



## Article

# The Relationship between Telomere Length and Gestational Weight Gain: Findings from the Mamma & Bambino Cohort

Andrea Maugeri <sup>1</sup>, Roberta Magnano San Lio <sup>1</sup>, Maria Clara La Rosa <sup>1</sup>, Giuliana Giunta <sup>2</sup>, Marco Panella <sup>2</sup>, Antonio Cianci <sup>2</sup>, Maria Anna Teresa Caruso <sup>3</sup>, Antonella Agodi <sup>1,\*</sup> and Martina Barchitta <sup>1</sup>

<sup>1</sup> Department of Medical and Surgical Sciences and Advanced Technologies “GF Ingrassia”, University of Catania, Via S. Sofia, 87, 95123 Catania, Italy; andrea.maugeri@unict.it (A.M.); roberta.magnanosanlio@phd.unict.it (R.M.S.L.); mariaclara.larosa@unict.it (M.C.L.R.); martina.barchitta@unict.it (M.B.)

<sup>2</sup> Obstetrics and Gynecology Unit, Department of General Surgery and Medical Surgical Specialties, University of Catania, Via S. Sofia, 78, 95123 Catania, Italy; giunta.giuliana@studium.unict.it (G.G.); mpanella@unict.it (M.P.); acianci@unict.it (A.C.)

<sup>3</sup> Cytogenetic Laboratory, Azienda Ospedaliero Universitaria Policlinico “G. Rodolico—San Marco”, Via S. Sofia, 78, 95123 Catania, Italy; m.caruso@ao-ve.it

\* Correspondence: agodia@unict.it

**Abstract:** Inadequate gestational weight gain (GWG) affects a growing number of pregnancies, influencing intrauterine environment and long-term health. Uncovering molecular mechanisms associated with GWG could be helpful to develop public health strategies for tackling this issue. Here, our study aimed to understand the relationship of DNA telomere length with weight gain during pregnancy, using data and samples from the ongoing prospective “Mamma & Bambino” study (Catania, Italy). GWG was calculated according to the Institute of Medicine (IOM) guidelines. Relative telomere length was assessed by real-time quantitative polymerase chain reaction in 252 samples of maternal leucocyte DNA (mlDNA) and 150 samples of cell-free DNA (cfDNA) from amniotic fluid. We observed that relative telomere length of mlDNA seemed to weakly increase with GWG. In contrast, telomere length of cfDNA exhibited a U-shaped relationship with GWG. Women with adequate GWG showed longer telomere length than those who gained weight inadequately. Accordingly, the logistic regression model confirmed the association between telomere length of cfDNA and adequate GWG, after adjusting for potential confounders. Our findings suggest an early effect of GWG on telomere length of cfDNA, which could represent a molecular mechanism underpinning the effects of maternal behaviours on foetal well-being.

**Keywords:** pregnancy; aging; telomere; weight gain; body mass index



**Citation:** Maugeri, A.; Magnano San Lio, R.; La Rosa, M.C.; Giunta, G.; Panella, M.; Cianci, A.; Caruso, M.A.T.; Agodi, A.; Barchitta, M. The Relationship between Telomere Length and Gestational Weight Gain: Findings from the Mamma & Bambino Cohort. *Biomedicines* **2022**, *10*, 67. <https://doi.org/10.3390/biomedicines10010067>

Academic Editor: Yegor E. Yegorov

Received: 28 November 2021

Accepted: 28 December 2021

Published: 30 December 2021

**Publisher’s Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Gestational weight gain (GWG)—which depends on body composition, weight of the foetus, placenta and amniotic fluid [1]—represents a natural response to host the growing foetus. The US-based Institute of Medicine (IOM) has promoted the development of recommendations to identify an optimal amount of GWG according to the pre-pregnancy Body Mass Index (BMI) [1–5]. However, more than half of pregnant women do not respect clinical practice guidelines for weight gain [6–8]. The adherence to these guidelines is crucial to reduce the risk of adverse outcomes for mothers and their newborns [9–13]. With respect to IOM guidelines, both greater and lower weight gain contribute to short- and long-term health complications [2,14–16]. For instance, excessive GWG is associated with an increased risk of high blood pressure [17], diabetes [18], caesarean section [19], postpartum weight retention [20] and obesity [12]. As regard newborns born from mothers who gained weight excessively, they are more likely to be large for gestational age [18,21,22] and to develop metabolic disorders during adolescence and adulthood [23,24]. By contrast,

low GWG is associated with increasing risks of pre-term delivery [25]. For all these reasons, there is the need for uncovering molecular mechanisms associated with GWG to identify mothers who could benefit more from preventive strategies.

In this scenario, telomere length represents a promising biomarker for biological aging and age-related diseases. Telomeres are repeating DNA sequences at the ends of chromosomes that progressively shorten with cell division [26]. To maintain genomic stability, telomeres protect the chromosomes from DNA damage and shorter telomeres are considered as a marker of the cumulative damage to which cells have been exposed [26]. Telomere integrity is maintained by the specific activity of telomerase, characterized by an enzymatic protein component that adds the telomeric DNA repeats at the end of chromosomes, and a telomerase RNA template for telomeric DNA synthesis [27]. In adults, shorter telomeres are associated with diabetes [28], cancer [29] and cardiovascular disease [30]. However, also before the onset of age-related diseases, obesity might contribute to cumulative burden of oxidative stress and chronic inflammation, accelerating the telomere shortening process. During pregnancy, adverse exposures such as maternal stress, smoking and higher levels of air pollution are associated with shorter telomeres measured in cord blood [31–33], placenta [34] and other children’s samples [35]. In particular, it has been demonstrated that maternal pre-pregnancy BMI was associated with telomere shortening in cord blood and placenta [36]. Some studies evaluated the association of telomere length in cord blood with preterm birth [37] and birth weight [38,39]. Moreover, since telomerase activity is time- and location-regulated in both embryo and placental tissues, associations between telomerase activity and pregnancy complications—such as intrauterine growth restriction—have been previously observed [40]. Circulating cell-free DNA (cfDNA) in plasma and serum has been proposed as a novel biomarker for prenatal diagnosis [41,42] and with applications in oncology [43,44]. Moreover, the relationship between cfDNA in serum and several diseases has raised much interest in investigating the role of cfDNA in other body fluids, such as urine [45], saliva [46] and amniotic fluid [47]. In line, amniotic fluid has been proposed as an alternative source of potential biomarkers for prenatal diagnosis [48]. Indeed, amniotic fluid surrounds the foetus with a continuous exchange with foetal organs and gestational tissues [49–51]. After removing the cellular components, the cell-free supernatant that remains reflects maternal and foetal well-being [51–59]. In particular, amniotic fluid contains a greater amount of cell-free foetal- and pregnancy-related DNA than maternal serum [60–63].

With these thoughts in mind, in order to study the relationship between telomere length and weight gain during pregnancy we used data and samples from the ongoing prospective “Mamma & Bambino” study, which enrolls mother–child pairs from Catania, Italy. Here, we report findings about the relationship between GWG and telomere length in maternal leucocyte DNA (mlDNA) and cfDNA of amniotic fluid.

## 2. Materials and Methods

### 2.1. Study Design

This study was conducted on samples and data obtained from mother–child pairs of the “Mamma & Bambino” cohort. It is an ongoing prospective study research with the aim of evaluating how the exposome affects mothers’ and children’s health [64–67]. Full details and protocols of the “Mamma & Bambino” cohort are described elsewhere [64–67]. In brief, the cohort consists of pregnant women attending for prenatal genetic counselling at the Azienda Ospedaliero Universitaria Policlinico “G. Rodolico -San Marco” (Catania, Italy). Those with multiple pregnancy, autoimmune and chronic diseases, pregnancy complications, intrauterine foetal death and congenital malformations are excluded. Women are generally recruited from 4 to 20 weeks of gestation and the study entails planned follow-ups at delivery and after two years birth. In the current analysis, we used data and samples from mothers who completed singleton pregnancy and with available data on GWG at delivery.

## 2.2. Assessment of Gestational Weight Gain

At recruitment, women were asked to report their height and pre-pregnancy weight to calculate pre-pregnancy body mass index (BMI) as  $\text{kg}/\text{m}^2$ . Women were classified as underweight, normal weight, overweight or obese based on their pre-pregnancy BMI, according to WHO criteria [68]. Firstly, maternal weight achieved at recruitment was calculated by subtracting the self-reported pre-pregnancy weight from the weight at recruitment. Next, maternal weight achieved at delivery was collected from clinical records and total GWG was calculated by subtracting the self-reported pre-pregnancy weight from the weight at delivery. As described by the IOM guidelines [6], GWG was classified as reduced, adequate, or excessive according to pre-pregnancy BMI.

## 2.3. Covariate Ascertainment

Beyond anthropometric measures, our analysis considered several covariates that might affect GWG, telomere length and their relationship. At recruitment, demographics, socio-economic information and lifestyles were assessed by trained epidemiologists through structured questionnaires [64–67,69–71]. Maternal age and gestational ages at recruitment and at delivery were considered because of their potential effect on sampling and telomere length. In addition, we used the educational level and employment status as two proxy indicators of socio-economic status. Educational level was classified as low (having primary education), medium (having secondary education) or high (having tertiary education). Employment status was categorized as employment or unemployment, which also included students and housewives. We also categorized women in those who have previously had at least a child and those who have not. Regarding lifestyles, we assessed smoking status, daily energy intake and adherence to the Mediterranean Diet (MD). Specifically, dietary data were collected using a 95-item semiquantitative Food Frequency Questionnaire (FFQ). This tool was referred to 30 days before recruitment [72] and, for each item, it asked to report both frequency of consumption and portion size. Daily energy intake was calculated considering the table of food composition released by the US Department of Agriculture and adapted to typical Italian foods. Adherence to MD was evaluated using the Mediterranean Diet Score, as described in detail elsewhere [73].

## 2.4. DNA Extraction

Biological samples included maternal blood obtained at recruitment and an aliquot of amniotic fluid from women who underwent amniocentesis. Full details on protocols of DNA extraction are reported elsewhere [67,71]. Genomic mlDNA was extracted from 200  $\mu\text{L}$  of maternal blood, while cfDNA was extracted from the supernatant of amniotic fluid obtained after centrifugation. DNA extraction and purification were performed using the QIAamp Blood Kit (Qiagen, Milan, Italy) on the QIAcube instrument (Qiagen, Milan, Italy), as described by the manufacturer's protocol. Concentration and purity of DNA were assessed by NanoDrop 1000 spectrometer and by Qubit 3.0 Fluorometer using dsDNA HS Assay Kit (Thermo Fisher Scientific, Carlsbad, CA, USA).

## 2.5. Relative Telomere Length

Relative telomere length was measured by real-time quantitative polymerase chain reaction (qPCR), using the Relative Human Telomere Length Quantification Assay Kit (ScienCell Research Laboratories, Carlsbad, CA, USA). The qPCR was performed on a QuantStudio 7 Flex Real-Time PCR System (Thermo Fisher Scientific, Carlsbad, CA, USA), according to the manufacturer's protocol. The following sets of primers were used: the telomere (T) primer set amplified telomere sequences; the single-copy reference (S) primer set amplified a 100 bp-long region on human chromosome 17 and was used as reference for data normalization. The specificity of these primer sets was validated by the manufacturer through qPCR with melt curve analysis. Each reaction contained 1  $\mu\text{L}$  of DNA (5 ng/ $\mu\text{L}$ ), 2  $\mu\text{L}$  of primer solution (telomere or SCR), 10  $\mu\text{L}$  of 2X GoldNStart TaqGreen qPCR master mix (ScienCell Research Laboratories, Carlsbad, CA, USA) and 7  $\mu\text{L}$  of nuclease-free water.

The PCR conditions were as follows: denaturation (95 °C for 10 min); 32 cycles of 95 °C for 20 s, 52 °C for 20 s and 72 °C for 45 s. The qPCR was calibrated including in each plate a serial dilution of DNA from randomly selected samples. All reactions were run in duplicate and relative telomere length was expressed as the average of telomere/single copy reference (T/S) ratio.

### 2.6. Statistical Analysis

Statistical analyses were performed using SPSS v.25. Descriptive statistics was initially performed using frequencies (percentage, %) or median and interquartile range (IQR) due to the skewness of quantitative variables. Bivariate analyses were conducted using the Mann–Whitney or the Kruskal–Wallis tests for quantitative variables and the Chi-squared test for trend for categorical variables. Relative telomere length was also plotted against weight gain at recruitment and at delivery to inspect linear or non-linear relationships. Next, we plotted relative telomere length by the tertile distribution of GWG, as well as by its classification in reduced, adequate, or excessive. We also applied a logistic regression model using adequate GWG as dependent variables and the following covariates: relative telomere length, maternal age, gestational age at recruitment, educational level, having children, pre-pregnancy BMI, total daily energy intake and gestational age at delivery. The adjusted association of relative telomere length with adequate GWG was reported as  $\beta$  coefficient and its Standard error (SE). All tests were two-sided and performed at a significance level  $\alpha = 0.05$ .

## 3. Results

### 3.1. Characteristics of Study Population

The current analysis included 270 mothers who completed singleton pregnancy, with available information on GWG at delivery. Table 1 describes the characteristics of the study population and their comparison across categories of GWG (i.e., 101 women gained weight adequately, while 91 and 78 reported reduced and excessive GWG, respectively). As expected, there were strong relationships between maternal anthropometric measures and GWG. Indeed, women with adequate GWG were those with the lowest pre-pregnancy weight and BMI. By contrast, weight at delivery increased from reduced to excessive GWG categories. A similar trend was observed for education, so that the proportion of women with low or medium educational level increased from reduced to excessive GWG. Moreover, the proportion of women who already had at least one child before the current study was higher in those with adequate GWG than their counterparts. With respect to dietary habits, we did not find any association with adherence to Mediterranean Diet (MD), but total daily energy intake increased across GWG categories.

### 3.2. Relationship of Telomere Length with Maternal Characteristics

Among recruited women, we collected 252 maternal blood samples used to analyse telomere length of mlDNA. Notably, relative telomere length of mlDNA did not correlate with maternal age, pre-pregnancy BMI, total energy intake, Mediterranean Diet Score (MDS) and gestational age at sampling and at delivery ( $p$ -values  $> 0.05$ ). Moreover, relative telomere length did not differ across categories of educational level, employment, smoking status, parity and pre-pregnancy BMI ( $p$ -values  $> 0.05$ ).

We also obtained 150 samples of amniotic fluid from those who underwent amniocentesis. These samples were used to evaluate telomere length of cfDNA. Relative telomere length of cfDNA and mlDNA did not correlate with each other ( $p > 0.05$ ). On the contrary, relative telomere length of cfDNA was negatively but weakly correlated with gestational age at sampling (Spearman coefficient =  $-0.152$ ;  $p = 0.046$ ) and positively with total energy intake (Spearman coefficient =  $0.157$ ;  $p = 0.038$ ). No correlations were evident with the remaining maternal characteristics, as well as with birth length and birth weight ( $p$ -values  $> 0.05$ ). Relative telomere length did not also differ across categories of educa-

tional level, employment, smoking status, parity, pre-pregnancy BMI, type of delivery and newborn gender ( $p$ -values > 0.05).

**Table 1.** Characteristics of women from the “Mamma & Bambino” cohort ( $n = 270$ ) according to gestational weight gain categories.

Characteristics	Overall ( $n = 270$ )	Reduced GWG ( $n = 91$ )	Adequate GWG ( $n = 101$ )	Excessive GWG ( $n = 78$ )	$p$ -Value <sup>a</sup>
Age <sup>b</sup>	37.0 (4.0)	37.0 (4.0)	38.0 (4.0)	37.0 (4.0)	0.699
Gestational age at sampling <sup>b</sup>	16.0 (4.0)	16.0 (4.0)	16.0 (3.0)	16.0 (2.0)	0.953
Educational level (%)					
Low	17.8%	16.5%	16.8%	20.5%	
Medium	47.8%	40.7%	45.5%	59.0%	0.038
High	34.4%	42.8%	37.7%	20.5%	
Working (%)					
Employment	57.4%	54.9%	61.4%	55.1%	
Unemployment	42.6%	45.1%	38.6%	44.9%	0.593
Smokers (%)	20.5%	15.4%	20.0%	27.3%	0.216
Having children (% yes)	67.7%	64.3%	76.8%	59.7%	0.041
Total energy intake <sup>b</sup>	1750 (620)	1667 (674)	1752 (545)	1858 (596)	0.045
MDS <sup>b</sup>	4.0 (2.0)	4.0 (2.0)	4.0 (2.0)	4.0 (2.0)	0.102
Pre-pregnancy weight <sup>b</sup>	61.0 (15.2)	62.0 (16.0)	59.0 (13.0)	64.5 (18.3)	0.012
Pre-pregnancy BMI <sup>b</sup>	22.8 (5.1)	22.8 (4.8)	22.0 (3.8)	25.0 (5.7)	0.002
Pre-pregnancy BMI categories					
Underweight	6.7%	6.6%	6.9%	6.4%	
Normal weight	64.1%	68.1%	77.2%	42.3%	
Overweight	17.4%	9.9%	8.9%	37.2%	<0.001
Obese	11.9%	15.4%	7.0%	14.1%	
Weight at delivery <sup>b</sup>	74.0 (15.0)	68.5 (11.5)	73.0 (12.7)	82.0 (15.2)	<0.001
Gestational age at delivery <sup>b</sup>	39.0 (2.0)	38.0 (2)	39.0 (2)	39.0 (2.0)	0.383

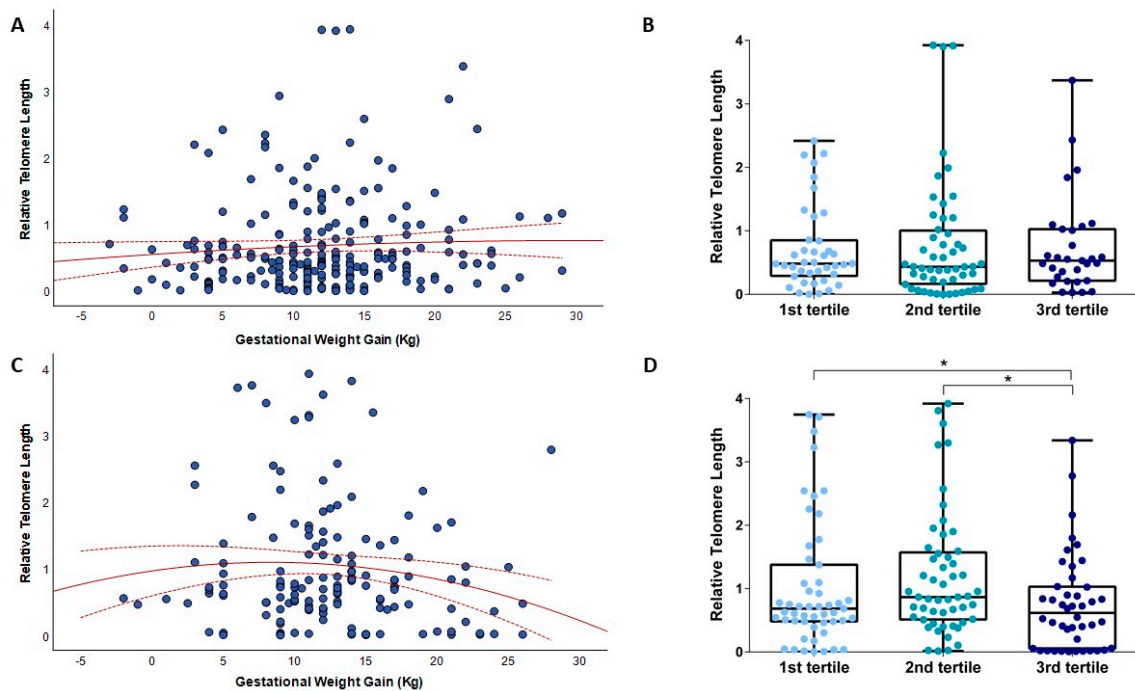
<sup>a</sup>  $p$ -values are based on the Kruskal–Wallis test for quantitative variables, or chi-squared test for categorical variables <sup>b</sup> Data are reported as median and interquartile range (IQR). Abbreviations: GWG, gestational weight gain; MDS, Mediterranean Diet Score; BMI, body mass index.

### 3.3. Relationships between Gestational Weight Gain and Telomere Length

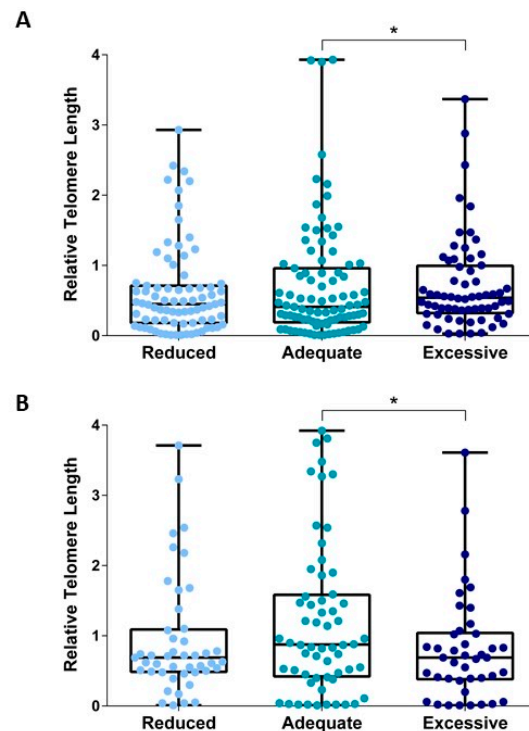
We next evaluated the relationship of relative telomere length in mlDNA and cfDNA with gestational weight gain (Figure 1). To do that, we also classified women according to the tertile distribution of GWG: first tertile from  $-2$  to  $9$  Kg; second tertile from  $10$  to  $13$  Kg; third tertile from  $14$  to  $28$  Kg. As depicted in Figure 1A, relative telomere length of mlDNA seemed to weakly increase with GWG. However, Figure 1B did not show a significant difference according to the tertile distribution of GWG ( $p = 0.559$ ). By contrast, Figure 1C suggested a U-shaped relationship between GWG and relative telomere length of cfDNA. The U-shaped relationship was confirmed by the comparison of relative telomere length across tertiles of GWG ( $p = 0.016$ ; Figure 1D). In particular, women in the third tertile showed shorter relative telomere length than those in the second tertile ( $p = 0.014$ ; Figure 1D).

We next compared relative telomere length across categories of GWG that considered reduced, adequate, or excessive weight gain during pregnancy. Regarding mlDNA, we showed longer telomere length in women with excessive GWG that in those who gained weight adequately ( $p = 0.017$ ; Figure 2A). By contrast, telomere length of cfDNA was lower in amniotic fluid from women with reduced or excessive GWG, if compared with those who gained weight adequately (Figure 2B). Yet, the difference was statistically significant for women with excessive GWG ( $p = 0.044$ ) but not for those with reduced GWG ( $p = 0.117$ ; Figure 2B). It is also worth mentioning that the relationship was already evident if considering weight gain at recruitment (Figure 3).

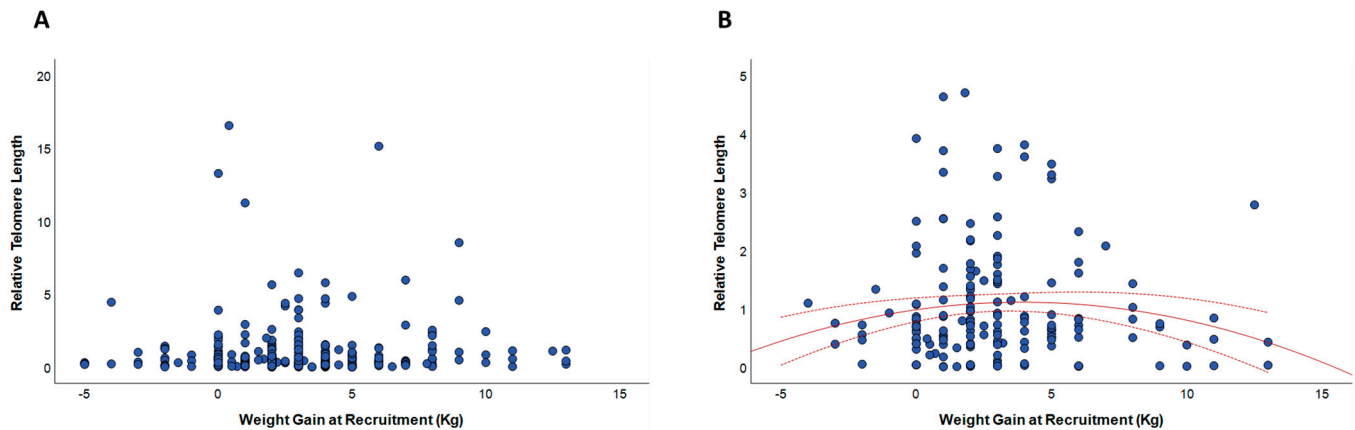




**Figure 1.** The relationship of gestational weight gain with relative telomere length. (A) shows the relationship of gestational weight gain with telomere length of mDNA; (B) shows the box plots of telomere length of mDNA by the tertile distribution of GWG; (C) shows the relationship of gestational weight gain with telomere length of cfDNA from amniotic fluid; (D) shows the box plots of telomere length of cfDNA by the tertile distribution of GWG. \*  $p$ -value  $< 0.05$  based on the Mann-Whitney or Kruskal-Wallis test.

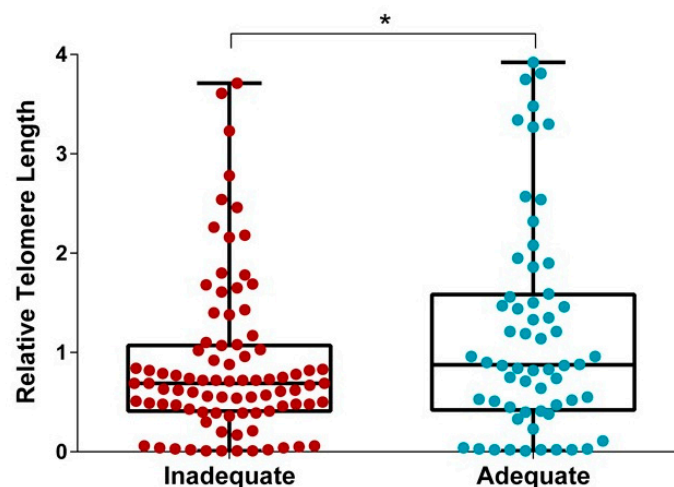


**Figure 2.** The relationship between categories of gestational weight gain and relative telomere length. (A) shows the box plots of telomere length of mDNA according to GWG categories; (B) shows the box plots of telomere length of cfDNA according to GWG categories. \*  $p$ -value  $< 0.05$  based on the Mann-Whitney test.



**Figure 3.** The relationship of weight gain at recruitment with relative telomere length. (A) shows the relationship of weight gain at recruitment with telomere length of mDNA; (B) shows the relationship of weight gain at recruitment with telomere length of cfDNA from amniotic fluid.

We next compared relative telomere length between women who gained weight adequately and those who did not. This comparison showed higher relative telomere length of cfDNA in women with adequate GWG ( $p = 0.017$ ; Figure 4). Finally, we applied a logistic regression model including other maternal characteristics (i.e., age, gestational age at sampling, educational level, parity, pre-pregnancy BMI, total daily energy intake, gestational age at delivery) that might affect telomere length and/or GWG. Interestingly, the association between cfDNA telomere length and adequate GWG was significant, after adjusting for the abovementioned maternal characteristics ( $\beta = 0.464$ ; SE = 0.189;  $p = 0.014$ ).



**Figure 4.** The comparison of telomere length of cfDNA from amniotic fluid between adequate and inadequate gestational weight gain. \*  $p$ -value < 0.05 based on the Mann–Whitney U Test.

#### 4. Discussion

In the present study, we show for the first time a link between relative telomere length and GWG, though the shape of the relationship depends on DNA source. In particular, we observed a U-shaped relationship when analysing cfDNA in amniotic fluid, with longer relative telomere length in samples from mothers who gained weight adequately. To place our results in context, it is worth mentioning that a meta-analysis of nearly 120,000 subjects suggested an inverse association between obesity and telomere length [29]. This is in line with a collaborative cross-sectional meta-analysis of 87 observational studies and 146,114 individuals, showing a 3.99 bp decrease in telomere length for each unit increase in BMI [74]. However, these analyses indicated a high degree of heterogeneity across studies and an overall lack of evidence on pregnant women [29,74]. This heterogeneity could, at

least in part, explained by the effect of chronological age on telomere length. In fact, it has been greatly demonstrated how telomere length decreased with increasing chronological age [75,76]. However, in our study, we did not find a correlation between maternal age and telomere length of mtDNA and cfDNA. Yet, during the gestational period, maternal body could experience changes at cellular and molecular levels that might prevent the relationship between age and telomere length [77]. For instance, it has been proposed that gestational age, rather than chronological age, could influence telomere shortening in placental DNA [78]. Moreover, other factors could interact with and/or mediate this relationship.

To our knowledge, only few studies investigated the effect of maternal pre-pregnancy BMI on telomere length, and none focused on maternal GWG. For instance, Martens and colleagues reported a decline in newborns' telomere length with increasing maternal pre-pregnancy BMI, as assessed in both cord blood and placental tissues [36]. Interestingly, this effect seemed to persist in childhood, as demonstrated by Clemente and colleagues [79]. The authors used data from the Human Early-Life Exposome (HELIX) study to demonstrate that child's leukocyte telomere length decreased with increasing maternal pre-pregnancy BMI [79]. However, the role of telomerase in the association between increased BMI and shortened telomere length is not well investigated. Epel and colleagues described reduced telomerase activity with increasing BMI in healthy women, which may be an important factor for the observed relationship between shorter telomeres and body weight. Further research is needed to evaluate whether the relationship between telomerase activity and BMI in pregnant women could be associated with altered neonatal telomerase activity [80].

We add to this knowledge, suggesting the influence of maternal weight gain during pregnancy on telomere length of cfDNA from amniotic fluid. The observed difference, though small, could add motivations to keep studying the effect of weight gain on aging biomarkers. Of note, the relationship remained significant after controlling for the potential effects of covariates (i.e., age, gestational ages at recruitment and at delivery, educational level, previous pregnancies, pre-pregnancy BMI and total daily energy intake). Interestingly, the effect of maternal weight gain on telomere length of cfDNA was already evident in early pregnancy (i.e., considering GWG at a median gestational age of 16 weeks). This was consistent with the Developmental Origins of Health and Disease (DOHaD) hypothesis, for which prenatal environment programs the foetus for challenges that it is likely to experience after birth [81]. For instance, it has been proposed a potential relationship between epigenetic mechanisms (i.e., DNA methylation) and telomere attrition rate in early life, which in turn could be influenced by internal and external stressors [82]. However, it is necessary to investigate if even small differences in telomere length could be associated with pregnancy and neonatal outcomes. In this scenario, identifying activators of telomerase that could complement the benefits of a healthy lifestyle will be an important field of research in the ongoing evaluation of the telomere system [83].

In line, there is the current need for developing non-invasive tests to understand foetal well-being. These tests should be based on maternal serum or urine, avoiding invasive tests such as amniocentesis. Yet, more than 80% of cfDNA fragments in the maternal serum are short and fragmented [84]. Compared to maternal serum, amniotic fluid contains a much greater concentration of cfDNA [85], which is largely uncontaminated by maternal- and trophoblast-derived nucleic acids. Thus, amniotic fluid represents a relatively pure foetal sample and its supernatant is a valuable and widely available but under-utilized resource [86]. It was not our intention to prefer amniotic fluid over maternal blood for the analysis of relative telomere length. However, in future, it will be interesting to evaluate if foetal DNA from maternal blood reflects the same difference observed in our study.

Our findings also provide further motivation to study telomere length and telomerase activity as potential molecular mechanisms underpinning the effects of maternal behaviours on the development of chronic disease later in life. Although mechanisms by which inadequate weight gain affects telomere length are not yet fully understood, it is plausible that they rely on a chronic inflammatory and oxidative state in utero [87]. Despite this



speculation, however, we currently need more experimental work to better understand how maternal weight gain affects telomere dynamics in the foetus.

Our study had some limitations that should be considered. Firstly, the information on pre-pregnancy weight was self-reported, which cannot completely exclude a potential reporting bias. It is also true, however, that previous studies demonstrated how self-reported pre-pregnancy weight correlated with that measured [88,89]. Secondly, we worked on total GWG reached at delivery without accounting for weight trajectories throughout pregnancy. Yet, our analysis at the time of recruitment already showed an effect of maternal weight gain on telomere length of cfDNA. Although we observed an early influence of GWG on telomere length, our analysis did not focus on their causal–effect relationship. Thirdly, although amniotic fluid seems a relatively pure foetal sample, a low proportion of cfDNA from placenta cannot be completely excluded [90]. Moreover, we assessed relative telomere length by qPCR, which has higher assay variability than terminal restriction fragment analysis [91]. Finally, the presence of residual confounders cannot be completely ruled out, such as that deriving from fatherly influence [92,93].

## 5. Conclusions

In conclusion, we found that relative telomere length of cfDNA is associated with maternal weight gain during pregnancy and at delivery. This suggests an early influence of GWG on telomere length, which could represent a molecular mechanism underpinning the effects of maternal behaviours on foetal well-being. However, further experimental studies are needed to biological events that regulate this relationship and to consider other factors influencing the uterine environment during pregnancy.

**Author Contributions:** Conceptualization, A.M., R.M.S.L., A.A. and M.B.; software, A.M. and R.M.S.L.; formal analysis, A.M., R.M.S.L., M.C.L.R., G.G., M.P., A.C., M.A.T.C. and M.B.; resources, A.A.; data curation, A.M., R.M.S.L., M.C.L.R. and M.B.; writing—original draft preparation, A.M. and R.M.S.L.; writing—review and editing, all the Authors; visualization, A.M. and R.M.S.L.; supervision, A.A. All authors have read and agreed to the published version of the manuscript.

**Funding:** The research “La coorte Mamma & Bambino: un approccio Multisetoriale Alla salute Materno-Infantile Mediante valutazione dell’Esposoma nelle Donne, MAMI-MED” was funded by the University of Catania, Italy, Department of Medical and Surgical Science and Advanced Technologies “GF Ingrassia” (Programma ricerca di ateneo UNICT 2020-22 linea 2, PIAno di inCEntivi per la Ricerca di Ateneo 2020/2022). This work was also supported by the Department of Medical and Surgical Science and Advanced Technologies “GF Ingrassia”, University of Catania, Italy (Piano Triennale di sviluppo delle Attività di Ricerca Scientifica del Dipartimento 2016–2018).

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Ethics Committee of Azienda Ospedaliero-Universitaria Policlinico-Vittorio Emanuele” and Ethics Committee “Catania 1” with the following protocol numbers: 47/2014/VE; 48/2015/EMPO; 186/2015/EMPO; 197/2016/EMPO; 213/2017/EMPO; 231/2018/EMPO; 263/2019/EMPO.

**Informed Consent Statement:** Informed consent was obtained from all subjects involved in the study.

**Data Availability Statement:** The datasets analysed during the current study are available from the corresponding author on reasonable request.

**Acknowledgments:** We are grateful to all women who gave their consent to participate in the study.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Institute of Medicine Guidelines. *Weight Gain During Pregnancy: Reexamining the Guidelines*; Institute of Medicine Guidelines: Washington, DC, USA, 2009.
2. Cedergren, M.I. Optimal gestational weight gain for body mass index categories. *Obstet. Gynecol.* **2007**, *110*, 759–764. [[CrossRef](#)] [[PubMed](#)]

3. Beyerlein, A.; Schiessl, B.; Lack, N.; von Kries, R. Optimal gestational weight gain ranges for the avoidance of adverse birth weight outcomes: A novel approach. *Am. J. Clin. Nutr.* **2009**, *90*, 1552–1558. [[CrossRef](#)] [[PubMed](#)]
4. Cheikh Ismail, L.; Bishop, D.C.; Pang, R.; Ohuma, E.O.; Kac, G.; Abrams, B.; Rasmussen, K.; Barros, F.C.; Hirst, J.E.; Lambert, A.; et al. Gestational weight gain standards based on women enrolled in the Fetal Growth Longitudinal Study of the INTERGROWTH-21st Project: A prospective longitudinal cohort study. *BMJ* **2016**, *352*, i555. [[CrossRef](#)]
5. Rasmussen, K.M.; Catalano, P.M.; Yaktine, A.L. New guidelines for weight gain during pregnancy: What obstetrician/gynecologists should know. *Curr. Opin. Obstet. Gynecol.* **2009**, *21*, 521–526. [[CrossRef](#)] [[PubMed](#)]
6. Chung, J.G.; Taylor, R.S.; Thompson, J.M.; Anderson, N.H.; Dekker, G.A.; Kenny, L.C.; McCowan, L.M.; Consortium, S. Gestational weight gain and adverse pregnancy outcomes in a nulliparous cohort. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2013**, *167*, 149–153. [[CrossRef](#)] [[PubMed](#)]
7. Durie, D.E.; Thornburg, L.L.; Glantz, J.C. Effect of second-trimester and third-trimester rate of gestational weight gain on maternal and neonatal outcomes. *Obstet. Gynecol.* **2011**, *118*, 569–575. [[CrossRef](#)]
8. Simas, T.A.; Liao, X.; Garrison, A.; Sullivan, G.M.; Howard, A.E.; Hardy, J.R. Impact of updated Institute of Medicine guidelines on prepregnancy body mass index categorization, gestational weight gain recommendations, and needed counseling. *J. Womens Health (Larchmt)* **2011**, *20*, 837–844. [[CrossRef](#)]
9. Cedergren, M. Effects of gestational weight gain and body mass index on obstetric outcome in Sweden. *Int. J. Gynaecol. Obstet.* **2006**, *93*, 269–274. [[CrossRef](#)] [[PubMed](#)]
10. Johnston, J.C.; McNeil, D.A.; Best, M.; MacLeod, C. A growth status measurement pilot in four Calgary area schools: Perceptions of grade 5 students and their parents. *J. Sch. Nurs.* **2011**, *27*, 61–69. [[CrossRef](#)]
11. Nohr, E.A.; Vaeth, M.; Baker, J.L.; Sørensen, T.I.; Olsen, J.; Rasmussen, K.M. Pregnancy outcomes related to gestational weight gain in women defined by their body mass index, parity, height, and smoking status. *Am. J. Clin. Nutr.* **2009**, *90*, 1288–1294. [[CrossRef](#)]
12. Rooney, B.L.; Schauburger, C.W.; Mathiason, M.A. Impact of perinatal weight change on long-term obesity and obesity-related illnesses. *Obstet. Gynecol.* **2005**, *106*, 1349–1356. [[CrossRef](#)]
13. Rooney, B.L.; Schauburger, C.W. Excess pregnancy weight gain and long-term obesity: One decade later. *Obstet. Gynecol.* **2002**, *100*, 245–252. [[CrossRef](#)] [[PubMed](#)]
14. Ferraro, Z.M.; Contador, F.; Tawfiq, A.; Adamo, K.B.; Gaudet, L. Gestational weight gain and medical outcomes of pregnancy. *Obstet. Med.* **2015**, *8*, 133–137. [[CrossRef](#)] [[PubMed](#)]
15. DeVader, S.R.; Neeley, H.L.; Myles, T.D.; Leet, T.L. Evaluation of gestational weight gain guidelines for women with normal prepregnancy body mass index. *Obstet. Gynecol.* **2007**, *110*, 745–751. [[CrossRef](#)] [[PubMed](#)]
16. Margerison Zilko, C.E.; Rehkopf, D.; Abrams, B. Association of maternal gestational weight gain with short- and long-term maternal and child health outcomes. *Am. J. Obstet. Gynecol.* **2010**, *202*, 574.e1–574.e8. [[CrossRef](#)]
17. de la Torre, L.; Flick, A.A.; Istwan, N.; Rhea, D.; Cordova, Y.; Dieguez, C.; Desch, C.; González-Quintero, V.H. The effect of new antepartum weight gain guidelines and prepregnancy body mass index on the development of pregnancy-related hypertension. *Am. J. Perinatol.* **2011**, *28*, 285–292. [[CrossRef](#)]
18. Mamun, A.A.; Kinarivala, M.; O’Callaghan, M.J.; Williams, G.M.; Najman, J.M.; Callaway, L.K. Associations of excess weight gain during pregnancy with long-term maternal overweight and obesity: Evidence from 21 y postpartum follow-up. *Am. J. Clin. Nutr.* **2010**, *91*, 1336–1341. [[CrossRef](#)] [[PubMed](#)]
19. Crane, J.M.; White, J.; Murphy, P.; Burrage, L.; Hutchens, D. The effect of gestational weight gain by body mass index on maternal and neonatal outcomes. *J. Obstet. Gynaecol. Can.* **2009**, *31*, 28–35. [[CrossRef](#)]
20. Mannan, M.; Doi, S.A.; Mamun, A.A. Association between weight gain during pregnancy and postpartum weight retention and obesity: A bias-adjusted meta-analysis. *Nutr. Rev.* **2013**, *71*, 343–352. [[CrossRef](#)] [[PubMed](#)]
21. Herring, S.J.; Rose, M.Z.; Skouteris, H.; Oken, E. Optimizing weight gain in pregnancy to prevent obesity in women and children. *Diabetes Obes. Metab.* **2012**, *14*, 195–203. [[CrossRef](#)]
22. Lucia Bergmann, R.; Bergmann, K.E.; Haschke-Becher, E.; Richter, R.; Dudenhausen, J.W.; Barclay, D.; Haschke, F. Does maternal docosahexaenoic acid supplementation during pregnancy and lactation lower BMI in late infancy? *J. Perinat. Med.* **2007**, *35*, 295–300. [[CrossRef](#)] [[PubMed](#)]
23. Poston, L. Gestational weight gain: Influences on the long-term health of the child. *Curr. Opin. Clin. Nutr. Metab. Care* **2012**, *15*, 252–257. [[CrossRef](#)]
24. Sridhar, S.B.; Darbinian, J.; Ehrlich, S.F.; Markman, M.A.; Gunderson, E.P.; Ferrara, A.; Hedderson, M.M. Maternal gestational weight gain and offspring risk for childhood overweight or obesity. *Am. J. Obstet. Gynecol.* **2014**, *211*, 259.e1–259.e8. [[CrossRef](#)]
25. Han, Z.; Lutsiv, O.; Mulla, S.; Rosen, A.; Beyene, J.; McDonald, S.D.; Group, K.S. Low gestational weight gain and the risk of preterm birth and low birthweight: A systematic review and meta-analyses. *Acta Obstet. Gynecol. Scand.* **2011**, *90*, 935–954. [[CrossRef](#)]
26. Blackburn, E.H.; Epel, E.S.; Lin, J. Human telomere biology: A contributory and interactive factor in aging, disease risks, and protection. *Science* **2015**, *350*, 1193–1198. [[CrossRef](#)]
27. Collins, K. The biogenesis and regulation of telomerase holoenzymes. *Nat. Rev. Mol. Cell Biol.* **2006**, *7*, 484–494. [[CrossRef](#)]
28. World Health Organization. *Obesity and Overweight 2012*; World Health Organization: Geneva, Switzerland, 2012.

29. Mundstock, E.; Sarria, E.E.; Zatti, H.; Mattos Louzada, F.; Kich Grun, L.; Herbert Jones, M.; Guma, F.T.; Mazzola In Memoriam, J.; Epifanio, M.; Stein, R.T.; et al. Effect of obesity on telomere length: Systematic review and meta-analysis. *Obesity (Silver Spring)* **2015**, *23*, 2165–2174. [[CrossRef](#)]
30. Njajou, O.T.; Cawthon, R.M.; Blackburn, E.H.; Harris, T.B.; Li, R.; Sanders, J.L.; Newman, A.B.; Nalls, M.; Cummings, S.R.; Hsueh, W.C. Shorter telomeres are associated with obesity and weight gain in the elderly. *Int. J. Obes. (Lond.)* **2012**, *36*, 1176–1179. [[CrossRef](#)] [[PubMed](#)]
31. Gorenjak, V.; Petrelis, A.M.; Stathopoulou, M.G.; Visvikis-Siest, S. Telomere length determinants in childhood. *Clin. Chem. Lab. Med.* **2020**, *58*, 162–177. [[CrossRef](#)]
32. Liu, B.; Song, L.; Zhang, L.; Wu, M.; Wang, L.; Cao, Z.; Xiong, C.; Zhang, B.; Li, Y.; Xia, W.; et al. Prenatal second-hand smoke exposure and newborn telomere length. *Pediatr. Res.* **2020**, *87*, 1081–1085. [[CrossRef](#)] [[PubMed](#)]
33. Salihu, H.M.; King, L.M.; Nwoga, C.; Paothong, A.; Pradhan, A.; Marty, P.J.; Daas, R.; Whiteman, V.E. Association Between Maternal-Perceived Psychological Stress and Fetal Telomere Length. *South. Med. J.* **2016**, *109*, 767–772. [[CrossRef](#)] [[PubMed](#)]
34. Martens, D.S.; Cox, B.; Janssen, B.G.; Clemente, D.B.P.; Gasparrini, A.; Vanpoucke, C.; Lefebvre, W.; Roels, H.A.; Plusquin, M.; Nawrot, T.S. Prenatal Air Pollution and Newborns' Predisposition to Accelerated Biological Aging. *JAMA Pediatr.* **2017**, *171*, 1160–1167. [[CrossRef](#)] [[PubMed](#)]
35. Isaevska, E.; Moccia, C.; Asta, F.; Cibella, F.; Gagliardi, L.; Ronfani, L.; Rusconi, F.; Stazi, M.A.; Richiardi, L. Exposure to ambient air pollution in the first 1000 days of life and alterations in the DNA methylome and telomere length in children: A systematic review. *Environ. Res.* **2021**, *193*, 110504. [[CrossRef](#)]
36. Martens, D.S.; Plusquin, M.; Gyselaers, W.; De Vivo, I.; Nawrot, T.S. Maternal pre-pregnancy body mass index and newborn telomere length. *BMC Med.* **2016**, *14*, 148. [[CrossRef](#)]
37. Vasu, V.; Turner, K.J.; George, S.; Greenall, J.; Slijepcevic, P.; Griffin, D.K. Preterm infants have significantly longer telomeres than their term born counterparts. *PLoS ONE* **2017**, *12*, e0180082. [[CrossRef](#)] [[PubMed](#)]
38. Lee, S.P.; Hande, P.; Yeo, G.S.; Tan, E.C. Correlation of cord blood telomere length with birth weight. *BMC Res. Notes* **2017**, *10*, 469. [[CrossRef](#)] [[PubMed](#)]
39. Sibert, N.T.; Ventura Ferreira, M.S.; Wagner, W.; Eipel, M.; Dreschers, S.; Brümmendorf, T.H.; Orlikowsky, T.; Beier, F. Cord blood telomere shortening associates with increased gestational age and birth weight in preterm neonates. *Exp. Ther. Med.* **2021**, *21*, 344. [[CrossRef](#)]
40. Fragkiadaki, P.; Tsoukalas, D.; Fragkiadoulaki, I.; Psycharakis, C.; Nikitovic, D.; Spandidos, D.A.; Tsatsakis, A.M. Telomerase activity in pregnancy complications (Review). *Mol. Med. Rep.* **2016**, *14*, 16–21. [[CrossRef](#)] [[PubMed](#)]
41. Bianchi, D.W. Circulating fetal DNA: Its origin and diagnostic potential—a review. *Placenta* **2004**, *25* (Suppl. A), S93–S101. [[CrossRef](#)]
42. Lo, Y.M. Recent advances in fetal nucleic acids in maternal plasma. *J. Histochem. Cytochem.* **2005**, *53*, 293–296. [[CrossRef](#)]
43. Taback, B.; Hoon, D.S. Circulating nucleic acids in plasma and serum: Past, present and future. *Curr. Opin. Mol. Ther.* **2004**, *6*, 273–278. [[PubMed](#)]
44. Deligezer, U.; Erten, N.; Akisik, E.E.; Dalay, N. Circulating fragmented nucleosomal DNA and caspase-3 mRNA in patients with lymphoma and myeloma. *Exp. Mol. Pathol.* **2006**, *80*, 72–76. [[CrossRef](#)] [[PubMed](#)]
45. Weikert, S.; Christoph, F.; Schrader, M.; Krause, H.; Miller, K.; Müller, M. Quantitative analysis of survivin mRNA expression in urine and tumor tissue of bladder cancer patients and its potential relevance for disease detection and prognosis. *Int. J. Cancer* **2005**, *116*, 100–104. [[CrossRef](#)]
46. Li, Y.; Zhou, X.; St John, M.A.; Wong, D.T. RNA profiling of cell-free saliva using microarray technology. *J. Dent. Res.* **2004**, *83*, 199–203. [[CrossRef](#)] [[PubMed](#)]
47. Larrabee, P.B.; Johnson, K.L.; Pestova, E.; Lucas, M.; Wilber, K.; LeShane, E.S.; Tantravahi, U.; Cowan, J.M.; Bianchi, D.W. Microarray analysis of cell-free fetal DNA in amniotic fluid: A prenatal molecular karyotype. *Am. J. Hum. Genet.* **2004**, *75*, 485–491. [[CrossRef](#)] [[PubMed](#)]
48. Soltani, M.; Nemati, M.; Maralani, M.; Estiar, M.A.; Andalib, S.; Fardiazar, Z.; Sakhinia, E. Cell-free fetal DNA in amniotic fluid supernatant for prenatal diagnosis. *Cell. Mol. Biol. (Noisy-le-grand)* **2016**, *62*, 14–17.
49. Underwood, M.A.; Gilbert, W.M.; Sherman, M.P. Amniotic fluid: Not just fetal urine anymore. *J. Perinatol.* **2005**, *25*, 341–348. [[CrossRef](#)]
50. Larrabee, P.B.; Johnson, K.L.; Lai, C.; Ordovas, J.; Cowan, J.M.; Tantravahi, U.; Bianchi, D.W. Global gene expression analysis of the living human fetus using cell-free messenger RNA in amniotic fluid. *JAMA* **2005**, *293*, 836–842. [[CrossRef](#)]
51. Zwemer, L.M.; Bianchi, D.W. The amniotic fluid transcriptome as a guide to understanding fetal disease. *Cold Spring Harb. Perspect. Med.* **2015**, *5*, a023101. [[CrossRef](#)]
52. Slonim, D.K.; Koide, K.; Johnson, K.L.; Tantravahi, U.; Cowan, J.M.; Jarrah, Z.; Bianchi, D.W. Functional genomic analysis of amniotic fluid cell-free mRNA suggests that oxidative stress is significant in Down syndrome fetuses. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 9425–9429. [[CrossRef](#)]
53. Koide, K.; Slonim, D.K.; Johnson, K.L.; Tantravahi, U.; Cowan, J.M.; Bianchi, D.W. Transcriptomic analysis of cell-free fetal RNA suggests a specific molecular phenotype in trisomy 18. *Hum. Genet.* **2011**, *129*, 295–305. [[CrossRef](#)]
54. Edlow, A.G.; Vora, N.L.; Hui, L.; Wick, H.C.; Cowan, J.M.; Bianchi, D.W. Maternal obesity affects fetal neurodevelopmental and metabolic gene expression: A pilot study. *PLoS ONE* **2014**, *9*, e88661. [[CrossRef](#)]

55. Massingham, L.J.; Johnson, K.L.; Scholl, T.M.; Slonim, D.K.; Wick, H.C.; Bianchi, D.W. Amniotic fluid RNA gene expression profiling provides insights into the phenotype of Turner syndrome. *Hum. Genet.* **2014**, *133*, 1075–1082. [[CrossRef](#)] [[PubMed](#)]
56. Kamath-Rayne, B.D.; Du, Y.; Hughes, M.; Wagner, E.A.; Muglia, L.J.; DeFranco, E.A.; Whitsett, J.A.; Salomonis, N.; Xu, Y. Systems biology evaluation of cell-free amniotic fluid transcriptome of term and preterm infants to detect fetal maturity. *BMC Med. Genom.* **2015**, *8*, 67. [[CrossRef](#)] [[PubMed](#)]
57. Cho, H.Y.; Cho, Y.; Shin, Y.J.; Park, J.; Shim, S.; Jung, Y.; Cha, D. Functional analysis of cell-free RNA using mid-trimester amniotic fluid supernatant in pregnancy with the fetal growth restriction. *Medicine (Baltimore)* **2018**, *97*, e9572. [[CrossRef](#)]
58. Jung, Y.W.; Shim, J.I.; Shim, S.H.; Shin, Y.J.; Chang, S.W.; Cha, D.H. Global gene expression analysis of cell-free RNA in amniotic fluid from women destined to develop preeclampsia. *Medicine (Baltimore)* **2019**, *98*, e13971. [[CrossRef](#)] [[PubMed](#)]
59. Tarca, A.L.; Romero, R.; Pique-Regi, R.; Pacora, P.; Done, B.; Kacerovsky, M.; Bhatti, G.; Jaiman, S.; Hassan, S.S.; Hsu, C.D.; et al. Amniotic fluid cell-free transcriptome: A glimpse into fetal development and placental cellular dynamics during normal pregnancy. *BMC Med. Genom.* **2020**, *13*, 25. [[CrossRef](#)] [[PubMed](#)]
60. Grisaru-Granovsky, S.; Reichman, B.; Lerner-Geva, L.; Boyko, V.; Hammerman, C.; Samueloff, A.; Schimmel, M.S.; Network, I.N. Population-based trends in mortality and neonatal morbidities among singleton, very preterm, very low birth weight infants over 16 years. *Early Hum. Dev.* **2014**, *90*, 821–827. [[CrossRef](#)] [[PubMed](#)]
61. Hug, L.; Alexander, M.; You, D.; Alkema, L.; Estimation, U.I.-a.G.f.C.M. National, regional, and global levels and trends in neonatal mortality between 1990 and 2017, with scenario-based projections to 2030: A systematic analysis. *Lancet Glob. Health* **2019**, *7*, e710–e720. [[CrossRef](#)]
62. Moutquin, J.M. Classification and heterogeneity of preterm birth. *BJOG* **2003**, *110* (Suppl. 20), 30–33. [[CrossRef](#)]
63. Goldenberg, R.L.; Culhane, J.F.; Iams, J.D.; Romero, R. Epidemiology and causes of preterm birth. *Lancet* **2008**, *371*, 75–84. [[CrossRef](#)]
64. Barchitta, M.; Maugeri, A.; Magnano San Lio, R.; Favara, G.; La Mastra, C.; La Rosa, M.C.; Agodi, A. Dietary Folate Intake and Folic Acid Supplements among Pregnant Women from Southern Italy: Evidence from the “Mamma & Bambino” Cohort. *Int. J. Environ. Res. Public Health* **2020**, *17*, 638. [[CrossRef](#)]
65. Maugeri, A.; Barchitta, M.; Agrifoglio, O.; Favara, G.; La Mastra, C.; La Rosa, M.C.; Magnano San Lio, R.; Panella, M.; Cianci, A.; Agodi, A. The impact of social determinants and lifestyles on dietary patterns during pregnancy: Evidence from the “Mamma & Bambino” study. *Ann. Ig.* **2019**, *31*, 81–89. [[PubMed](#)]
66. Maugeri, A.; Barchitta, M.; Favara, G.; La Rosa, M.C.; La Mastra, C.; Magnano San Lio, R.; Agodi, A. Maternal Dietary Patterns Are Associated with Pre-Pregnancy Body Mass Index and Gestational Weight Gain: Results from the “Mamma & Bambino” Cohort. *Nutrients* **2019**, *11*, 1308. [[CrossRef](#)]
67. Barchitta, M.; Maugeri, A.; La Rosa, M.C.; Magnano San Lio, R.; Favara, G.; Panella, M.; Cianci, A.; Agodi, A. Single Nucleotide Polymorphisms in Vitamin D Receptor Gene Affect Birth Weight and the Risk of Preterm Birth: Results From the “Mamma & Bambino” Cohort and A Meta-Analysis. *Nutrients* **2018**, *10*, 1172. [[CrossRef](#)]
68. Eveleth, P.B.; Andres, R.; Chumlea, W.C.; Eiben, O.; Ge, K.; Harris, T.; Heymsfield, S.B.; Launer, L.J.; Rosenberg, I.H.; Solomons, N.W.; et al. Uses and interpretation of anthropometry in the elderly for the assessment of physical status. Report to the Nutrition Unit of the World Health Organization: The Expert Subcommittee on the Use and Interpretation of Anthropometry in the Elderly. *J. Nutr. Health Aging* **1998**, *2*, 5–17.
69. Magnano San Lio, R.; Maugeri, A.; La Rosa, M.C.; Cianci, A.; Panella, M.; Giunta, G.; Agodi, A.; Barchitta, M. The Impact of Socio-Demographic Factors on Breastfeeding: Findings from the “Mamma & Bambino” Cohort. *Medicina (Kaunas)* **2021**, *57*, 103. [[CrossRef](#)]
70. Barchitta, M.; Maugeri, A.; Magnano San Lio, R.; La Rosa, M.C.; La Mastra, C.; Favara, G.; Giunta, G.; Cianci, A.; Agodi, A. Vaccination Status of Mothers and Children from the ‘Mamma & Bambino’ Cohort. *Vaccines* **2021**, *9*, 168.
71. Maugeri, A.; Barchitta, M.; Magnano San Lio, R.; La Rosa, M.C.; La Mastra, C.; Favara, G.; Ferlito, M.; Giunta, G.; Panella, M.; Cianci, A.; et al. The Effect of Alcohol on Telomere Length: A Systematic Review of Epidemiological Evidence and a Pilot Study during Pregnancy. *Int. J. Environ. Res. Public Health* **2021**, *18*, 5038. [[CrossRef](#)] [[PubMed](#)]
72. Barchitta, M.; Maugeri, A.; Quattrocchi, A.; Barone, G.; Mazzoleni, P.; Catalfo, A.; De Guidi, G.; Iemmolo, M.G.; Crimi, N.; Agodi, A. Mediterranean Diet and Particulate Matter Exposure Are Associated With LINE-1 Methylation: Results From a Cross-Sectional Study in Women. *Front. Genet.* **2018**, *9*, 514. [[CrossRef](#)]
73. Maugeri, A.; Barchitta, M.; Fiore, V.; Rosta, G.; Favara, G.; La Mastra, C.; La Rosa, M.C.; Magnano San Lio, R.; Agodi, A. Determinants of Adherence to the Mediterranean Diet: Findings from a Cross-Sectional Study in Women from Southern Italy. *Int. J. Environ. Res. Public Health* **2019**, *16*, 2963. [[CrossRef](#)]
74. Gielen, M.; Hageman, G.J.; Antoniou, E.E.; Nordfjall, K.; Mangino, M.; Balasubramanyam, M.; de Meyer, T.; Hendricks, A.E.; Giltay, E.J.; Hunt, S.C.; et al. Body mass index is negatively associated with telomere length: A collaborative cross-sectional meta-analysis of 87 observational studies. *Am. J. Clin. Nutr.* **2018**, *108*, 453–475. [[CrossRef](#)]
75. Vaiserman, A.; Krasnienkov, D. Telomere Length as a Marker of Biological Age: State-of-the-Art, Open Issues, and Future Perspectives. *Front. Genet.* **2020**, *11*, 630186. [[CrossRef](#)]
76. Rizvi, S.; Raza, S.T.; Mahdi, F. Telomere length variations in aging and age-related diseases. *Curr. Aging Sci.* **2014**, *7*, 161–167. [[CrossRef](#)]
77. Giller, A.; Andrawus, M.; Gutman, D.; Atzmon, G. Pregnancy as a model for aging. *Ageing Res. Rev.* **2020**, *62*, 101093. [[CrossRef](#)]



78. Gielen, M.; Hageman, G.; Pachen, D.; Derom, C.; Vlietinck, R.; Zeegers, M.P. Placental telomere length decreases with gestational age and is influenced by parity: A study of third trimester live-born twins. *Placenta* **2014**, *35*, 791–796. [[CrossRef](#)]
79. Clemente, D.B.P.; Maitre, L.; Bustamante, M.; Chatzi, L.; Roumeliotaki, T.; Fossati, S.; Grazuleviciene, R.; Gützkow, K.B.; Lepeule, J.; Martens, D.S.; et al. Obesity is associated with shorter telomeres in 8 year-old children. *Sci. Rep.* **2019**, *9*, 18739. [[CrossRef](#)]
80. Epel, E.S.; Lin, J.; Wilhelm, F.H.; Wolkowitz, O.M.; Cawthon, R.; Adler, N.E.; Dolbier, C.; Mendes, W.B.; Blackburn, E.H. Cell aging in relation to stress arousal and cardiovascular disease risk factors. *Psychoneuroendocrinology* **2006**, *31*, 277–287. [[CrossRef](#)]
81. Barker, D.J. The origins of the developmental origins theory. *J. Intern. Med.* **2007**, *261*, 412–417. [[CrossRef](#)]
82. Wang, C.; Nawrot, T.S.; Van Der Stukken, C.; Tylus, D.; Sleurs, H.; Peusens, M.; Alfano, R.; Langie, S.A.S.; Plusquin, M.; Martens, D.S. Different epigenetic signatures of newborn telomere length and telomere attrition rate in early life. *Aging (Albany N. Y.)* **2021**, *13*, 14630–14650. [[CrossRef](#)]
83. Skordalakes, E. Telomerase and the benefits of healthy living. *Lancet Oncol.* **2008**, *9*, 1023–1024. [[CrossRef](#)]
84. Kamath-Rayne, B.D.; Smith, H.C.; Muglia, L.J.; Morrow, A.L. Amniotic fluid: The use of high-dimensional biology to understand fetal well-being. *Reprod. Sci.* **2014**, *21*, 6–19. [[CrossRef](#)] [[PubMed](#)]
85. Bianchi, D.W.; LeShane, E.S.; Cowan, J.M. Large amounts of cell-free fetal DNA are present in amniotic fluid. *Clin. Chem.* **2001**, *47*, 1867–1869. [[CrossRef](#)]
86. Hui, L.; Bianchi, D.W. Cell-free fetal nucleic acids in amniotic fluid. *Hum. Reprod. Update* **2011**, *17*, 362–371. [[CrossRef](#)]
87. de Heredia, F.P.; Gómez-Martínez, S.; Marcos, A. Obesity, inflammation and the immune system. *Proc. Nutr. Soc.* **2012**, *71*, 332–338. [[CrossRef](#)] [[PubMed](#)]
88. Harris, H.E.; Ellison, G.T. Practical approaches for estimating prepregnant body weight. *J. Nurse Midwifery* **1998**, *43*, 97–101. [[CrossRef](#)]
89. Oken, E.; Taveras, E.M.; Kleinman, K.P.; Rich-Edwards, J.W.; Gillman, M.W. Gestational weight gain and child adiposity at age 3 years. *Am. J. Obstet. Gynecol.* **2007**, *196*, 322.e1–322.e8. [[CrossRef](#)]
90. Lun, F.M.; Chiu, R.W.; Leung, T.Y.; Leung, T.N.; Lau, T.K.; Lo, Y.M. Epigenetic analysis of RASSF1A gene in cell-free DNA in amniotic fluid. *Clin. Chem.* **2007**, *53*, 796–798. [[CrossRef](#)]
91. Aviv, A.; Hunt, S.C.; Lin, J.; Cao, X.; Kimura, M.; Blackburn, E. Impartial comparative analysis of measurement of leukocyte telomere length/DNA content by Southern blots and qPCR. *Nucleic Acids Res.* **2011**, *39*, e134. [[CrossRef](#)]
92. Soubry, A.; Murphy, S.K.; Wang, F.; Huang, Z.; Vidal, A.C.; Fuemmeler, B.F.; Kurtzberg, J.; Murtha, A.; Jirtle, R.L.; Schildkraut, J.M.; et al. Newborns of obese parents have altered DNA methylation patterns at imprinted genes. *Int. J. Obes. (Lond.)* **2015**, *39*, 650–657. [[CrossRef](#)]
93. Soubry, A.; Schildkraut, J.M.; Murtha, A.; Wang, F.; Huang, Z.; Bernal, A.; Kurtzberg, J.; Jirtle, R.L.; Murphy, S.K.; Hoyo, C. Paternal obesity is associated with IGF2 hypomethylation in newborns: Results from a Newborn Epigenetics Study (NEST) cohort. *BMC Med.* **2013**, *11*, 29. [[CrossRef](#)] [[PubMed](#)]