

# Multitopic Supramolecular Detection of Chemical Warfare Agents by Fluorescent Sensors

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## Supporting Information

**ABSTRACT:** Two new naphthalimide derivatives (hosts), bearing one or two ethanolamine arms, respectively, were synthesized and tested as fluorescent probes for the detection of a chemical warfare agent simulant (guest). The resulting host–guest hydrogen bonds allowed a recognition mechanism based on supramolecular interactions. The sensor with two ethanolamine groups showed excellent sensibility and selectivity in a turn-on fluorescent response.



## INTRODUCTION

Terrorist attacks are nowadays the most dangerous threat in the international scenario. In particular, some of these attacks have already been conducted by using chemical substances, called chemical warfare agents (CWAs). Although CWAs are outlawed and prohibited in many countries, the recent terrorist attacks in the Middle East and also in Europe highlight the importance to detect in real time the presence of these agents.<sup>1</sup> From a chemical point of view, CWAs are organophosphorus (OP) compounds, classified into G and V series.<sup>2,3</sup> For security reasons, the use of these compounds is precluded also for research activity; thus, the development of new and more efficient sensors and relative recognition studies can only be conducted by using less toxic simulants, such as dimethyl methylphosphonate (DMMP).<sup>4a,b</sup> In fact, recent studies demonstrated that DMMP is one of the best simulants for G series nerve agents.<sup>4c</sup> The detection via mass spectrometry is very sensitive but, due to the instrumental complexity, difficult to be realized in real time. Thus, sensing by using molecular probes is the preferred way. Recently, two mechanisms have been exploited in the CWAs recognition: (i) the covalent and (ii) the supramolecular approaches. The covalent mechanism is based on a covalent and irreversible reaction between the sensor and the analyte, leading to a detectable response.<sup>5</sup> The main limitations of this approach are represented by scarce selectivity, impossibility to reuse the sensor, and some false-positive responses.<sup>6</sup> Conversely, the more recent supramolecular approach shows many advantages.<sup>7</sup> In fact, the possibility to exploit noncovalent interactions leads to a reversible detection mechanism thus affording reusable sensors.<sup>8</sup> In addition, a “multitopic approach” involves more than one site of the OP analyte, thus increasing the selectivity.<sup>9</sup>

Therefore, this new frontier of “supramolecular recognition of CWAs via a multitopic approach” paves the way for a new generation of CWAs sensors. In fact, the chemical structures of the proposed receptors are designed to simultaneously recognize two or more sites of the target analyte, thus increasing the selectivity and reducing the false-positive responses. So far, few examples of supramolecular sensors that interact with the simulant by hydrogen bonds (gel)<sup>10–15</sup> or hydrophobic effects (synthetic macrocycles),<sup>16–20</sup> have been reported. In both these examples, the recognition mechanisms are based on a monotopic approach. Recently, our research group has developed the first example of a multitopic receptor based on two different recognition sites.<sup>9</sup>

In the present study, we report on the syntheses of two new fluorescent sensors that show one or two ethanolamine arms (naphthyl-mono-AE and naphthyl-di-AE, Chart 1), respectively, for the detection of DMMP via a multitopic supramolecular approach.

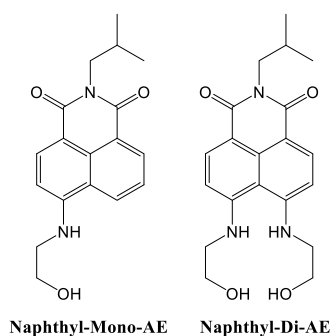
In particular, these new probes allow the formation of multiple hydrogen bonds with the DMMP analyte, thus increasing the affinity and selectivity. Molecular recognition studies have been performed by using fluorescence, UV–vis, one-dimensional–two-dimensional NMR (1D–2D NMR), and mass spectrometry. Naphthyl-di-AE shows many advantages with respect to already reported sensors.<sup>11–20</sup> In fact, it represents the first multitopic/metal-free/turn-on fluorescent sensor that is able to recognize DMMP with high selectivity. In addition, the easy synthetic pathway, the large Stock-shift

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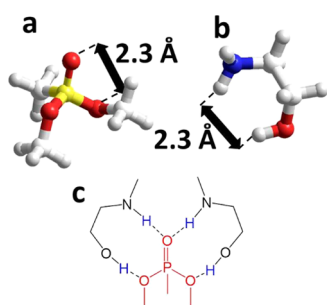
**Chart 1. Chemical Structures of Naphthyl-mono-AE (Left) and Naphthyl-di-AE (Right)**



emission, and the possibility to functionalize the organic scaffold to obtain heterogeneous devices pave the way for many applications of **naphthyl-di-AE** in real conditions.

## RESULTS AND DISCUSSION

The multitopic recognition approach requires at least two binding sites for the guest. DMMP shows three possible H-bond acceptor sites: P=O and two  $-\text{OCH}_3$  groups that are ca. 2.3 Å away from P=O (Figure 1a). The ethanolamine



**Figure 1.** Geometry optimizations of (a) DMMP, (b) ethanolamine and relative distances between atoms involved in molecular recognition (force field MM+), (c) multitopic proposed approach.

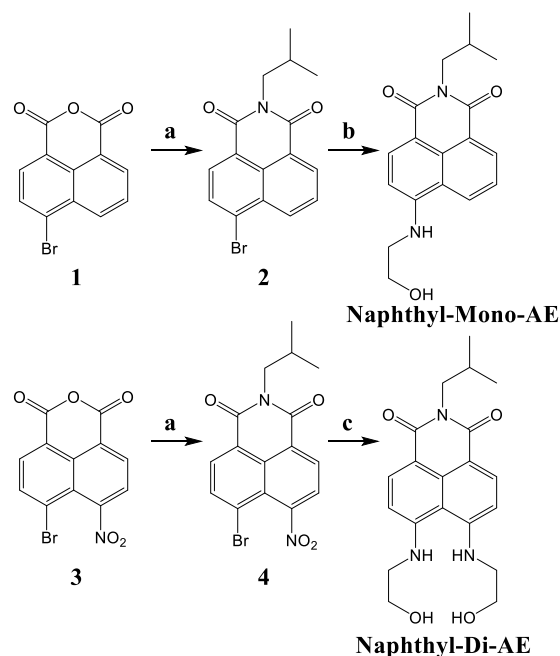
molecule shows appropriate geometry to simultaneously interact with P=O and oxymethylene groups of DMMP (Figure 1b) and, Figure 1c shows the possible resulting H-bond network. On the basis of the above considerations, we designed two molecular probes (**naphthyl-mono-AE** and **naphthyl-di-AE**), bearing naphthalimide moiety, that are able to give fluorescent response upon the recognition event.<sup>21,22</sup>

Scheme 1 reports the synthesis of the receptors. **Naphthyl-mono-AE** was synthesized starting from Br-anhydride **1**, which in the presence of isobutylamine in ethanol led to the Br-imide derivative **2** (see the Supporting Information). After the treatment of **2** with excess ethanolamine in 2-methoxyethanol at reflux, **naphthyl-mono-AE** was isolated in high yield (Scheme 1, see the Supporting Information).

Following the same synthetic strategy, **naphthyl-di-AE** was prepared starting from Br-NO<sub>2</sub>-anhydride **3** by using a well-known procedure to functionalize Br-NO<sub>2</sub>-naphthalimide scaffold.<sup>21,22</sup> A large excess of ethanolamine (10 equiv) and a longer reaction time (36 h) in the last step were employed.

The UV-vis spectrum of **naphthyl-mono-AE** shows an absorption band centered at 425.2 nm ( $\epsilon = 15600$ ) (see the Supporting Information). The fluorescence spectrum, obtained by excitation at  $\lambda_{\text{ex}} = 340$  nm, shows a strong emission band

**Scheme 1. Synthesis of the Receptors<sup>a</sup>**

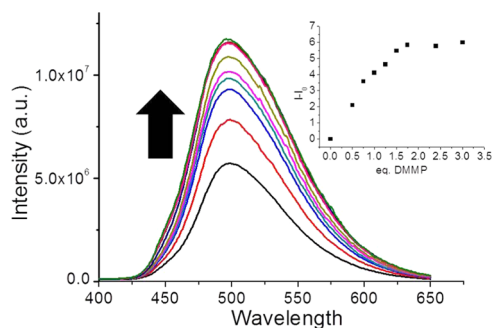


<sup>a</sup>Reagents and conditions: (a) isobutylamine, ethanol, 50 °C, 18 h, 92% for **2**, 56% for **4**; (b) ethanolamine, 2-methoxyethanol, reflux, 18 h, 71%; (c) ethanolamine, 2-methoxyethanol, reflux, 36 h, 47%.

centered at 470 nm, with a strong Stokes shift (see the Supporting Information). Recognition properties of **naphthyl-mono-AE** and DMMP were evaluated by following the increase of the emission band at 470 nm. In particular, upon the progressive addition of the guest, a turn-on fluorescence response was observed, probably due to a photoinduced electron transfer mechanism.<sup>23</sup> The obtained binding constant value was 3467 M<sup>-1</sup> (see the Supporting Information), in agreement with the formation of two hydrogen bonds with DMMP.<sup>24</sup>

To increase the affinity for DMMP, we introduced a second ethanolamine arm in the naphthalimide scaffold, thus obtaining **naphthyl-di-AE** (see the Supporting Information). The related UV-vis spectrum shows two main bands centered at 344.6 nm ( $\epsilon = 5100$ ) and 438.4 nm ( $\epsilon = 19800$ ), respectively (see the Supporting Information). By excitation at 340 nm, the emission band at 509 nm shows a higher Stokes shift with respect to **naphthyl-mono-AE**, thus confirming the potential application as a real fluorescent sensor. Fluorescence measurements upon addition of progressive amounts of DMMP show a strong turn-on response (see Figure 2), with a calculated binding constant value of 10 471 M<sup>-1</sup>, more than 3 times larger than that observed for **naphthyl-mono-AE**, thus demonstrating that the introduction of the second chelating arm leads to a stronger supramolecular complex with DMMP. The increase of the binding constant value is in agreement with our previous study performed by using uranyl salen complexes.<sup>8,9</sup> In particular, the results here obtained confirm that the presence of additional two H-bonds in the host-guest complex leads to a binding constant value 3 times higher. This result can be fundamental for the design of other CWA receptors.

The detection limit of DMMP is 1 ppm, both in the case of **naphthyl-mono-AE** and **naphthyl-di-AE**. The time-dependent fluorescence emission spectra, recorded for both receptors before and after the addition of 1 equiv of DMMP, show an



**Figure 2.** Fluorescence intensity behavior of **naphthyl-di-AE** ( $1 \times 10^{-5}$  M in dry toluene solution,  $\lambda_{\text{ex}}$  340 nm) upon the DMMP additions; the inset shows the intensity increase upon the progressive addition of DMMP equivalents.

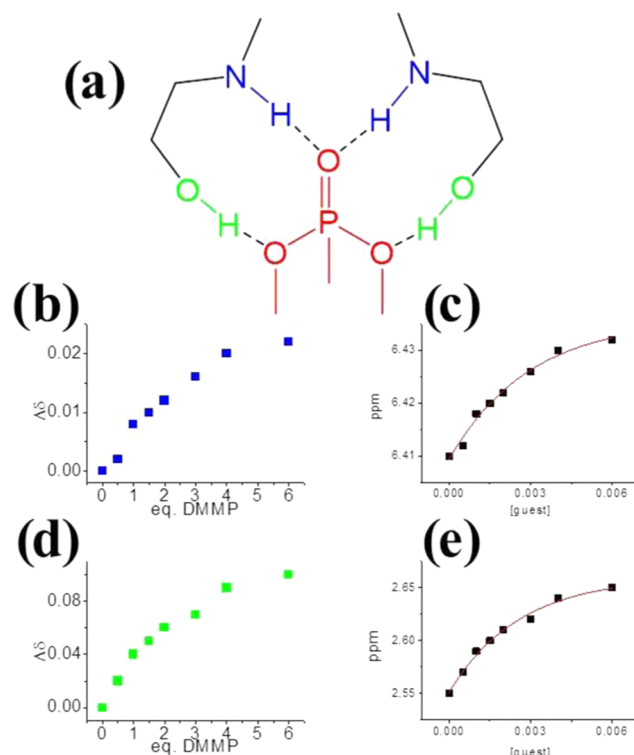
instant emission response, stable for at least 10 min (see the [Supporting Information](#)).

Electrospray ionization-mass spectrometry (ESI-MS) measurements also confirmed the supramolecular complex stoichiometry. In fact, the observed peak at  $m/z = 518.4$  is ascribed to the  $[\text{naphthyl-di-AE@DMMP+Na}]^+$  ion (see the [Supporting Information](#)). The geometry of the host–guest system was elucidated by  $^1\text{H}$  NMR experiments. In particular, titration of **naphthyl-di-AE** with DMMP in  $\text{CDCl}_3$  shows that (i) signals relative to NH and OH protons of the host (6.40 and 2.55 ppm, respectively) undergo a downfield shift after the addition of the guest in a fast-exchange mechanism compared to the  $^1\text{H}$  NMR time scale ([Figure 3b,d](#)); and (ii) the multiplicity of the methylene signal of the ethanolamine moieties changes from quartet to triplet. These overall data suggest the synergy of NH and OH groups toward the DMMP chelation, thus confirming the multitopic recognition process ([Figure 3a](#)). Nonlinear curve fits relative to these data show a binding constant value of  $468 \text{ M}^{-1}$  ([Figure 3c,e](#)), lower with respect to those obtained with fluorescence measurements, probably due to the different solvent used (toluene is less competitive than chloroform in the hydrogen bond formation).

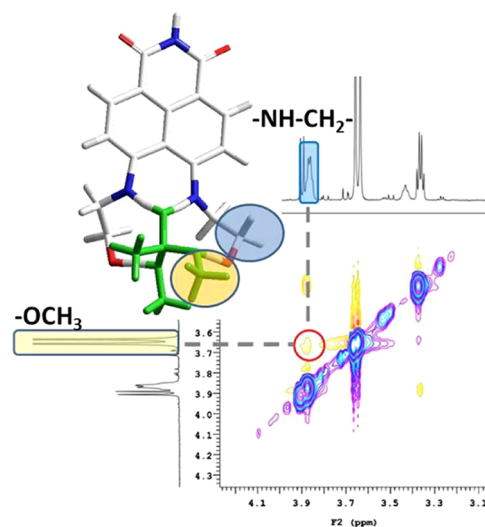
To further confirm the geometry of **naphthyl-di-AE@DMMP** supramolecular complex, TROESY measurements were also carried out (see the [Supporting Information](#)). In particular, an equimolar solution ( $1 \times 10^{-3}$  M in  $\text{CDCl}_3$ ) of **naphthyl-di-AE** and DMMP shows ROE contacts between the methylene groups bound to the  $-\text{NH}$  function ( $-\text{NH}-\text{CH}_2-$ ) of the receptor and the  $-\text{OCH}_3$  substituent of DMMP ([Figure 4](#)). This further result unambiguously confirms the chelation of DMMP by the **naphthyl-di-AE** receptor.

High selectivity is of crucial importance for real applications, and the classical CWAs sensing based on covalent reactions between a nucleophilic sensor substituent and a given nerve agent simulant can lead to false-positive response due to the low specificity of this reaction. In contrast, the goal of the multitopic supramolecular approach consists in the formation of reversible interactions with more than two sites of the guest, thus increasing the selectivity and reducing the possibility of a false-positive detection.

For this reason, we also performed selectivity and competition tests. In particular, atmospheric air (in which we measured 24 000 ppm of water, 400 ppm  $\text{CO}_2$ , 5 ppm  $\text{NO}$ , and 10 ppm  $\text{CO}$ ) was bubbled for 30 min into a  $1 \times 10^{-5}$  M toluene solution of **naphthyl-di-AE**. The fluorescence spectra of the receptor were acquired before and after air bubbling.



**Figure 3.** (a) Multitopic proposed recognition mechanism; chemical shift variation of signals relative to (b) NH (blue) and (d) OH (green) groups of the ethanolamine moieties during the  $^1\text{H}$  NMR titration of **naphthyl-di-AE** with DMMP in  $\text{CDCl}_3$ . (c, e) The nonlinear curve fits relative to NH and OH, respectively; although other molecular receptors are able to detect CWAs simulant by using Zn as metal Lewis acid, **naphthyl-di-AE** recognizes DMMP without the assistance of metal ion, as confirmed by ESI-MS measurements (see the [Supporting Information](#)). In addition, we tested the ability of **naphthyl-di-AE** to chelate  $\text{Zn}^{2+}$  ion, without positive results.

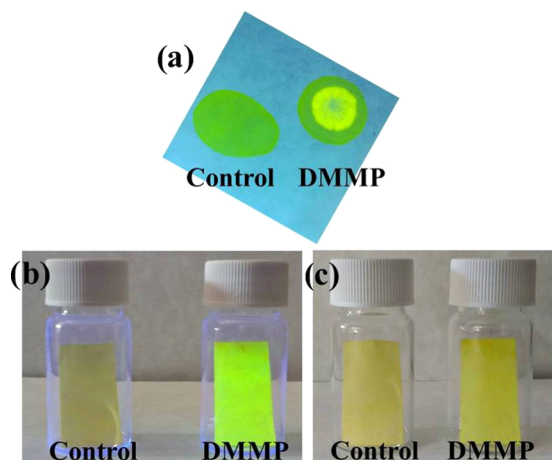


**Figure 4.** Selected region of the TROESY spectrum of an equimolar ( $1 \times 10^{-3}$  M in  $\text{CDCl}_3$ ) solution of **naphthyl-di-AE** and DMMP. The inset shows the minimized structure (force field MM+) of the supramolecular complex, highlighting the functional groups involved in the ROE contacts (DMMP is shown in green; isobutyl group of the receptor was omitted for clarity).

The two obtained spectra are superimposable (see the [Supporting Information](#)), thus demonstrating that the

fluorescence profile of **naphthyl-di-AE** is not affected by the standard analytes/contaminants contained in air. Then, this air-saturated **naphthyl-di-AE** solution was exposed to DMMP and we observed an increase in the intensity of the emission spectra. These results confirm the high selectivity of **naphthyl-di-AE** toward DMMP also in real atmospheric conditions. Selectivity test was further conducted by using acetic acid as a competitor. Also, in this case, the **naphthyl-di-AE** emission remains unaffected up to 10 equiv of acetic acid.

Then, we performed a practical test using a filter paper adsorbed with **naphthyl-di-AE** (Test Strip, Figure 5). A



**Figure 5.** Test Strip with vapors and solution: (a) solution of **naphthyl-di-AE** ( $1 \times 10^{-3}$  M) adsorbed on filter paper without (control on the left) and with the presence of  $1 \mu\text{L}$  of DMMP (on the right) under UV-vis lamp (365 nm); (b) solution of **naphthyl-di-AE** ( $1 \times 10^{-3}$  M) adsorbed on filter paper ( $2 \times 3.5$  cm) without (control on the left) and with the presence of vapors of DMMP (40 ppm, on the right) under UV-vis lamp (365 nm); (c) filter papers showed in (b) under daylight. Due to the low volatility of DMMP (bp  $181^\circ\text{C}$ , vapor pressure  $0.962$  mm Hg at  $25^\circ\text{C}$ ), Test Strip was conducted for 1 h and then the filter paper was exposed under the UV-vis lamp.

stronger fluorescent spot appeared in correspondence of the addition of a drop of simulant (Figure 5a), thus confirming the possibility of a fast and reliable practical test. Then, we adsorbed common filter paper with a (1 mM) solution of the receptor. After its exposure to DMMP vapors (40 ppm, in a closed vial at room temperature), a clear enhancement of emission was observed, both under UV-vis (Figure 5b) and daylight (Figure 5c) irradiations.

These experiments confirm the ability of **naphthyl-di-AE** to detect DMMP with high affinity and sensibility, also in a common practical device.

The photostability of **naphthyl-mono-AE** and **naphthyl-di-AE** in air was evaluated to validate these receptors for long operation time. In particular, after exposure of sensors to air and daylight for 1 month, they preserve the recognition abilities without the loss of efficiency and sensibility. This is due to the high stability of naphthalimide derivatives under normal conditions.<sup>25</sup>

## CONCLUSIONS

Two new fluorescent metal-free sensors for a CWA simulant, differing in the number of ethanolamine arms, have been synthesized. Both probes recognize DMMP via multitopic supramolecular approach by the formation of a H-bond

network. The recognition event detected by fluorescence emission, with a large Stokes shift and a turn-on emission fluorescence, highlights the presence of DMMP in a solution. The probe with two ethanolamine functionalities (**naphthyl-di-AE**) shows higher affinity for DMMP with respect to **naphthyl-mono-AE**. The supramolecular host-guest geometry between **naphthyl-di-AE** and DMMP, as well as the related stoichiometry was elucidated by ESI-MS and NMR 1D–2D measurements. Both selectivity and specificity, also in competition conditions, were evaluated thus demonstrated the importance of a multitopic approach for the recognition mechanism. Test Strip demonstrated the possibility to realize practical device for the detection of CWAs.

With the aim to realize real sensor devices, these receptors show many interesting features: (i) easy synthetic pathway; (ii) absence of metal ions; (iii) turn-on fluorescence detection of the analyte; (iv) selectivity and specificity; (v) reversibility, due to the nature of the interaction with the analyte; (vi) possibility to easily functionalize the scaffold (particularly in the imide moiety) to anchor it onto solid supports; (vii) positive to the Test Strip; and (viii) photostability and fast-emission response. Actually, we are working on the functionalization of this organic scaffold to obtain solid devices.

## EXPERIMENTAL SECTION

**General Experimental Methods.** The NMR experiments were carried out at  $27^\circ\text{C}$  on a Varian UNITY Inova 500 MHz spectrometer ( $^1\text{H}$  at 499.88 MHz,  $^{13}\text{C}$  NMR at 125.7 MHz) equipped with a pulse-field gradient module (Z axis) and a tunable 5 mm Varian inverse detection probe (ID-PFG). The ESI mass spectra were acquired on a ES-MS Thermo-Finnigan LCQ-DECA using  $\text{CH}_3\text{CN}$  (positive or negative ion mode). Luminescence measurements were carried out using a Cary Eclipse Fluorescence spectrophotometer at room temperature. The emission was recorded at  $90^\circ$  with respect to the exciting line beam, using 5:5 slit widths for all measurements. All chemicals were reagent grade and used without further purification. The three-dimensional minimized structure of **naphthyl-di-AE** was obtained by using HyperChem v8.0.7, MM+ force field. The NMR characterization of compound **2** is in tune with the literature data.<sup>26</sup>

**Procedure for  $^1\text{H}$  NMR Titrations.** Two mother solutions of host and guest ( $7.0 \times 10^{-3}$  M) in  $\text{CDCl}_3$  were prepared. From these, different solutions with different host/guest ratio were prepared, and  $^1\text{H}$  NMR spectra were recorded at  $27^\circ\text{C}$ . TROESY spectrum was acquired by using equimolar solution of host and guest. With this data treatment, the apparent binding constants were determined using HypNMR (version 4.0.71), a software designed to extract equilibrium constants from potentiometric and/or spectrophotometric titration data.

**Procedure for Test Strip.** Fifty microliters of a solution of **naphthyl-di-AE** ( $1 \times 10^{-3}$  M in  $\text{CH}_2\text{Cl}_2$ ) was adsorbed onto two portions of filter paper ( $2 \times 3.5$  cm) and solvent was removed under air flow. One of these paper filter was inserted into a vial (20 mL) containing  $1 \mu\text{L}$  of DMMP 0.1 M in  $\text{CH}_2\text{Cl}_2$ . The other one was inserted in a vial (20 mL) without DMMP (control in Figure 5). Both vials were maintained at room temperature for 1 h. After this time, filter papers were observed under UV-vis lamp at 365 nm.

**Procedure for Fluorescence Titrations.** Two mother solutions of host and guest ( $1.0 \times 10^{-3}$  M) in dry solvent were prepared. From these, different solutions with different receptor/guest ratios were prepared, and the emission spectra

were recorded at 25 °C. Fluorescence titrations with **naphthyl-mono-AE** and **naphthyl-di-AE** were carried out using  $\lambda_{\text{exc}} = 340$  nm in dry toluene,  $\lambda_{\text{em}} = 470$  nm for **naphthyl-mono-AE**, and  $\lambda_{\text{em}} = 509$  nm for **naphthyl-di-AE** at 25 °C. The apparent binding affinities of receptors with DMMP were estimated using HypSpec (version 1.1.33),<sup>27</sup> a software designed to extract equilibrium constants from potentiometric and/or spectrophotometric titration data. HypSpec starts with an assumed complex formation scheme and uses a least-squares approach to derive the spectra of the complexes and the stability constants.  $\chi^2$  test (chi-square) was applied, where the residuals follow a normal distribution (for a distribution approximately normal, the  $\chi^2$  test value is around 12 or less). In all of the cases,  $\chi^2 \leq 10$  were found, as obtained by 3 independent measurements sets.

**Synthesis of 4-Br-N-isobutyl-1,8-naphthalimide (2).** A 1.5 g (5.4 mmol) of (1) was dissolved in 90 mL of absolute ethanol at 70 °C for 30 min. Then, the mixture was kept at 50 °C, and 800  $\mu\text{L}$  (8.1 mmol) of isobutylamine in ethanol (20 mL) was added dropwise under nitrogen. The reaction mixture was stirred at 50 °C for 18 h, checking the reaction by thin-layer chromatography (TLC) (*n*-hexane/ethyl acetate 7:3). The solid precipitate was filtered to give 1.7 g (5.1 mmol, yield 92%) of (2). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  8.66 (d, *J* = 8.0 Hz, 1H), 8.58 (d, *J* = 7.5 Hz, 1H), 8.41 (d, *J* = 8 Hz, 1H), 8.05 (d, *J* = 7.5 Hz, 1H), 7.85 (t, *J* = 7.5 Hz, 1H), 4.04 (d, *J* = 7.5 Hz, 2H), 2.23 (m, 1H), 0.98 (d, *J* = 6.5 Hz, 6H) ppm.<sup>26</sup>

**Synthesis of Naphthyl-mono-AE.** A 0.3 g (0.9 mmol) of (2) was dissolved in 2-methoxyethanol (30 mL) with 600  $\mu\text{L}$  (10 mmol) of ethanolamine. The reaction mixture was stirred at reflux for 18 h under nitrogen. Then, the solvent was removed, and the crude compound was purified by column chromatography (silica gel, CHCl<sub>3</sub>/MeOH 9:1) to obtain 200 mg (0.64 mmol, 71% yield) of **naphthyl-mono-AE**. <sup>1</sup>H NMR (500 MHz, acetone-*d*<sub>6</sub>):  $\delta$  8.60 (d, *J* = 8.5 Hz, 1H), 8.51 (d, *J* = 8.5 Hz, 1H), 8.36 (d, *J* = 8.5 Hz, 1H), 7.68 (t, *J* = 7.5, 1H), 6.94 (bs, 1H), 6.90 (d, *J* = 7.5 Hz, 1H), 3.97 (d, *J* = 7.5 Hz, 2H), 3.92 (q, *J* = 5.5 Hz, 2H), 3.62 (q, *J* = 5.5 Hz, 2H), 2.2 (m, 1H), 0.92 (d, *J* = 7.0 Hz, 6H) ppm. <sup>13</sup>C NMR (125 MHz, methanol-*d*<sub>4</sub>):  $\delta$  20.7, 28.8, 46.9, 48.2, 61.1, 105.4, 109.5, 121.8, 123.4, 126.1, 129.1, 131.0, 132.4, 136.8, 152.2, 166.1, 166.5 ppm. Anal. calcd for C<sub>18</sub>H<sub>20</sub>N<sub>2</sub>O<sub>3</sub>: C, 69.21; H, 6.45; O, 15.37. Found: C, 69.18; H, 6.41; O, 15.33. ESI-MS: *m/z* 336.0 [M + Na]<sup>+</sup>, 353.1 [M + K]<sup>+</sup>.

**Synthesis of 4-Br-5-nitro-N-isobutyl-1,8-naphthalimide (4).** A 1 g (3.1 mmol) of (3)<sup>21a</sup> was dissolved in 90 mL of absolute ethanol at 70 °C for 30 min. Then, the mixture was kept at 50 °C, and 320  $\mu\text{L}$  (3.1 mmol) of isobutylamine in absolute ethanol (20 mL) was added dropwise. The reaction mixture was stirred at 50 °C for 18 h under nitrogen, checking the reaction by TLC (*n*-hexane/ethyl acetate 7:3). The solid precipitate was filtered, obtaining 620 mg (1.64 mmol, 53% yield) of (4). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  8.71 (d, *J* = 7.5 Hz, 1H), 8.52 (d, *J* = 8.0 Hz, 1H), 8.22 (d, *J* = 8.0 Hz, 1H), 7.93 (d, *J* = 8.0 Hz, 1H), 4.04 (d, *J* = 7.5 Hz, 2H), 2.23 (m, 1H), 0.98 (d, *J* = 6.5 Hz, 6H) ppm. <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>):  $\delta$  = 163.09, 162.33, 135.96, 132.42, 131.28, 130.60, 125.74, 124.09, 123.52, 122.45, 121.22, 47.64, 27.32, 20.22 ppm. ESI-MS *m/z* 377.1 [M + H]<sup>+</sup>. Anal. calcd for C<sub>16</sub>H<sub>13</sub>BrN<sub>2</sub>O<sub>4</sub>: C, 50.95; H, 3.47; O, 16.97. Found: C, 50.91; H, 3.41; O, 16.94.

**Synthesis of Naphthyl-di-AE.** 172 mg (0.46 mmol) of (4) was dissolved in 20 mL of 2-methoxyethanol under

nitrogen atmosphere, and 10 equiv of ethanolamine was added to the solution. The reaction mixture was stirred at reflux for 36 h. Then, the solvent was removed, and **naphthyl-di-AE** (80 mg, 0.2 mmol, 47% yield) were obtained after purification by flash chromatography (silica gel, CHCl<sub>3</sub>/MeOH 9:1). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  8.43 (d, *J* = 8.5 Hz, 2H), 6.74 (d, *J* = 8.5 Hz, 2H), 6.60 (t, *J* = 5.0 Hz, 2H), 4.05 (q, *J* = 5.0 Hz, 4H), 4.01 (d, *J* = 7.5 Hz, 2H), 3.42 (q, *J* = 5.0 Hz, 4H), 2.55 (t br, 2H), 2.24 (m, 1H), 0.96 (d, *J* = 6.5 Hz, 6H) ppm. <sup>13</sup>C NMR (125 MHz, acetone-*d*<sub>6</sub>):  $\delta$  164.92, 153.87, 134.02, 133.27, 112.43, 112.33, 107.74, 60.76, 47.50, 46.90, 28.09, 20.64 ppm. ESI-MS *m/z* 372.3 [M + H]<sup>+</sup>, 394.2 [M + Na]<sup>+</sup>. Anal. calcd for C<sub>20</sub>H<sub>25</sub>N<sub>3</sub>O<sub>4</sub>: C, 64.67; H, 6.78; O, 17.23. Found: C, 64.61; H, 6.72; O, 17.18.

## ■ ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.9b00502.

<sup>1</sup>H and <sup>13</sup>C NMR, ESI-MS, UV-vis, emission spectra, <sup>1</sup>H NMR titration, selectivity, and photostability tests (PDF)

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### Notes

The authors declare no competing financial interest.

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