



ABSTRACT

Identification of the Sequence Cleavage Preference for the Protease BACE2 Using Proteomics, Mass Spectrometry, and Bioinformatics

Hyojung Kim¹ and Joseph L. Johnson¹

¹Department of Chemistry and Biochemistry, University of Minnesota Duluth

Proteases play critical roles in vital biological processes including the regulation, breakdown, and recycling of proteins. Understanding the preferred amino acid sequence for a given protease can elucidate its substrates and therefore its function. The information from this characterization can be used to develop inhibitors of a protease to treat disease or to suggest potential undesired effects resulting from its inhibition.

β -secretase (BACE) membrane associated aspartyl protease family consists of two homologs, BACE1 and BACE2. BACE1 has been studied in depth as the primary member of this family leading to Alzheimer's disease (AD) while its homolog of BACE2 has been largely ignored because BACE2 mRNA is minimally expressed in the brain. However, recent studies suggest that both BACE1 and BACE2 are involved in the generation of beta-amyloid (A β) which aggregates to form the plaques that are a hallmark pathology of AD. Thus, characterization of the substrate cleavage sequence preference of BACE2 should help to identify putative substrates and its native and AD-related functions.

Proteomic identification of cleavage sites (PICS) is a combined proteomic and mass spectrometric method for characterizing the cleavage site preferences of proteases. It is less biased than previous methods requiring the synthesis of a small number of peptides designed around a specific amino acid sequence. Briefly, peptide fragments generated by the BACE2 cleavage of semi-random peptide libraries are sequenced using mass spectrometry. The remaining portion of recognition and cleavage sequence is determined using bioinformatics and database searching. With the preferred sequence(s) determined, we will then search the human proteome for novel putative substrates of BACE2. In summary, a first step towards a better understanding of BACE2 is to analyze its substrate specificity and then use that information to identify its native substrates which will enable us to elucidate its native and AD-related functions.

DLPC Liposomes Inhibit A β Fibrillation and Remodel Preformed Fibrils Through a Detergent-like Mechanism

Kyle Korshavn¹, Cristina Satriano², Rongchun Zhang³, Mark Dulchavsky⁴, Anirban Bhunia⁵, Magdalena Ivanova⁴, Carmelo La Rosa², Mi Hee Lim⁶, Ayyalusamy Ramamoorthy^{1,3}

¹Department of Chemistry, University of Michigan, Ann Arbor, MI 48109, USA, ²Department of Chemical Sciences, University of Catania, Viale A. Doria 6, 95125 Catania, Italy, ³Biophysics Program, University of Michigan, Ann Arbor, MI 48109, USA, ⁴Department of Neurology, University of Michigan, Ann Arbor, MI 48109, USA, ⁵Department of Biophysics, Bose Institute, Kolkata 700 054, India, ⁶Department of Chemistry, Ulsan National Institute of Science and Technology (UNIST), Ulsan, 44919, Republic of Korea.

The aggregation of amyloid- β (A β) on neuronal lipid bilayers is implicated in the neurotoxicity associated with Alzheimer's disease. It is suggested that the aggregation on lipid bilayers is accelerated. This aggregation, in turn, is capable of disrupting bilayer integrity which induces cell death. We have found, however, that lipid bilayers composed of dilauroyl phosphatidylcholine (DLPC) undergo a unique mechanism of disruption. Through a combination of fluorescence measurements, microscopy, CD spectroscopy, and solid state NMR we have found that DLPC liposomes are rapidly disrupted through interactions with monomeric and early oligomeric forms of Ab. This disruption generates free lipid which successfully impedes amyloid formation through a detergent-like mechanism and stabilizes non-toxic, off pathway oligomers. DLPC liposomes are also capable of remodeling preformed fibrils of Ab into on-pathway, toxic protofibrillar species. All previous evidence of membrane disruption by A β has



ABSTRACT

been the result of fibrillation, pointing to a unique interaction occurring at the surface of DLPC membranes which is capable of halting amyloid formation in the early stages.

Supermetallization of Peptides and Proteins Studied by High Resolution Mass Spectrometry

Eugeny Kukaev^{1, 3, 4}, Yury Kostyukevich^{1, 2}, Alexey Kononikhin^{1, 3}, Maria Indeykina^{3, 4}, Igor Popov^{1,4}, Eugene Nikolaev^{1, 2, 3, 4}

¹Moscow Institute of Physics and Technology, Moscow, ²Skolkovo Institute of Science and Technology, Skolkovo, ³Institute for Energy Problems of Chemical Physics, Moscow, ⁴Emanuel Institute of Biochemical Physics, Moscow, Russia

Transition metal ions are involved in neurodegenerative diseases.

Recently we have reported the supermetallization phenomenon, which is the formation of complex ions of peptide-metal in the gas phase when the peptide accepts an unexpectedly large number of metal atoms. This supermetallization takes place during electrospray ionization when charged droplets are evaporating at relatively high temperature (ca 400°C). The effect has been demonstrated for the divalent Zn²⁺ atom and several peptides and proteins. It is observed that small protein ubiquitin can incorporate up to 20 Zn atoms and the attachment of each Zn requires the removal of two hydrogen atoms. Here we report the first observation, to our knowledge, of gas-phase complexes of peptides and proteins with tetravalent metals.

Supermetallized complexes were studied by FTICR MS in combination with CID/ECD fragmentation. It was found that during ECD complexes with many metal atoms do not dissociate and only decrease their charge by capturing the electron. Using high resolution FTICR it was observed that with the increase of the number of metal atoms in the supermetallized complex there is a trend toward forming the z+1 fragments instead of the z fragments in ECD experiments. The effect of hydrogen rearrangement for c fragments was also observed but was found to be very weak.

This work was supported by the project of Russian Science Foundation 16-14-00181.

References:

1. Y. Kostyukevich et al. *J. Mass Spectrom.* 50, 1079–1087 (2015).
2. Y. Kostyukevich et al. *Eur. J. Mass Spectrom.* 22,39–42 (2016).

Key asparagine and glutamine residues promote cross-species prion conversion

Timothy D. Kurt¹, Lin Jiang², Nazilla Alderson¹, Jun Liu¹, David Eisenberg², and Christina J. Sigurdson^{1,3}

¹Departments of Pathology and Medicine, UC San Diego, La Jolla, CA 92093, USA, ²UCLA-DOE Institute, Howard Hughes Medical Institute, and Molecular Biology Institute, UCLA, Los Angeles, California 90095, USA, ³Department of Pathology, Immunology, and Microbiology, UC Davis, Davis, CA 95616, USA

The central event underlying prion disease transmission is the conversion of the cellular prion protein, Pr^{PC}, into a misfolded, self-templating conformer called Pr^{Sc}. Sequence similarity between Pr^{Sc} and Pr^{PC} is critical for efficient prion templating, yet the specific interacting residues and the mechanism by which a residue promotes or inhibits conversion remain unclear. Previous studies have shown that single residue differences between Pr^{Sc} and Pr^{PC} may dramatically impede conversion, as illustrated by the G127V variant of human Pr^{PC}, which confers protection against transmission of human prions (1). However, the transmission of prions between species suggests that many Pr^{PC} residue differences are not protective, or may even promote prion cross-seeding. Recent studies have found that the Pr^{PC} sequence of bank voles, a European rodent, is permissive to conversion by a wide array of prions from many species (2, 3) despite residue differences between the Pr^{Sc} and vole Pr^{PC} sequences. We recently demonstrated that human Pr^{PC} having only two bank vole amino acid substitutions (E168Q