CA). Single-stranded cDNA was synthesized with SuperScript III (Invitrogen), by priming with oligo-(dT)<sub>20</sub>. Aliquots of cDNA were amplified in parallel reactions with external standards at known amounts, using specific primer pairs for D<sub>3</sub>R, RACK1, BDNF and S18 ribosomal RNA (reference gene). Each PCR reaction (20 µl final volume) contained 0.5 µM primers, 1.6 mM Mg<sup>2+</sup>, and 1× Light Cycler-Fast Start DNA Master SYBR Green I (Roche Diagnostics, Indianapolis, IN). Amplifications were carried out in a Light Cycler 1.5 instrument (Roche Diagnostics). Quantification was obtained by the ΔCt comparative method.



*Western Blot Analysis*

Protein extracts from striatum and cerebellum were run in SDS-PAGE, blotted and probed for non-phosphorylated and phosphorylated forms of DARPP-32, GSK-3 $\beta$  and Trkb, with primary antibodies (Cell Signalling Technology, Beverly, MA), diluted at 1:1,000, and secondary antibody (goat anti-rabbit IRDye; Li-Cor Biosciences, Lincoln, NE). Blots were scanned with an Odyssey Infrared Imaging System (Li-Cor Biosciences) and analyzed with ImageJ software (NIH, Bethesda, MD; <http://rsb.info.nih.gov/ij/index.html>).

*Statistical analysis*

Data were analyzed using one- or two-way analysis of variance (ANOVA). The post hoc Newman-Keuls test was used for multiple comparisons; p values less than 0.05 were considered as significant.

## Results

### *D<sub>3</sub>R<sup>-/-</sup> mice exhibited lower ethanol intake*

As shown in Figure 1a,b WT mice exhibited a high intake of ethanol-containing solution. In contrast, D<sub>3</sub>R<sup>-/-</sup> mice showed a low ethanol intake (Figure 1a,b). During the entire period of observation (44 days) WT mice maintained their preferential intake of ethanol, whereas D<sub>3</sub>R<sup>-/-</sup> mice maintained a preferential intake of water ( $F_{(1,307)} = 1170,08$   $p < 0.001$ ). There was no difference between WT and D<sub>3</sub>R<sup>-/-</sup> in terms of total amount of fluid intake (ethanol + water) (Figure 1c). In the DID paradigm, D<sub>3</sub>R<sup>-/-</sup> mice also showed a lower ethanol intake compared to their WT counterparts ( $F_{(3,97)} = 13.90$   $p < 0.01$  2<sup>nd</sup> day;  $F_{(3,97)} = 21.04$   $p < 0.001$  3<sup>th</sup> day; Figure 2a).

### *Blockade of D<sub>3</sub>R inhibited ethanol intake*

In the two-bottle choice paradigm, after 30 days of stable ethanol/water intake, mice were treated with D<sub>3</sub>R antagonists (U99194A or SB277011A). As shown in Figure 1d,e, treatment of WT with each D<sub>3</sub>R antagonist decreased voluntary ethanol intake ( $F_{(2,56)} = 55.23$   $p < 0.01$ , for both U99194A and SB277011A). Treatment of D<sub>3</sub>R<sup>-/-</sup> with U99194A and SB277011A did not change ethanol intake (data not shown). Neither in WT nor in D<sub>3</sub>R<sup>-/-</sup> total fluid intake was affected by treatments (Figure 1f and data not shown). SB277011A also significantly decreased ethanol intake in WT mice tested in the DID ( $F_{(3,48)} = 8.67$   $p < 0.01$  1<sup>st</sup> day;  $p < 0.05$  2<sup>nd</sup> day; Figure 2b), while it did not change ethanol intake of D<sub>3</sub>R<sup>-/-</sup> in the DID paradigm (Figure 2c).

### *RACK1, BDNF and Dopamine D<sub>3</sub>R expression were increased in the striatum of WT mice following chronic ethanol intake*

BDNF induces D<sub>3</sub> receptor expression in the ventral striatum, both during

development and in adulthood (Guillin *et al*, 2001). RACK1, a mediator of chromatin remodeling, regulates in an exon-specific manner the expression of the BDNF gene (He *et al*, 2010) and the RACK1/BDNF pathway is activated upon exposure to ethanol (McGough *et al*, 2004). We therefore assessed D<sub>3</sub>R, BDNF and RACK1 mRNA expression in striatum of WT that had free access to either water only or to both water and ethanol. Figure 3a shows that chronic ethanol intake increased D<sub>3</sub>R mRNA expression in striatum ( $F_{(3,23)} = 170.4$ ,  $p < 0.05$ ). Long-term access to ethanol also increased BDNF (Figure 3b,  $F_{(7,47)} = 48.05$ ,  $p < 0.01$ ) and RACK1 (Figure 3c,  $F_{(7,47)} = 21.14$ ,  $p < 0.01$ ) mRNA in striatum of WT mice.

Long-term ethanol exposure appeared to be associated with BDNF/RACK1 overexpression, but interpretation of these data was made difficult by the different ethanol intake in the two genetic groups, as it was very high in WT and very low in D<sub>3</sub>R<sup>-/-</sup>. To address this issue, some WT and D<sub>3</sub>R<sup>-/-</sup> mice were subjected to forced ethanol intake, i.e. they had access to ethanol 10% solution only. As shown in Figure 3d and 3e, forced ethanol intake induced a significant overexpression of BDNF ( $F_{(7,47)} = 48.05$ ,  $p < 0.05$ ,  $p < 0.01$ ) and RACK1 ( $F_{(7,47)} = 21.14$ ,  $p < 0.05$ ,  $p < 0.05$ ) mRNAs in striatum of both WT and D<sub>3</sub>R<sup>-/-</sup> mice. We also tested the effects of the D<sub>3</sub>R antagonists SB277011A and buspirone (see also below) on mRNA expression of D<sub>3</sub>R, BDNF and RACK1. None of these values were changed by a 14-day treatment with SB277011A or buspirone (Figure 3f-h).

#### *Blockade of the BDNF receptor TrkB inhibited ethanol intake and decreased D<sub>3</sub>R expression*

TrkB is the high affinity receptor for BDNF, belonging to the family of tyrosine kinase receptors, that undergo autophosphorylation upon agonist binding (Soppet *et al*, 1991). In order to assess the role of BDNF pathway in ethanol intake we used the recently available TrkB selective antagonist ANA-12 (Cazorla *et al*, 2011). After 30 days of stable

ethanol/water intake, mice received daily i.p. injections of either vehicle or ANA-12 (Figure 4a,b). ANA-12 reversed the stable ethanol intake of WT mice ( $F_{(7,42)} = 30.53$ ,  $p < 0.001$ ) but did not change the voluntary and the forced ethanol intake of  $D_3R^{-/-}$  (data not shown). Neither in WT nor in  $D_3R^{-/-}$  total fluid intake was affected by treatment with ANA-12 (Figure 4c and data not shown). Also in the DID paradigm ANA-12 was effective in reducing ethanol intake in WT mice ( $F_{(3,55)} = 6.64$ ,  $P < 0.05$ , Figure 4d), whereas it did not change ethanol intake in  $D_3R^{-/-}$  (Figure 4e).

To assess the selective blockade of the BDNF receptor in striatum by ANA-12, we determined, by immunoblot, the abundance of phosphorylated TrkB. As shown in Figure 4f, treatment of WT with ANA-12 significantly decreased phosphorylation of TrkB ( $F_{(3,35)} = 184.5$ ,  $p < 0.01$ ). Finally and more interestingly, ANA-12 strongly decreased  $D_3R$  mRNA expression in the striatum of WT mice exposed to voluntary ethanol intake (Figure 4f,  $F_{(3,35)} = 184.5$   $P < 0.001$ ).

#### *Buspironone inhibited ethanol intake*

In the two-bottle choice paradigm, after 30 days of stable ethanol/water intake, mice were treated with buspironone (1 mg/kg/day). As shown in Figure 5a,b, treatment of WT with buspironone significantly decreased voluntary ethanol intake ( $F_{(1,28)} = 20.88$   $p < 0.05$ ). Treatment of  $D_3R^{-/-}$  with buspironone did not change ethanol intake (data not shown). Neither in WT nor in  $D_3R^{-/-}$  total fluid intake was affected by treatment (Figure 5c and data not shown). The treatment with buspironone also significantly decreased ethanol intake in WT mice when tested in the DID. Dose ranging of buspironone (0.1, 1, 3 and 10 mg/kg) showed that treatment of WT with buspironone at the doses of 3 and 10 mg/kg significantly decreased ethanol intake both in the 1<sup>st</sup> day ( $F_{(4,75)} = 31.24$ ,  $p < 0.05$ ) and in the 2<sup>nd</sup> day ( $F_{(4,75)} = 31.24$ ,  $p < 0.01$  3 mg/kg;  $p < 0.05$  10 mg/kg) of the behavioral paradigm (Figure 5d). Buspironone did not change ethanol intake of WT in the 3<sup>rd</sup> and 4<sup>th</sup> days of DID (Figure 5d).

Furthermore, in the DID paradigm, 3 mg/kg buspirone did not change ethanol intake in  $D_3R^{-/-}$  (data not shown). Because buspirone is also known as a 5-HT<sub>1A</sub> agonist, the  $D_3R$  specific effect of buspirone in decreasing ethanol intake was confirmed by using the selective 5-HT<sub>1A</sub> agonist, 8-OH-DPAT. As shown in Figure 5e, treatment with 8-OH-DPAT (1mg/kg, i.p.) in WT and  $D_3R^{-/-}$  mice did not affect ethanol intake (Figure 5e and data not shown). As expected, the 5-HT<sub>1A</sub> selective agonist 8-OH-DPAT decreased the body temperature of WT mice ( $F_{(2,39)} = 14.99$ ,  $p < 0.001$ ) (Figure 5f). Buspirone (3 mg/kg) decreased the body temperature of WT mice only transiently (Figure 5f).

#### *Dopamine receptor signaling in striatum of WT and $D_3R^{-/-}$ mice exposed to ethanol*

Activation of  $D_1$  receptor results in activation of adenylyl cyclase/cAMP/protein kinase A (PKA) signaling; a major substrate for PKA in the striatum is DARPP-32.  $D_2$ -like receptors regulate the activity of the protein kinases Akt and GSK3 $\beta$ ; stimulation of either  $D_2$  or  $D_3$  receptors results in phosphorylation of Akt and GSK3 $\beta$  (Mannoury la Cour *et al*, 2011). In order to assess activation of dopaminergic transmission in striatum, we determined, by immunoblot, the abundance of phosphorylated DARPP-32 (Thr 34) and of phosphorylated GSK3 $\beta$  (Ser 9). As shown in Figure 66, posphoGSK3 $\beta$  was more abundant in striatum of  $D_3R^{-/-}$  than in WT mice, while phosphoDARPP-32 showed the same tendency, though it did not reach statistical significance. Treatment of WT mice with SB277011A induced phosphorylation of DARPP-32 and GSK3 $\beta$ , up to the level seen in  $D_3R^{-/-}$  mice. In contrast, in cerebellum there was no difference in the level phosphoDARPP-32 and posphoGSK3 $\beta$  between WT e  $D_3R^{-/-}$ , nor it was influenced by SB277011A-treatment in WT.

## Discussion

This study demonstrates that D<sub>3</sub>R is necessary for ethanol consumption in mice, because either D<sub>3</sub>R gene deletion or D<sub>3</sub>R pharmacological blockade by selective D<sub>3</sub>R experimental antagonists or the approved drug buspirone, inhibits alcohol intake. The D<sub>3</sub>R overexpression induced by ethanol intake associated with the activation of RACK1/BDNF may represent the basis for a reinforcing mechanism of ethanol intake. Indeed, while selective blockade of the TrkB reversed stable intake of ethanol in WT mice and decreased D<sub>3</sub>R expression levels in their striatum, it was ineffective in D<sub>3</sub><sup>-/-</sup> mice.

It seems that D<sub>3</sub>R, among D<sub>2</sub>-like receptors, is the key player in addiction, particularly in reward mechanisms. Indeed, while the D<sub>2</sub>R is associated with mesocortical and mesohippocampal DA pathway, the D<sub>3</sub>R is associated with the ventral mesolimbic DA system (Sokoloff *et al*, 1990). Previous studies reported low levels of D<sub>2</sub>R both in animal models and in patients addicted to cocaine, alcohol, metamphetamaine, and nicotine (Volkow *et al*, 2009). Conversely, up regulation of D<sub>3</sub>R expression has been reported following exposure to DA elevating drugs (Boileau *et al*, 2012; Heidbreder and Newman, 2010; Le Foll *et al*, 2005b; Mash, 1997; Segal *et al*, 1997; Staley and Mash, 1996).

An important interpretative issue is the genetic background on which the D<sub>3</sub>R null mutation was placed. Specific behavioral phenotypes are differently expressed in different strains of mice (Nelson and Young, 1998). The D<sub>3</sub>R<sup>-/-</sup> mice we used are on the C57BL/6J background (Accili *et al*, 1996), a strain where ethanol preference and sensitivity is well documented (Crabbe *et al*, 1983). Interestingly, D<sub>3</sub>R<sup>-/-</sup> mice have extracellular DA levels twice as high as their WT littermates (Joseph *et al*, 2002; Koeltzow *et al*, 1998); this enhanced DA tone and the resulting adaptations may reflect removal of the inhibitory influence of D<sub>3</sub>R in the control of basal extracellular DA levels (Le Foll *et al*, 2005a), giving support to an autoreceptor role for D<sub>3</sub>R in the mesolimbic areas of the brain (Diaz *et al*, 2000). The increased DA activity in D<sub>3</sub>R<sup>-/-</sup> mice is consistent with their phenotype, including

higher basal levels of grooming behavior, hyper-locomotion and reactivity to drug-paired environmental cues (Accili *et al*, 1996; Le Foll *et al*, 2005a; Le Foll *et al*, 2002).

Here we found that  $D_3R^{-/-}$  mice chronically exposed to the voluntary ethanol intake paradigm, drink very low quantities of ethanol in comparison with their WT littermates. This observation cannot be attributed to differences in metabolism (McQuade *et al*, 2003), locomotor activity (Harrison *et al*, 2009) or taste reactivity (McQuade *et al*, 2003) between WT and  $D_3R^{-/-}$  mice. The lower ethanol intake of  $D_3R^{-/-}$  in comparison with their WT control mice seems apparently in contrast with the only two previous studies testing  $D_3R^{-/-}$  mice in the ethanol voluntary intake paradigm (Boyce-Rustay and Risinger, 2003; McQuade *et al*, 2003). This may be due, at least in part, to some important differences in experimental procedures used. Indeed, McQuade *et al*. (2003), that have shown no difference between  $D_3R^{-/-}$  and WT in the 24-hours access paradigm, used a different experimental procedure in the two-bottle choice paradigm. First, they used just 4 days of adaptation period before ethanol exposure. Second, they tested both  $D_3R^{-/-}$  and WT animals with increasing concentrations of ethanol in subsequent 7-day steps. In the first step 3% ethanol, in the second step 6%, in the third step 10%, in the 4<sup>th</sup> 15% and finally, in the 5<sup>th</sup> 20% ethanol. Thus, the behavioral paradigm used by McQuade and coworkers is quite different from our paradigm. From our experience, for these mice it is important to have a long period of habituation in the two-bottle paradigm (15 days) before to start with the ethanol access procedure. It is likely that the progressive increase of the ethanol concentration every 7 days, may induce an adaptation to the ethanol that damps the difference between  $D_3R^{-/-}$  and WT mice. Furthermore, in the McQuade's study, the relative positions of the ethanol and water bottle were determined randomly each day, whereas in our experiments the position of tubes was interchanged (left/right) every 24 h, to prevent acquisition of position bias. The random change of bottles may expose a given animal to access the same solution (either ethanol or water) in the same position for two/three days consecutively,

which may interfere with the results of the experiment during a short period of observation (7 days).

In the study by Boyce-Rustay and Risinger (2003), C57 animals were used as control of  $D_3R^{-/-}$  mice. These experiments are not comparable to our experiments using WT littermates as controls. Moreover, again, in this study increasing concentrations of ethanol were used in 8-day steps (3% and 10%). Thus, (i) the behavioral procedure is different; (ii) an adaptation to ethanol may occur and damp the difference between genotypes.

To obtain pharmacological evidence for a functional role of  $D_3R$  in the control of voluntary ethanol intake we tested two  $D_3R$  antagonists, U99194A and SB277011A at doses reported to selectively target the  $D_3R$  (Carr *et al*, 2002; Reavill *et al*, 2000). Before administering these drugs, we performed a molecular modeling study to gain information on the interaction of U99194A and SB277011A with  $D_3R$ . As illustrated in Supplementary information, *in silico* analysis showed that the two  $D_3R$  antagonists were i) highly selective for the  $D_3R$  subtype and ii) displayed a distinct interaction (different binding energy, different interaction patterns) with  $D_3R$ , consistent with their distinct chemical structure. We found that both U99194A and SB277011A induced a significant decrease in voluntary ethanol intake in WT but not in  $D_3R^{-/-}$ . This pharmacological evidence reinforces the view that the  $D_3R$  is necessary for ethanol consumption in mice and is consistent with rat data showing that  $D_3R$  antagonism reduces relapse-like drinking and cue-induced ethanol-seeking behavior (Vengeliene *et al*, 2006).

We confirmed the primary role of  $D_3R$  in the control of ethanol drinking behavior in a binge-like ethanol drinking paradigm (Crabbe *et al*, 2011; Rhodes *et al*, 2005; Rhodes *et al*, 2007). Here, again,  $D_3R^{-/-}$  mice exposed to DID drank lower quantities of ethanol in comparison with their WT littermates, and  $D_3R$  blockade by SB277011A decreased ethanol intake in WT but not in  $D_3R^{-/-}$ . No differences were recorded in the DID at day 4.



Indeed, there was neither a genotype-effect between WT and  $D_3R^{-/-}$  nor a treatment-effect with the SB277011A in WT mice. In general, the binge-like behavior is captured by the 2h-time window, that detects differences between treatments/genotypes better than the 4h-window, because the cumulative intake over 4 h makes smaller the proportion of differences (Rhodes et al, 2005). Thus, it is likely that, the lack of differences on day 4 is due to the longer lasting access to ethanol that produced overall a higher consumption, potentially masking the genotype/treatment effect on binge-like drinking behavior occurring in the first 2 h.

Enhanced  $D_3R$  expression in striatum following long-term alcohol consumption has been previously reported in both mice and rats (Jeanblanc *et al*, 2006; Vengeliene *et al*, 2006). Our data show and confirm that chronic voluntary ethanol intake up-regulated  $D_3R$  mRNA expression in the striatum of WT mice. Interestingly,  $D_3R$  expression is increased by exposure to other addictive drugs, such as nicotine and cocaine, in caudate-putamen (Neisewander *et al*, 2004) and in nucleus accumbens of rats (Le Foll *et al*, 2003, 2005b) and humans (Staley *et al*, 1996). Expression of  $D_3R$  therefore appears to be a potential basis for a reinforcing mechanism in reward-related behavior associated with voluntary intake of addictive drugs and ethanol.

A number of studies have linked  $D_3R$  expression in the nucleus accumbens to BDNF derived from cortical sources (Guillin *et al*, 2001; Le Foll *et al*, 2005b); furthermore, ethanol exposure increases both BDNF and  $D_3R$  within the striatum itself (Jeanblanc *et al*, 2006; McGough *et al*, 2004). The scaffolding protein RACK1 is a key regulator of BDNF expression; RACK1 translocates to the nucleus after exposure of neurons to ethanol and increases expression of BDNF (McGough *et al*, 2004). Jeanblanc et al. (Jeanblanc *et al*, 2006) proposed that the RACK1/BDNF/ $D_3R$  pathway is involved in the control of ethanol consumption in mice. Our hypothesis is that activation of RACK1/BDNF by ethanol may induce expression of  $D_3R$ , which in turn controls and maintains ethanol consumption. This

hypothesis is supported by the data we generated showing that: i) ethanol intake is negligible in  $D_3R^{-/-}$  and robust in WT; ii) increase in RACK1/BDNF/ $D_3R$  is maintained during chronic ethanol intake in WT; iii) forced ethanol intake increases RACK1/BDNF even in  $D_3R^{-/-}$ . Furthermore, chronic voluntary ethanol intake increased  $D_3R$  expression in striatum concomitant with increased expression of BDNF. It is noteworthy that, in the basal condition,  $D_3R^{-/-}$  mice exhibited higher BDNF than WT, consistent with a tendency reported in a recent study (Xing *et al*, 2012). When subjected to forced ethanol intake,  $D_3R^{-/-}$  mice showed a robust increase in BDNF expression in the striatum. Therefore, chronic ethanol intake increases BDNF independently of  $D_3R$  receptor stimulation. The finding that chronic ethanol intake increased RACK1 in striatum of both WT and  $D_3R^{-/-}$  provides additional evidence for the role of RACK1/BDNF/ $D_3R$  pathway in ethanol intake; chronic ethanol intake stimulates RACK1/BDNF pathway leading to  $D_3R$  overexpression and addictive behavior in WT, but not in  $D_3R^{-/-}$ , because this latter lacks  $D_3R$ .

To provide additional evidence, we blocked the BDNF pathway by using the TrkB specific antagonist, ANA-12. We found that ANA-12 reversed ethanol intake both in the two-bottle choice and DID paradigms and strongly decreased the expression of  $D_3R$  in the striatum of WT treated mice. Recently,  $D_3R$  on VTA-SN dopaminergic neurons were found to mediate neuroplasticity effects of several addictive drugs (Collo *et al*, 2012; Collo *et al*, 2013). Therefore, our conclusion about the engagement of striatal RACK1, BDNF and  $D_3R$  in mediating ethanol consumption may be only a part of a more complex mechanism, whose elucidation may require an assessment of the effects of ethanol intake in the VTA-SN dopaminergic neurons.

Finally, in a translational perspective, we tested buspirone, a drug marketed for anxiety disorders, endowed with  $D_3R$  antagonist (Bergman *et al*, 2013; Le Foll and Boileau, 2013; Newman *et al*, 2012) and 5-HT<sub>1A</sub> partial agonist activity (Wong *et al*, 2007). Of note, buspirone shows also high affinity for other D<sub>2</sub>-like receptors (Bergman *et al*,

2013; Kula *et al*, 1994; Tallman *et al*, 1997). D<sub>3</sub>R antagonists may be effective for treating substance use disorders and buspirone has proven effective in several preclinical model of drug abuse (Heidbreder *et al*, 2010; Higley *et al*, 2011; Song *et al*, 2012), but no studies have, so far, investigated its D<sub>3</sub>R antagonist action in ethanol consumption. By both radioligand binding and molecular modeling studies (see Supplementary Information) we found that buspirone: i) shows slight higher affinity at D<sub>3</sub>R than at D<sub>2</sub>R (K<sub>i</sub>, 29 versus 62 nM respectively) and may forms interactions comparable to those of SB277011A in D<sub>3</sub>R, having the antagonist binding mode at D<sub>3</sub> receptor, ii) displays a distinct interaction from the other two antagonists SB277011A and U99194A (different binding energy, different interaction patterns) with D<sub>3</sub>R, consistent with their distinct chemical structure. Thereafter, we found that buspirone induced a significant decrease in ethanol intake in both two-bottle choice and DID paradigms. The dose of 1 mg/kg inhibited ethanol intake in both paradigms, though its effect did not reach statistical significance in DID; 3 and 10 mg/kg, however produced a significant effect in DID. We confirmed the specificity of D<sub>3</sub>R effect by using a selective 5-HT<sub>1A</sub> agonist, 8-OH-DPAT, in the DID. Treatment with 8-OH-DPAT did not impact ethanol intake, whereas, as expected, decreased the body temperature in a stable manner. In a translational perspective, an important issue is the actual availability of buspirone to bind D<sub>3</sub>R in human CNS. Reported buspirone's affinity toward human recombinant D<sub>3</sub>R ranges from 3.5 to 98 nM (Bergman *et al*, 2013; Newman *et al*, 2012), which partially overlaps its affinity for 5-HT<sub>1A</sub> receptors; because buspirone binding to 5-HT<sub>1A</sub> is considered the basis of its anxiolytic activity in humans, it is likely that anxiolytic doses are sufficient to occupy also D<sub>3</sub>R in human CNS. However, the D<sub>3</sub>R-related therapeutic potential of buspirone requires more detailed information, including measurements of D<sub>3</sub>R receptor occupancy in human PET studies, as an essential prerequisite to clinical application.

Finally, as D<sub>3</sub>R<sup>-/-</sup> mice have been shown to exhibit extracellular DA levels

substantially higher than WT, as assessed by microdialysis (Koeltzow *et al*, 1998), a phenomenon related to the lack of autoreceptor function (Joseph *et al*, 2002), we hypothesized that ethanol intake effectively stimulates DA release and transmission in WT, but not in  $D_3R^{-/-}$ , presumably because this latter already displays high extracellular DA levels. To test the hypothesis that treatment with  $D_3R$  antagonists mimicked the high DA phenotype documented in  $D_3R^{-/-}$  (Koeltzow *et al*, 1998), we assessed phosphorylation of DARPP32, that is increased by different addictive drugs, including ethanol (Nuutinen *et al*, 2011; Svenningsson *et al*, 2005), and of GSK3 $\beta$ , that is linked to  $D_2$ -like receptors signaling cascade (Beaulieu *et al*, 2007; Li *et al*, 2009), particularly under hyper-DAergic conditions (Li *et al*, 2009). Treatment with SB277011A increased phosphorylation of DARPP32 and of GSK3 $\beta$  to a level similar to that of  $D_3R^{-/-}$ . Thus, chronic blockade of the  $D_3R$  or its genetic deletion increased DA transmission in striatum, consistent with increased extracellular DA (Joseph *et al*, 2002; Koeltzow *et al*, 1998).

In conclusion, either  $D_3R$  gene deletion or  $D_3R$  pharmacological blockade inhibit ethanol intake. Thus, pharmacological antagonism selectively targeting  $D_3R$  may provide a basis for novel weaning treatments to inhibit ethanol consumption. In this context, buspirone, a drug marketed as anxiolytic since more than 25 years and endowed with  $D_3R$  antagonist activity, exhibits translational potential for treating alcohol addiction.

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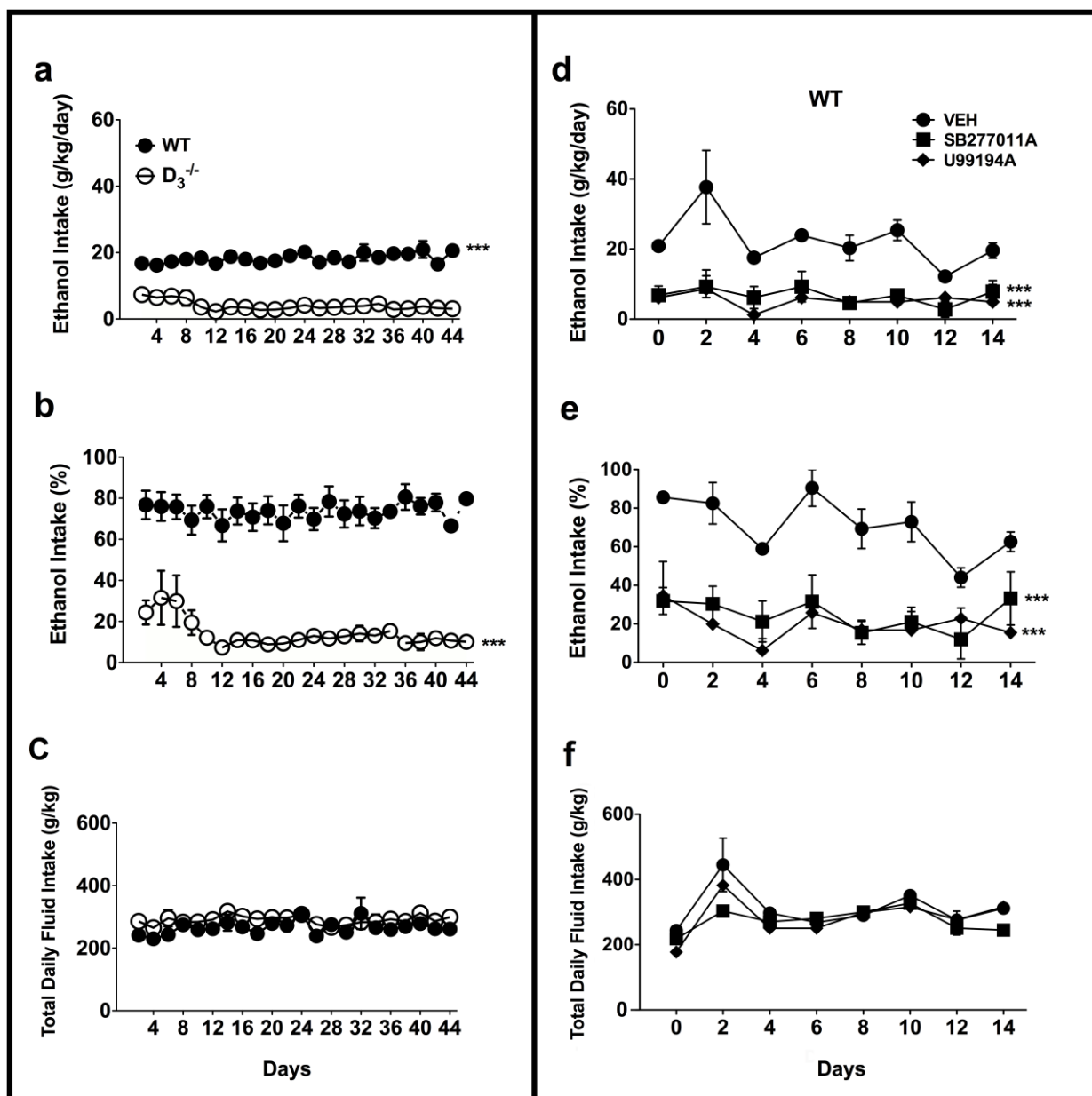
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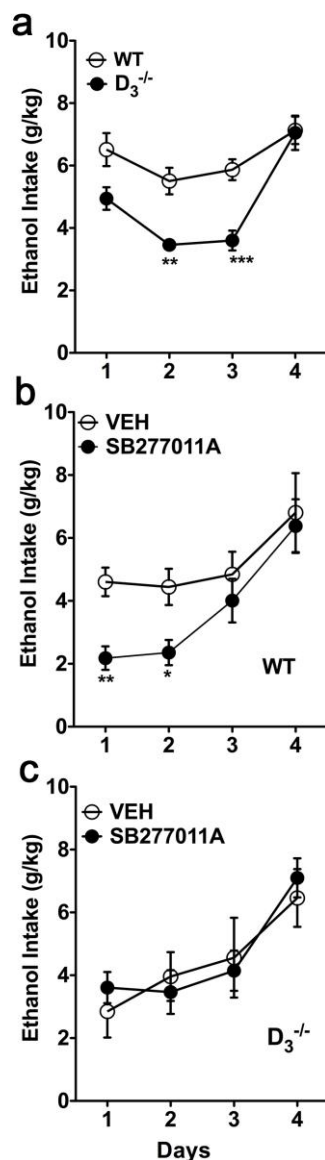
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**Figure 1**

In the two-bottle choice paradigm,  $D_3^{-/-}$  mice show a lower voluntary ethanol intake as compared to wild type (WT).  $D_3$  pharmacological antagonism inhibits ethanol intake in WT mice. In **a** and **b**, voluntary ethanol intake was measured every 24 h, for 44 days, in WT ( $n=30$ ) and  $D_3^{-/-}$  ( $n=30$ ) mice that had free access to water and ethanol solution (10%). Panel **c** shows total fluid intake that was not different in the two groups. In **d** and **e**, voluntary ethanol intake was measured as in **a**, but in mice that had received the day before and kept receiving daily i.p. injection of either saline (vehicle, VEH,  $n=10$ ), U99194A ( $n=10$ ) or SB277011A ( $n=10$ ), either drug at 10 mg/kg. Panel **f** shows total fluid intake in either group that was not affected by drug-treatment.

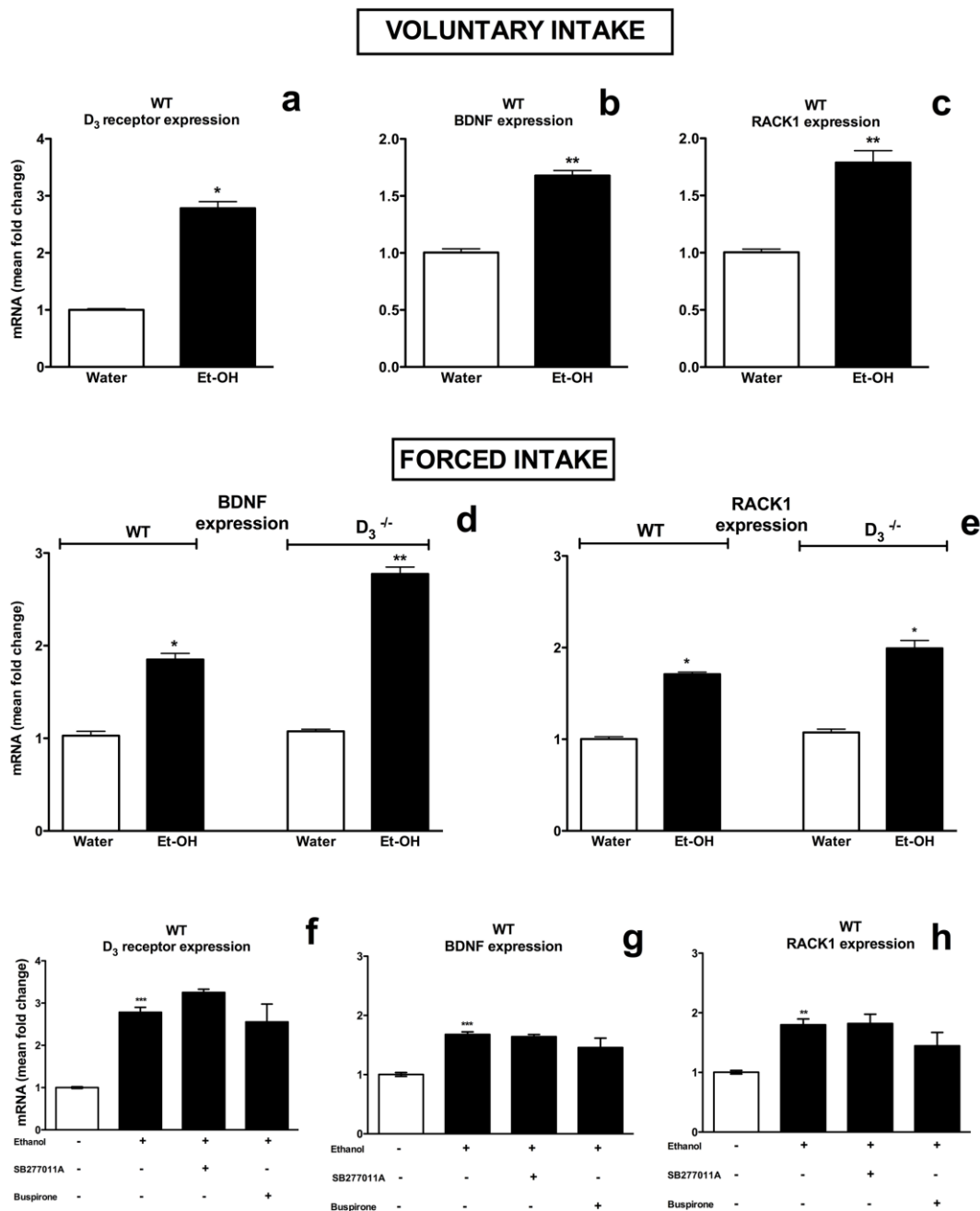
\*\*\*  $P < 0.001$  vs. water or vehicle (VEH). One-way ANOVA and Newman-Keuls post hoc test.



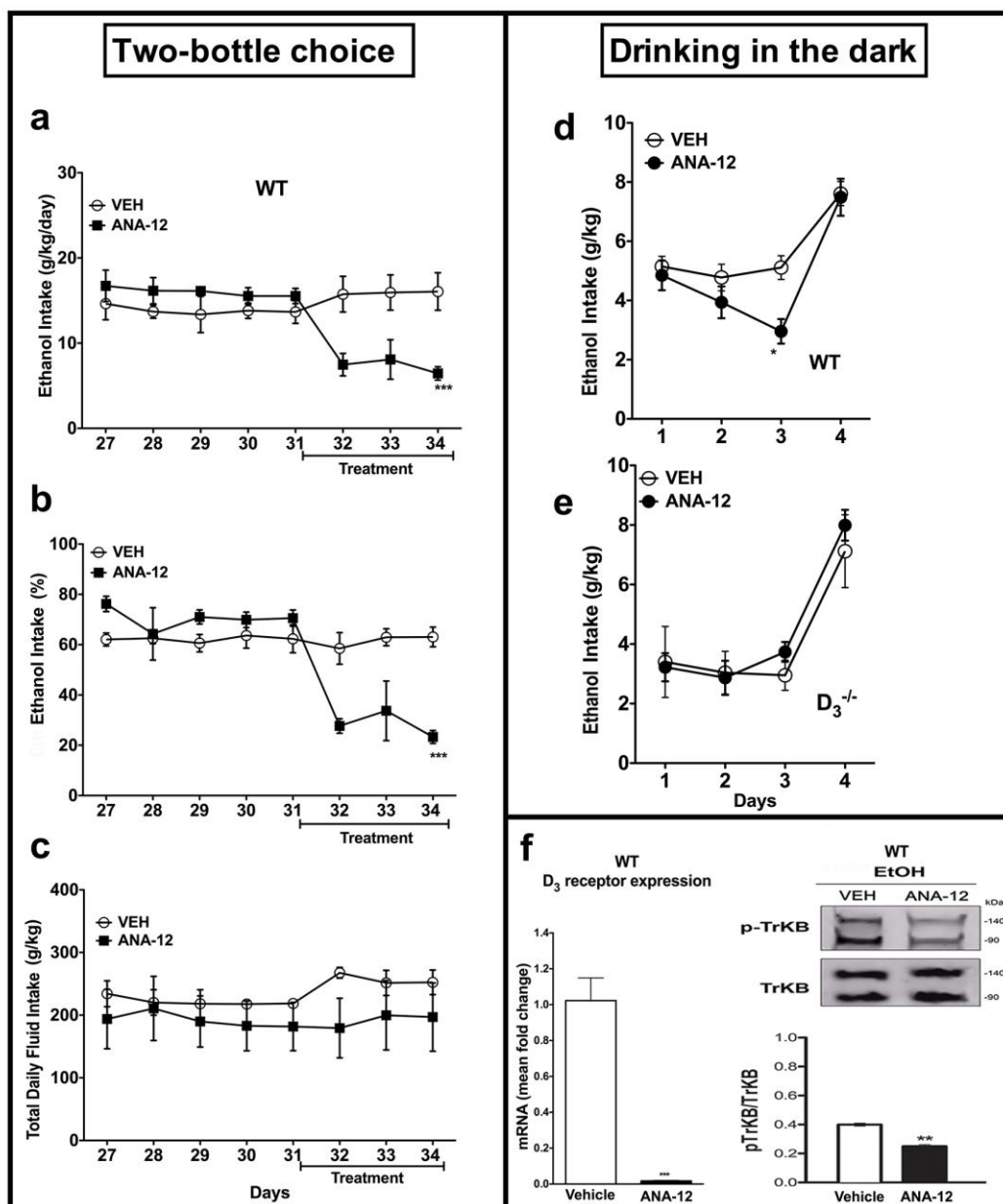
**Figure 2**

In the drinking in the dark (DID) paradigm,  $D_3^{-/-}$  mice show a lower ethanol intake as compared to their wild type (WT) littermates. The  $D_3$  antagonist SB277011A inhibits ethanol intake of WT but not in  $D_3^{-/-}$  mice. In **a**, DID was measured, for 4 days, in WT (n=12) and  $D_3^{-/-}$  (n=12) mice that had limited access (2 hours/day for 3 days and 4 hours the 4<sup>th</sup> day) to ethanol solution (20%). In **b** and **c**, voluntary ethanol intake was measured as in **a**, but in mice that had received the day before and kept receiving daily i.p. injection of either saline (vehicle, VEH, n=10), or SB277011A (n=10), at 10 mg/kg.

\* P<0.05 \*\* P<0.01, \*\*\* P<0.001 vs. wild type (WT) or vehicle (VEH). One-way ANOVA and Newman-Keuls post hoc test.

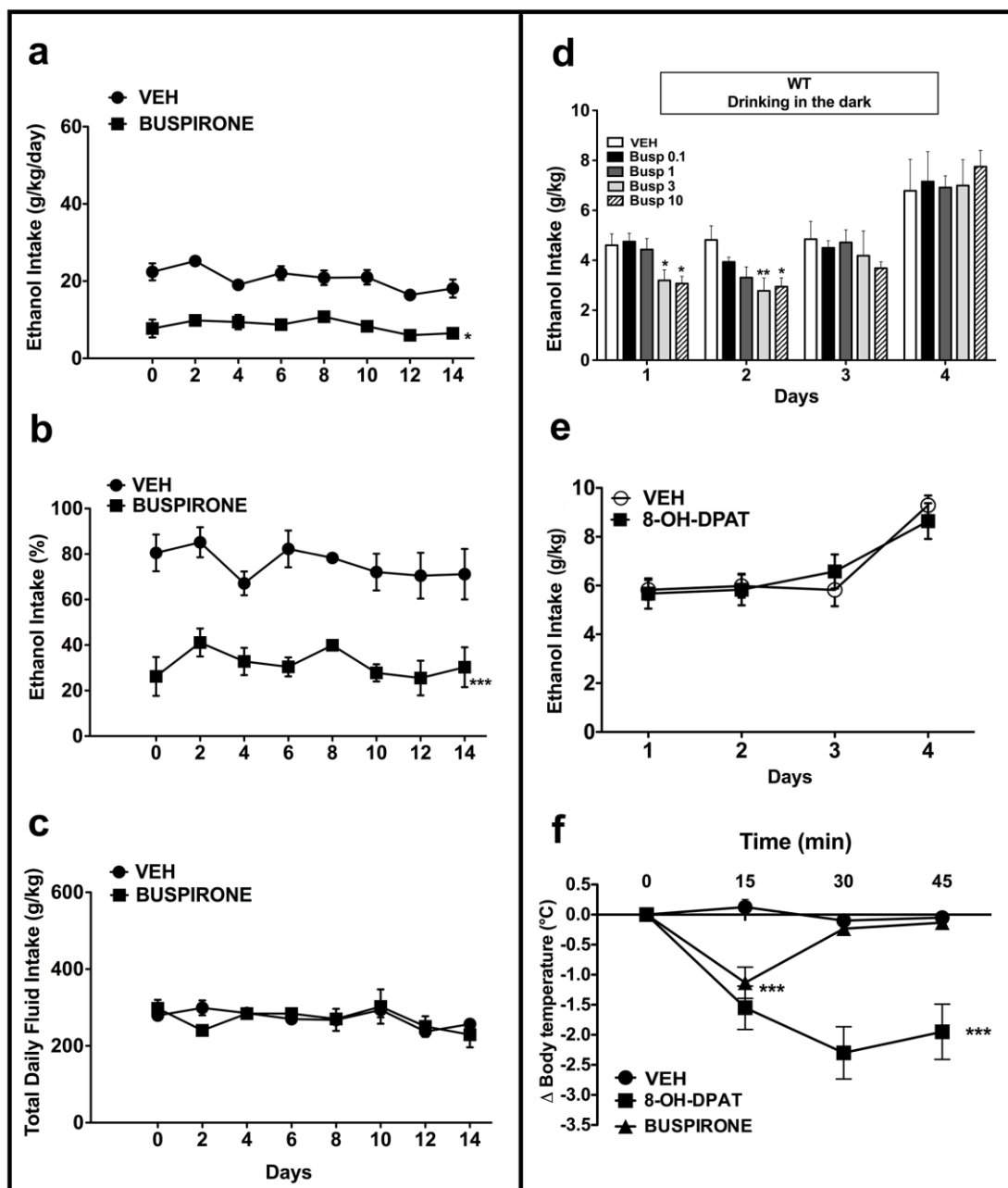
**Figure 3**

Chronic ethanol intake induces D<sub>3</sub> up-regulation, associated with activation of BDNF/RACK1 pathway. Abundance of transcripts in striatum was assessed by quantitative RT-PCR after 44 days of free access to water only (white columns), or to both water and ethanol (black columns, upper panels) or forced ethanol intake (black columns lower panels). In the forced alcohol-drinking procedure (panels d-h), D<sub>3</sub>R<sup>-/-</sup> and WT received 10% ethanol only, with or without SB277011A or buspirone for 14 days. In **a** and **f**, D<sub>3</sub> expression profile in WT; in **b**, **d** and **g**, brain-derived neurotrophic factor (BDNF) expression profile in WT and D<sub>3</sub><sup>-/-</sup>; in **c**, **e** and **h** RACK1 expression profile in WT and D<sub>3</sub><sup>-/-</sup>. Mean fold changes are expressed relative to transcript levels in controls (WT having access to water only). Each column is the mean (± S.E.M.) from 5 different samples. \*P<0.05, \*\*P<0.01 vs. water. One-way ANOVA and Newman-Keuls post hoc test.

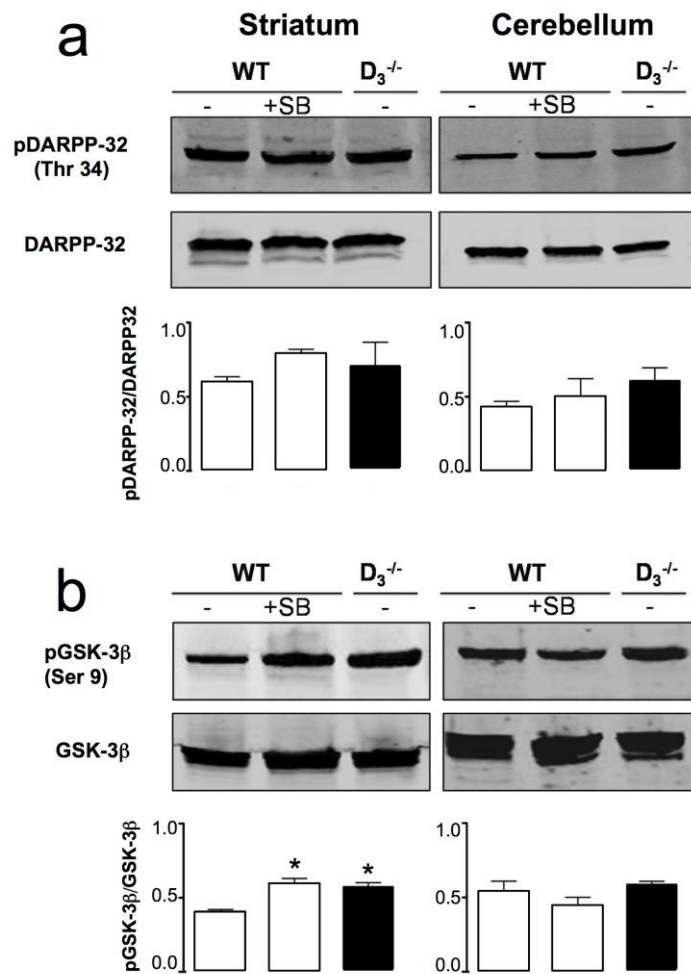
**Figure 4**

The selective TrkB antagonist, ANA-12 reverses ethanol intake of WT mice and induces  $D_3$  receptor down-regulation but does not change ethanol intake of  $D_3^{-/-}$  mice. In **a**, and **b** voluntary ethanol intake was measured every 24 h, for 34 days, in WT ( $n=30$ ) and  $D_3^{-/-}$  ( $n=20$ ) mice that had free access to water and ethanol solution (10%). At the day 31 mice received daily i.p. injection of either vehicle (VEH), or ANA-12 at 0.5 mg/kg. In panel **d** and **e** drinking in the dark (DID) was measured, for 4 days, in WT ( $n=9$ ) and  $D_3^{-/-}$  ( $n=9$ ) mice that had limited access (2 hours/day for 3 days and 4 hours the 4<sup>th</sup> day) to ethanol solution (20%), daily injected with vehicle or ANA-12 1h before the test. \* $P<0.05$ , \*\* $P<0.01$ , \*\*\* $P<0.001$  vs. VEH, One-way ANOVA and Newman-Keuls post hoc test. In **f**, the abundance of transcripts of  $D_3$  receptor in striatum was assessed by quantitative RT-PCR in WT mice exposed to chronic voluntary ethanol intake. Mean fold changes are expressed relative to transcript levels in controls. the abundance of phosphorylated TrkB was assessed by immunoblot, in the striatum WT treated with ANA-12 and exposed to the voluntary ethanol intake. Bars show mean ( $\pm$  S.E.M.). \*\* $P<0.01$ , \*\*\* $P<0.001$  vs. vehicle, One-way ANOVA and Newman-Keuls post hoc test. One-way ANOVA and Newman-Keuls post hoc test.



**Figure 5**

Buspirone inhibits ethanol intake in WT mice both in the two bottle choice and DID paradigm. In **a**, **b**, voluntary ethanol intake was measured every 24 h, for 44 days, in WT ( $n=20$ ) and  $D_3^{-/-}$  ( $n=20$ ) mice that had free access to water and ethanol solution (10%). Mice received for 14 days, from day 31, daily i.p. injection of either vehicle (VEH) or buspirone at 1 mg/kg. Panel **c** shows total fluid intake that was not changed by buspirone. \* $P<0.05$ , \*\*\*  $P<0.01$  vs VEH. One-way ANOVA and Newman-Keuls post hoc test. Panel **d** shows the dose ranging of buspirone (0.1, 1, 3 and 10mg/kg) in WT mice exposed to the drinking in the dark (DID) paradigm. DID was measured, for 4 days, in WT ( $n=33$ ) that had limited access (2 hours/day for 3 days and 4 hours the 4<sup>th</sup> day) to ethanol solution (20%). \* $P<0.05$ , \*\*  $P<0.01$  vs VEH. One-way ANOVA and Newman-Keuls post hoc test. In **e**, the effect of the selective 5-HT<sub>1A</sub> agonist, 8-OH-DPAT in DID paradigm. 8-OH-DPAT at 1mg/kg did not change ethanol intake. In **f**, the action on 5-HT<sub>1A</sub> of 3 mg/kg buspirone was compared to 1 mg/kg 8-OH-DPAT by assessing the pharmacologically-induced hypothermia. \*\*\*  $P<0.001$  vs. VEH. One-way ANOVA and Newman-Keuls post hoc test.

**Figure 6**

Dopamine receptor signaling is enhanced in striatum of  $D_3R^{-/-}$  mice and of SB277011A-treated WT mice. The abundance of phosphorylated DARPP-32 (Thr 34, **a**) and phosphorylated GSK3 $\beta$  (Ser 9, **b**) was assessed by immunoblot, in the striatum of WT mice exposed to the long-term voluntary ethanol intake (white columns) and injected i.p. for 14 days with either vehicle or 10 mg/kg SB277011A and in  $D_3^{-/-}$  (black columns). Brain tissues were taken 1 hour after the last administration of either vehicle or SB277011A. Bar graphs show mean ( $\pm$  S.E.M.) of intensities normalized against the respective non-phosphorylated protein. Each column is the mean ( $\pm$  S.E.M.) from 5 different samples.\*  $P < 0.05$  vs. control (vehicle-injected WT). Two-way ANOVA and Newman-Keuls post hoc test.