

Potential therapeutic applications of mesenchymal stem cells for the treatment of eye diseases

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Abstract

Stem cell-based treatments have been extensively explored in the last few decades to develop therapeutic strategies aimed at providing effective alternatives for those human pathologies in which surgical or pharmacological therapies produce limited effects. Among stem cells of different sources, mesenchymal stem cells (MSCs) offer several advantages, such as the absence of ethical concerns, easy harvesting, low immunogenicity and reduced tumorigenesis risks. Other than a multipotent differentiation ability, MSCs can release extracellular vesicles conveying proteins, mRNA and microRNA. Thanks to these properties, new therapeutic approaches have been designed for the treatment of various pathologies, including ocular diseases. In this review, the use of different MSCs and different administration strategies are described for the treatment of diabetic retinopathy, glaucoma, and retinitis pigmentosa. In a large number of investigations, positive results have been obtained by *in vitro* experiments and by MSC administration in animal models. Most authors agree that beneficial effects are likely related to MSC paracrine activity. Based on these considerations, many clinical trials have already been carried out. Overall, although some adverse effects have been described, promising outcomes are reported. It can be assumed that in the near future, safer and more effective protocols will be developed for more numerous clinical applications to improve the quality of life of patients affected by eye diseases.

Key Words: Mesenchymal stem cells; Eye diseases; Diabetic retinopathy; Glaucoma; Retinitis pigmentosa; Regenerative medicine

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Core Tip: Stem cell-based treatments have been extensively explored in the last few decades to develop therapeutic strategies. Mesenchymal stem cells (MSCs) offer several advantages, such as the absence of ethical concerns, easy harvesting, low immunogenicity and reduced tumorigenesis risks. MSC-based therapeutic approaches have been designed for the treatment of various ocular diseases, such as diabetic retinopathy, glaucoma and retinitis pigmentosa. In a large number of investigations, positive results have been obtained by *in vitro* experiments and in animal models. Many clinical trials have already been carried out and, although some adverse effects have been described, promising outcomes are reported.

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INTRODUCTION

Stem cell-based approaches have attracted considerable interest in the last few decades in view of their novel therapeutic applications. The regenerative potential of various types of stem cells has been widely explored, such as embryonic stem cells, induced pluripotent stem cells, and mesenchymal stem cells (MSCs). MSCs feature numerous advantages, such as the absence of ethical concerns, easy harvesting, low immunogenicity and tumorigenesis risks[1-3], and a marked ability to home to injury sites[4].

In the adults, MSCs are found in virtually all tissues where, by their self-renewal and multipotent differentiation capability, they ensure tissue growth, remodeling and repair[5]. When cultured *in vitro*, MSCs show a fibroblast-like morphology and, as indicated by the International Society for Cellular Therapy, they are positive for typical MSC markers[6] and are able to differentiate into adipocytes, osteoblasts, and chondrocytes. When administered *in vivo*, they induce beneficial effects, mostly thanks to the production of a variety of cytokines and growth factors (GF), such as basic fibroblast GF (bFGF) vascular endothelial GF (VEGF), transforming GF- β (TGF- β), hepatocyte GF, insulin-like GF-1 (IGF-1), ciliary neurotrophic factor (CNTF), brain-derived neurotrophic factor (BDNF), and glial cell-derived neurotrophic factor (GDNF)[2,7,8]. Moreover, they can release extracellular vesicles (EVs) conveying proteins, mRNA and microRNA (miRNA). In addition, like other cell types, MSCs feature immunomodulatory actions[9-11]. Because of these properties, new therapeutic approaches are being developed for the treatment of various pathologies, including ocular diseases[12]. Eye diseases may be due to different causes, such as photoreceptor loss, retinal ganglion cell (RGC) degeneration, and intraocular pressure (IOP) elevation. In this review, the use of different MSCs and different administration strategies are described for the treatment of diabetic retinopathy, glaucoma, and retinitis pigmentosa (RP). Most studies used MSCs derived from bone marrow (BMSCs) or adipose tissue (ASCs), but also those derived from the umbilical cord (UCMSCs) or conjunctiva (CJMSCs) were tested.

DIABETIC RETINOPATHY

Diabetic retinopathy (DR) is a vascular complication that occurs in diabetic patients. DR pathogenesis is associated with inflammation processes leading to microcirculation damage. As a result, a disruption of the blood-retina barrier (BRB) develops, mainly due to pericyte loss, which represents one of the earliest hallmarks of DR[13,14]. To date, only limited beneficial effects have been obtained with available therapeutic strategies. In fact, pericyte loss is virtually irreversible since in the adult retina they are not able to replicate[15,16]. Medical options trying to mitigate DR-induced damage include laser surgery, vitrectomy, and pharmacotherapy. However, these treatments are generally carried out when the disease is already in an advanced state. In addition, DR also affects retinal neuronal and glial cells. In fact, the visual loss occurring in the

early stages of DR is related to retinal sensory dysfunction, mainly due to RGC loss. For these reasons, many studies are focusing on developing therapies that can mitigate or arrest disease progression before irreversible damage occurs[17,18].

MSC-based treatments

Several studies *in vitro* or in animal models suggest that MSC-based strategies may represent a valuable tool for DR treatment. Main results are summarized in Table 1. A successful pericyte-like differentiation of ASCs was achieved by using specific culture media, as demonstrated by the expression of typical pericyte markers [α -smooth muscle actin, nerve/glial-antigen 2, platelet-derived growth factor receptor (PDGFR)] as well as by their perivascular localization in three-dimensional cultures[19,20]. In an *in vitro* model of the BRB, pericyte-like ASCs were also able to reinforce endothelial junctions, as indicated by the increased expression of VE-Cadherin, zone occludens-1 and occludin. In these experiments, also trans-endothelial electrical resistance values were increased[20]. It is also suggested that a high glucose (HG) concentration does not affect the pericyte-like functions of ASCs[21,22]. However, a conditioned medium (CM) from ASCs chronically cultured in HG conditions was able to normalize the HG-challenged retinal endothelial cells to normal glucose levels[23]. HG exposure of retinal endothelial cells activated NF- κ B and increased expression of downstream proinflammatory genes, such as interleukin (IL)-1B, VEGF and tumor necrosis factor (TNF). In bovine retinal endothelial cells, ASC-CM suppressed proinflammatory genes such as prostaglandin-endoperoxide synthase 2, and the proangiogenic gene VEGF. On the contrary, ASC-CM obtained using a shorter time of HG exposure did not induce beneficial effects. *In vitro* experiments indicated that, acting through Notch signaling, ASCs induce a stabilization of DR dysfunctional microvascular networks; in fact, genetic knockdown of Notch2 in ASCs disturbed the vascular network formation of human umbilical cord vein endothelial cells[24]. However, *in vivo* angiogenesis did not seem significantly influenced by Notch signaling, whose downregulation impairs ASC migratory ability, likely decreasing PDGFR expression.

Promising results were reported in experiments carried out in a murine model of DR[19]. A retinal capillary dropout was prevented and the revascularization of the central retina was achieved by an intravitreal injection of native or pretreated ASCs. Following intravitreal injection, it was also reported that BMSCs can repair retinal damage by their engrafting as photoreceptors[25]. In a mouse model of DR induced by streptozotocin, intravitreal administration of ASCs prevented RGC loss. In these studies, a retinal neuroprotection was obtained by reducing oxidative damage and increasing the intraocular levels of several neurotrophic factors[26]. In particular, mRNA and protein levels of nerve growth factor (NGF), bFGF, and GDNF were increased in treated eyes compared to controls. In addition, mRNA and protein levels of thrombospondin-1, a potent anti-angiogenic factor, also increased. In a similar rat model, intravitreal administration of ASCs was carried out two months after diabetes onset. Compared to controls, retinal histopathological evaluation revealed a significant decrease of vascular leakage and apoptotic cells around the retinal vessels. Moreover, a significantly improved “b” wave amplitude was observed in the electroretinogram. The authors suggest that ASCs may counteract retinal inflammation by blocking the up-regulation of several pro-inflammatory cytokines and DR-related genes[27].

Retinal stabilization against vascular insults was also reported in hyperglycemic Akimba mouse, by intravitreal ASC administration after retinal capillary dropout[28]. To compare their therapeutic efficacy, ASCs from either healthy or diabetic mice were injected. It was found that ASCs from healthy mice were more effective than diabetic ones for retinal protection. Moreover, engrafted ASCs were found associated with the retinal vasculature. ASC-mediated vascular stability is likely due to the secretion of FGF, IGF binding protein-3, monocyte chemoattractant protein-1, and stromal cell-derived factor (SDF-1). However, CM administration produced no significant effects, both when obtained from diabetic and healthy ASCs. However, small differences in the CM composition were noted: CM obtained from healthy mice was richer in some growth factors, such as IGF binding protein-3 and SDF-1. Dynamic effects were observed after intravitreal injection of both ASCs and their CM in mouse DR models.

Three weeks post-injection, mice treated with ASCs showed less vascular leakage and improved electroretinography tests, whereas visual acuity was not enhanced[17]. However, all visual deficits were ameliorated following CM administration. The better CM performance in this case was likely due to a cytokine ASC pretreatment before CM collection. Despite some discrepancies, there is a general agreement that many beneficial effects are exerted by MSC-derived EVs, which contain a considerable amount of bioactive molecules, such as miRNAs, cytokines, and immunomodulatory factors[29].

Table 1 Summary of the main results obtained in mesenchymal stem cell-based experiments for diabetic retinopathy

MSC source	Treatment	Results	Ref.
Human/mouse ASCs	<i>In vitro</i> pericyte-like differentiation of ASCs by addition of TGF- β 1	Increased expression of typical pericyte markers (α -smooth muscle actin and Nerve/glia-antigen 2)	Mendel <i>et al</i> [19], 2013
	Intravitreal injection of native or pericyte-like differentiated MSCs in a murine model of diabetic retinopathy	Prevention of retinal capillary dropout and improved revascularization of the central retina	
Human ASCs	<i>In vitro</i> pericyte-like differentiation of ASCs by using a culture medium designed for pericytes	Increased expression of typical pericyte markers (α -smooth muscle actin and Nerve/glia-antigen 2)	Mannino <i>et al</i> [20], 2020
	Co-cultures of pericyte-like ASCs and human retinal endothelial cells	Enhanced endothelial expression of junction proteins (VE-cadherin, zone occludens 1 and occludin). Increased trans endothelial electrical resistance in a blood-retina barrier model	
Human ASCs	High glucose cultures of retinal endothelial cells treated with conditioned medium from ASCs cultured in high glucose conditions	Suppression of pro-inflammatory and proangiogenic genes. Restoration of retinal endothelial cell parameters to normal glucose conditions	Hajmoussa <i>et al</i> [23], 2018
Human ASCs	ASC effects on endothelial cells of the human umbilical cord veins	Vascular network stabilization by ASC expression of NOTCH2. NOTCH2 downregulation impaired ASC migratory ability	Terlizzi <i>et al</i> [24], 2018
Mouse ASCs	Intravitreal ASC injection in streptozotocin-induced diabetic mouse model	Retinal ganglion cell loss was prevented. Reduced oxidative damage and increased intraocular levels of neurotrophic and anti-angiogenic factors	Ezquer <i>et al</i> [26], 2016
Human ASCs	Intravitreal ASC injection in streptozotocin-induced diabetic mouse model	Significant decrease of vascular leakage and apoptotic cells around the retinal vessels. Down-regulation of inflammatory gene expression in diabetic retina. Significantly improved "b" wave amplitude in the electroretinogram	Rajashekhar <i>et al</i> [27], 2014
Mouse ASCs	Intravitreal injections in Akimba mice of ASCs from diabetic or healthy mice, and respective conditioned media.	ASCs from healthy mice better improved retinal protection, likely because of higher levels of insulin-like growth factor binding protein-3 and stromal cell-derived factor-1	Cronk <i>et al</i> [28], 2015
Human ASCs	Intravitreal injection of cytokine-stimulated ASCs or their conditioned medium in a mouse model	ASC injection ameliorated electroretinogram parameters and reduced vascular leakage. Conditioned medium injections also improved visual acuity	Elshaer <i>et al</i> [17], 2018
Engineered Mouse BMSCs	High glucose treated Müller cells exposed to extracellular vesicles obtained from BMSCs overexpressing miRNA-486-3p	Improvement of Müller cell proliferation and inhibition of Toll-like receptor 4-mediated oxidative stress, inflammation and apoptosis	Li <i>et al</i> [30], 2020
Engineered Rat BMSCs	Intravitreal injections of BMSCs overexpressing C-X-C chemokine receptor type 4 in a streptozotocin-induced diabetic rat model	Reduced retinal damage and pro-inflammatory cytokine levels. Increased expression of rhodopsin and neuron specific enolase protein. Improved migratory ability	Wang <i>et al</i> [31], 2018
Engineered Rat BMSCs	Co-cultures of blue light injured retinal pigment epithelium cells with BMSCs overexpressing ciliary neurotrophic factor	Improved antioxidant ability and autophagic flux of retinal cells, whereas VEGF secretion and apoptosis were decreased	Lin <i>et al</i> [32], 2018
Engineered Human placenta MSCs	Co-cultures of oxidative stress-injured retinal pigment epithelium cells with BMSCs overexpressing pigment epithelium-derived factor	Improved mitochondrial functions and reduced apoptosis of retinal cells	Kim <i>et al</i> [33], 2021
	Intravitreal injection of engineered BMSCs in a rat model of retinal degeneration	Decreased levels of pro-inflammatory cytokines and vascular endothelial growth factor. Restoration of retina structure and function	
Autologous BMSCs	Pilot clinical trial carried out by intravitreal injection of autologous BMSCs in patients with diabetic retinopathy	Fasting blood glucose and hypersensitive C-reactive protein were reduced. Eyes of patients with non-proliferative diabetic retinopathy showed a macular thickness reduction and a significant improvement of best corrected visual acuity	Gu <i>et al</i> [34], 2018

ASCs: Adipose tissue stem cells; MSCs: Mesenchymal stem cells; TGF: Transforming growth factor; BMSCs: Bone marrow stem cells.

Genetically engineered MSCs

In an *in vitro* study, beneficial effects were observed in HG-treated Müller cells when exposed to EVs released by genetically engineered BMSCs. In particular, compared to control conditions, EVs from BMSCs overexpressing miRNA-486-3p were able to inhibit Toll-like receptor 4-mediated oxidative stress, inflammation and apoptosis, as well as promoting Müller cell proliferation [30]. BMSC ability to home to the injury site is guided by SDF-1 through its C-X-C chemokine receptor type 4 (CXCR4) [31]. To

improve this ability, MSCs were infected by a lentivirus constructed with CXCR4. *In vitro* observations revealed that transduced MSCs had high expression levels of the CXCR4 gene and protein, showing improved migration activities. When intravitreally injected, they reduced retinal injury in a DR animal model. In particular they overexpressed rhodopsin and neuron specific enolase, while decreasing the inflammatory cytokines IL-6 and TNF- α . The authors conclude that this strategy may be useful in DR treatment. The neuroprotective ability of CNTF-overexpressing BMSCs was tested in co-cultures with blue light-injured retinal pigment epithelium (RPE) cells[32]. In fact, it was demonstrated that CNTF protected RPE cells and modulated the secretion of neurotrophic factors and cytokines. Results showed an improved antioxidant ability and autophagic flux of RPE cells, whereas VEGF secretion and apoptosis were decreased. In another investigation, placenta-derived MSCs were genetically modified to overexpress pigment epithelium-derived factor (PEDF). They were then able to maintain stemness and showed an increased mRNA expression of biogenesis regulators and antioxidant enzymes[33]. When co-cultured with RPE cells injured by oxidative stress, engineered MSCs improved mitochondrial functions and reduced apoptosis by activating the cell survival JNK/ERK signaling pathway. Beneficial effects were also observed after their intravitreal administration in a rat model with H₂O₂-induced retinal degeneration. In fact, proinflammatory cytokines and VEGF expression decreased, whereas an increased PEDF expression promoted a restoration of retina structure and function.

Clinical trials

In a pilot clinical trial, the safety and efficacy of intravenous administration of autologous BMSCs were evaluated for the treatment of patients with proliferative or non-proliferative DR. Results show that fasting blood glucose and hypersensitive C-reactive protein were reduced in both groups of patients, whereas no significant differences were observed concerning hemoglobin A_{1c} or IL-6 levels. Only eyes of patients with non-proliferative DR showed a macular thickness reduction and a significant improvement of best corrected visual acuity (BCVA)[34].

GLAUCOMA

More than 70 million people suffer irreversible blindness due to glaucoma. Whereas primary glaucoma is of unknown origin, secondary glaucoma is associated with identifiable causes leading to increased IOP, optic nerve injury, and vision loss[29]. Moreover, two major types of glaucoma are genetically distinct: closed-angle and open-angle glaucoma. Overall, glaucoma is an age-related multifactorial and slow progressive eye disease, in which IOP elevation may result from an increased production and/or decreased outflow of aqueous humor, generally consequent to the degeneration of the trabecular meshwork (TM), the ocular tissue regulating aqueous humor outflow[35]. The main consequences comprise degeneration of RGCs and their axons, optic nerve shrinkage, and changes in brain visual areas[36]. The slow progression of the disease requires long-term ophthalmic medications, and 15%-25% of patients progressively lose sight, even if IOP is kept under control. Other mechanisms are likely involved in disease progression[29,37-39]. *Lamina cribrosa* changes may reduce RGC axonal transport and favor apoptotic mechanisms[40]. In fact, RGC axons normally support the retina with various neurotrophic factors, such as NGF, BDNF, CNTF, and GDNF. Since RGC regeneration is problematic[29], their loss or dysfunction severely impairs vision and quality of life. By targeting TM, the main therapeutic approaches for glaucoma are currently aimed at reducing IOP, and avoiding further RGC loss, but these interventions only delay disease progression. Indeed, some studies report the presence of MSC-like cells within the TM and between the TM and the corneal endothelial periphery, but they are not able to efficiently restore cell damage[41].

MSC-based treatments

Results obtained following MSC-based treatments are summarized in Table 2. Most of them indicate an improved RGC survival, a production of functional RGC-like cells, and an expansion and differentiation of resident retinal stem cells in mature RGCs[42, 43]. Although intravitreal administration often produces an exiguous number of MSC engraftments and retinal differentiation, beneficial effects are also attributable to MSC release of neurotrophic factors[36]. Co-culturing BMSCs with pigmented cells from the ciliary margin, the photoreceptor specific marker, cone-rod homeobox (Crx), was

Table 2 Summary of the main results obtained in mesenchymal stem cell-based experiments for glaucoma

MSC source	Treatment	Results	Ref.
Rat BMSCs	<i>In vitro</i> co-cultures of BMSCs with pigmented cells from ciliary margin	Enhanced expression of cone-rod homeobox, at both mRNA and protein level	Li <i>et al</i> [43], 2017
Mouse BMSCs	BMSC injection into the ocular anterior chamber of a laser-induced rat model	Decreased intraocular pressure likely due to MSC paracrine activity	Manuguerra-Gagné <i>et al</i> [44], 2013
Rat BMSCs	BMSC injection into the ocular anterior chamber of a rat model obtained by cauterization of episcleral veins	Reduced intraocular pressure, associated to MSC engrafting to the ciliary processes and to the trabecular meshwork	Roubeix <i>et al</i> [45], 2015
	BMSC-conditioned medium treatment of TGF- β 2-induced fibrotic human trabecular meshwork cell cultures	Improved survival of trabecular meshwork cells, by activation of anti-apoptotic signaling pathway Akt	
Rat BMSCs	Intravitreal or intravenous administration of labeled BMSCs in a laser-induced ocular hypertension rat model	Local but not systemic BMSC administration improved retinal ganglion cell survival. Transplanted BMSCs were preferentially located in a perivascular position	Johnson <i>et al</i> [47], 2010
Rat/humanBMSCs	Co-cultures of retinal explants with rat and human BMSCs, or rat and human fibroblasts	BMSCs but not fibroblasts improved retinal ganglion cell survival. An increased density of cells expressing islet-1 and NeuN was found in retinal ganglion cell layer	Johnson <i>et al</i> [48], 2014
	Exposure of retinal explant cultures to conditioned media from BMSCs or fibroblast	Moderate improvements were observed only for BMSC conditioned medium treatment	
Human BMSCs	Intravitreal injections of extracellular vesicles obtained from BMSCs of fibroblasts in two rat models of glaucoma	BMSC extracellular vesicles more efficiently reduced retinal ganglion cell apoptosis	Mead <i>et al</i> [38], 2018
Engineered mouse BMSCs	Indirect co-cultures of BMSCs with retinal ganglion cells in a transwell system	BMSCs enhanced anti-apoptotic effects on retinal ganglion cells by upregulation of stanniocalcin-1. Improved effects were obtained by miRNA-21-overexpressing BMSCs	Su <i>et al</i> [49], 2017
Autologous BMSCs	Clinical trial carried out by intravitreal injection of autologous BMSCs in patients with advanced stage of glaucoma	No significant improvements were reported	Vilela <i>et al</i> [50], 2021

MSCs: Mesenchymal stem cells; TGF: Transforming growth factor; BMSCs: Bone marrow stem cells; miRNA: MicroRNA.

found increased at both mRNA and protein expression levels[43]. In addition, MSC retinal differentiation was supported by increased percentages of cells that were immunopositive for typical retinal markers such as rhodopsin, visual system homeobox 2 and heparin sulphate. In fact, these markers are specific for rods, bipolar neurons and Müller glia, respectively. Many studies have been aimed at promoting TM regeneration; for example, Manuguerra-Gagné *et al*[44] reported that restored IOP levels were observed in a rat model after BMSC administration into the anterior chamber, this was likely due to paracrine mechanisms. Neuroprotective effects *via* TM protection were also reported in another study by BMSC injection in a rat model of glaucoma-like ocular hypertension[45]. Rapid and long-lasting BMSC-induced effects were associated with MSC engrafting to the ciliary processes and the TM. The same authors also showed that increased TM cell *in vitro* survival was obtained using MSC-CM, likely activating the anti-apoptotic pathway Akt. In fact, MSC anti-apoptotic effects had been previously reported[46]. At the same time, TM cell relaxation was induced by decreasing myosin phosphorylation and the TGF- β 2-dependent profibrotic phenotype was inhibited. In a laser-induced rat model of glaucoma, it was reported that only local BMSC administration, but not systemic administration, induced neuroprotective effects by improving RGC axon survival[47]. Most BMSCs were preferentially located to a perivascular position, and some of them migrated within the nerve fibers and the retinal ganglion cells. In co-cultures of retinal explants with rat or human BMSCs, a reduction of apoptosis and an RGC survival increase was observed. In fact, compared to cultures of the retina alone, a high density of cells expressing Islet-1 and NeuN was observed in the RGC layer. On the contrary, the addition of fibroblasts in analogous conditions was not able to reproduce these effects. Although inducing lesser neuroprotective effects, also the use of BMSC-CM has been proven effective, compared with fibroblast-CM. The differential analysis between the two conditioned media revealed that a significant difference was related to the presence of PDGF, which was contained only in BMSC-CM. It was then concluded that PDGF is responsible for these neuroprotective effects[48]. According to Mead *et al*[38], most MSC positive effects are due to their paracrine release of neurotrophic factors and EVs.

Intravitreal BMSC- or fibroblast-EV administration was carried out on two glaucoma rat models obtained by intracameral injection of microbeads or laser photocoagulation of the TM and limbal vessels. Data reported that decreased RGC apoptosis was obtained following BMSC-EV treatment, rather than fibroblast-EV injection. BMSC-miRNA would play an important role in RGC regeneration mechanisms by modulating the expression of the phosphatase and tensin homolog, a suppressor of RGC axonal growth and survival. Overall, more evident improvements were found in aged rats, characterized by a faster RGC loss when compared with young animals[37].

Genetically engineered MSCs

In a work by Su *et al*[49], BMSCs were co-cultured with RGCs in a transwell system mimicking *in vivo* conditions. Whereas RGCs cultured alone showed significantly decreased cell viability if subjected to oxygen-glucose deprivation and reperfusion, co-cultures with MSCs significantly increased RGC viability in a dose-dependent manner. Improved RGC survival was likely due to MSC-induced production of stanniocalcin 1, thus inhibiting caspase-8-mediated apoptosis. MSC transfection with miRNA-21, essential for stanniocalcin 1 production, further ameliorated RGC survival.

Clinical trials

No significant effects have been described to date in clinical trials. In patients with advanced glaucoma, intravitreal injection of autologous BMSCs did not induce appreciable improvements on visual acuity or the visual field[50].

RETINITIS PIGMENTOSA

Retinitis pigmentosa (RP) is a genetic disease affecting 1.5 million people worldwide. It leads to night blindness, tunnel vision, progressive central vision loss and total blindness. Modified molecular processes involved in the turnover of rhodopsin progressively induce the total loss of rod cells[51-53]. Subsequently, an increase of reactive oxygen species (ROS) levels occurring in RPE induces cone loss at later stages [54-57]. In fact, ROS oxidize fatty acids triggering inflammation, synaptic injury, apoptosis, and cell death[58]. Since there are no satisfactory therapeutic approaches for this disease, new strategies are being developing to replace or repair damaged or dead cells by using stem cell therapy[59,60].

MSC-based treatments

MSC intraocular administration would improve visual function[3,61-63] through different mechanisms: cell replacement following MSC trans-differentiation, paracrine effects, or inflammation modulation. In addition, also electroretinographic parameters could be improved[61,62,64]. Main results are summarized in Table 3.

In vitro studies have reported the ability of MSCs to differentiate into retinal progenitor cells, photoreceptors, and retinal neuron-like cells[65,66]. It has been shown that either in the presence of retinal cells, conditioned media from retinal cell cultures, or retinal cell extracts, MSCs differentiate into retinal-like cells, expressing typical genes and markers[67]. By using a taurine containing induction medium, CJMSCs were differentiated into photoreceptor-like cells in fibrin gel, as indicated by the expression of typical markers such as Nestin, rhodopsin, and RPE65. The authors claim that this 3D scaffold, fibrin gel-based strategy may lead to increased availability of photoreceptor progenitor cells for retina repair and regeneration[68].

When BMSCs were intravitreally injected in a rat model of retina degeneration, they expressed rhodopsin and pancytokeratin (PCK) markers. Rhodopsin indicates a photoreceptor-like differentiation, while PCK expression is indicative of an RPE-like phenotype[65]. Injected in the subretinal space of rhodopsin knockout mouse, engrafted BMSCs were found not only in the RPE, but also in the neuroretina, acquiring neuronal and glial morphologies, and prolonging photoreceptor survival [69]. In a rat model of RP, intravenous BMSC administration before photoreceptor loss was able to preserve both rod and cone cells. By multiunit activity recording from the superior colliculus, it was concluded that also visual functions, such as BCVA and luminance threshold, were significantly preserved[70].

Intravitreal injection of mouse ASCs or human UCMSCs, and their derived EVs, promoted repair and protective actions in rodent models of laser-induced retinal damage[71]. It was found that both MSCs and their exosomes induced comparable beneficial effects. They were able to reduce retinal damage, inhibit apoptosis, and suppress inflammatory responses, thus obtaining an improved visual function.

Table 3 Summary of the main results obtained in mesenchymal stem cell-based experiments for retinitis pigmentosa

MSC source	Treatment	Results	Ref.
ConjunctivaMSCs	Retinal-like MSC differentiation in 3D scaffold fibrin gel, by an induction medium containing taurine	Increased expression of typical retinal markers (nestin, rhodopsin, and RPE65)	Soleimannejad <i>et al</i> [68], 2017
Rat BMSCs	Intravitreal BMSC injections in a rat model of retina degeneration	Increased expression of typical photoreceptor marker (rhodopsin) and retinal pigment epithelium (pancytokeratin)	Huo <i>et al</i> [65], 2010
Mouse BMSCs	BMSC injections in the subretinal space of rhodopsin knockout mice	MSC engrafting in retinal pigment epithelium and neuroretina, acquiring neuronal and glial morphology, and prolonging photoreceptor survival	Arnhold <i>et al</i> [69], 2007
Rat BMSCs	Intravenous BMSC administration in a rat model of retinitis pigmentosa, before photoreceptor loss	Rod and cone cell preservation. Improved visual functions	Wang <i>et al</i> [70], 2010
Mouse ASCs/human UCMSCs	Intravitreal MSC or their exosomes in a laser-induced mouse model of retinal injury	Both MSCs and their exosomes reduced retinal damage, apoptosis and inflammatory responses	Yu <i>et al</i> [71], 2016
Human BMSCs	Physically separated co-cultures of BMSCs with porcine neuroretina	BMSC secretomes induced increased levels of anti-inflammatory cytokines, antioxidant and anti-apoptotic proteins, and anti-neurodegeneration proteins, Photoreceptors and the retinal general organization were better preserved	Usategui-Martin <i>et al</i> [72], 2020
Engineered human UCMSCs	Photoreceptor-like differentiation of UCMSCs overexpressing erythropoietin, by an induction medium containing taurine	Transduced UCMSCs better overexpressed cone-rod homeobox and rhodopsin	Ding <i>et al</i> [73], 2019
Engineered human BMSCs	Photoreceptor-like differentiation of BMSCs overexpressing miRNA-183/96/182 cluster	Transduced BMSCs overexpressed recoverin, a neuronal calcium-binding protein primarily detected in photoreceptors.	Mahmoudian-Sani <i>et al</i> [74], 2019
Engineered conjunctiva MSCs	Photoreceptor-like differentiation of MSCs overexpressing miRNA-9	In a time dependent manner, MSCs overexpressed photoreceptor markers recoverin, rhodopsin, PKC, and peripherin	Rahmani <i>et al</i> [75], 2020
Engineered mouse BMSCs	Intravitreal injections of BMSCs genetically modified to overexpress neurotrophin-4, in a mouse model of retinopathy	Neurotrophin-4-overexpressing BMSCs induced a significant retinal protection, as indicated by bioelectrical recordings and optical coherence tomography.	Machalińska <i>et al</i> [76], 2013
Autologous MSCs	In a phase I clinical trial, the safety, feasibility, and efficacy of autologous BMSC was tested after their intravitreal injection in patients with advanced retinitis pigmentosa	In spite of several mild and transient adverse effects, a significant improvement of best corrected visual acuity was observed. Only one severe, though manageable adverse effect was observed after three years	Tuekprakhon <i>et al</i> [77], 2021
Human UCMSCs	In a phase 3 clinical study, UCMSC administration was carried out by surgical procedures in the suprachoroidal area of 82 patients with retinitis pigmentosa	Over 6-month follow-up period, no serious adverse events were reported. Significant improvements were detected for best corrected visual acuity, visual field and multifocal electroretinography recordings. Beneficial effects might be due to MSC secretion of growth factors.	Kahraman <i>et al</i> [78], 2020
Human UCMSCs	In a phase I/II clinical trial, the protocol safety and efficacy was tested by intravenous administration of UCMSCs in 32 patients with advanced retinitis pigmentosa	The intravenous infusion of UCMSCs was safe, since no serious local or systemic adverse effects occurred over a period of 12 mo. Most patients improved their best corrected visual acuity, especially in the first 3 months. It was concluded that this strategy may improve the quality of life of patients with retinitis pigmentosa.	Zhao <i>et al</i> [79], 2020

ASCs: Adipose tissue stem cells; MSCs: Mesenchymal stem cells; TGF: Transforming growth factor; BMSCs: Bone marrow stem cells; UCMSCs: Umbilical cord mesenchymal stem cells.

In physically separated co-cultures of BMSCs with neuroretinas, it was found that neuroprotective effects were induced by BMSC secretomes. In particular, anti-inflammatory cytokines, antioxidant and anti-apoptotic proteins, as well as anti-neurodegeneration proteins, were increased in these co-cultures. Probably by secreting these neuroprotective molecules, photoreceptors and the retinal general organization were better preserved[72].

Genetically engineered MSCs

In an *in vitro* study, UCMSCs were tested for photoreceptor differentiation by using a specific culture medium containing taurine[73]. Indeed, compared to control cultures, taurine-treated cells showed an increased expression of Crx and rhodopsin. However, an enhanced expression of these photoreceptor markers was obtained when UCMSCs

were previously transduced with lentivirus particles encoding erythropoietin (EPO). In fact, EPO was proven to play a role in neuro-regeneration and protection, cell differentiation and rescue potential on dying photoreceptor cells. Since the fluorescence intensity was higher in rhodopsin-positive cells, it was suggested that a rod photoreceptor differentiation was favored.

In an *in vitro* work by Mahmoudian-Sani *et al*[74], human BMSCs were induced to differentiate into photoreceptor cells *via* transfection with the miR-183/96/182 cluster. Results showed that recoverin, a neuronal calcium-binding protein primarily detected in photoreceptors, was upregulated. Therefore, the authors concluded that overexpression of miR-183 can induce a successful reprogramming of BMSCs into photoreceptor cells.

A new approach was introduced by Rahmani *et al*[75] to guide the differentiation of CJMSCs into photoreceptor-like cells by miRNA-9 transduction. By using neither chemical nor biological growth factors, transduced cells were able to express, in a time-dependent manner, photoreceptor markers such as recoverin, rhodopsin, PKC, and peripherin. In particular, rhodopsin was the most enhanced marker after miRNA-9 overexpression.

BMSCs genetically modified to overexpress neurotrophin-4 (NT-4) were intravitreally injected in a mouse model of retinopathy[76]. NT-4-overexpressing BMSCs migrated into the injured retina, actively producing NT-4 and inducing a significant retinal protection. In particular, the preservation of retinal bioelectrical activity was indicated by electroretinography, and a complete restoration of the retina organization was shown by optical coherence tomography. Overall, the authors claim that this treatment induced a positive impact on neural cell survival, growth, and differentiation.

Clinical trials

In a phase I clinical trial, Tuekprakhon *et al*[77] investigated the safety, feasibility, and efficacy of autologous BMSC intravitreal injection in patients with advanced RP. In three groups of patients, BCVA, visual field (VF), central subfield thickness (CST), and subjective experiences were evaluated over 12 mo. In spite of several mild and transient adverse effects, a statistically significant improvement of BCVA was observed, whereas VF and CST remained stable, indicating no further progression of the disease. Only one severe, though manageable, adverse effect was observed after three years.

In a phase 3 clinical study, UCMSC administration was carried out in the suprachoroidal area of 82 RP patients by a surgical procedure[78]. Over a 6-month follow-up period, no serious adverse events or ocular complications were reported. Overall, statistically significant improvements in BCVA and VF were observed, as well as improvements in multifocal electroretinography recordings. The authors conclude that beneficial effects may possibly be due to MSC secretion of growth factors.

In a phase I/II clinical trial, intravenous administration of UCMSCs was performed in 32 advanced RP patients to evaluate the safety and efficacy protocol[79]. After a single dose injection, all subjects were followed-up over a period of 12 mo. Results showed that the intravenous infusion of UCMSCs was safe, since no serious local or systemic adverse effects occurred. Most patients improved their BCVA, especially in the first 3 mo. It was concluded that the intravenous infusion of UCMSCs may improve the quality of life of advanced RP patients.

CONCLUSION

In the last few decades, stem cell-based strategies have been extensively explored to develop therapeutic protocols aimed at counteracting human pathologies when other remedies fail. Although a variety of stem cells feature formidable potential therapeutic applications, in many cases they cannot be currently used because of ethical issues and/or safety problems. MSCs probably represent an acceptable compromise, being easily available even for autologous administration. Many experimental data obtained both in *in vitro* studies and in animal models report encouraging results, although after transplantation, only a limited fraction of injected cells really engrafted and differentiated into functional cells. Because of the wide MSC paracrine activity, also cell-free treatments have been explored with positive effects. Genetic manipulations of MSCs have also been performed to improve their ability to home to damaged sites and to overexpress cytokine/growth factors helping retinal restoration. Many clinical trials have already been carried out and, despite some adverse effects, promising outcomes

are reported. It is not difficult to assume that in the near future, safer and more effective protocols will be designed for clinical applications, to improve the quality of life of patients affected by eye disease.

REFERENCES

- 1 **Romanov YA**, Darevskaya AN, Merzlikina NV, Buravkova LB. Mesenchymal stem cells from human bone marrow and adipose tissue: isolation, characterization, and differentiation potentialities. *Bull Exp Biol Med* 2005; **140**: 138-143 [PMID: 16254640 DOI: 10.1007/s10517-005-0430-z]
- 2 **Lindroos B**, Suuronen R, Miettinen S. The potential of adipose stem cells in regenerative medicine. *Stem Cell Rev Rep* 2011; **7**: 269-291 [PMID: 20853072 DOI: 10.1007/s12015-010-9193-7]
- 3 **Öner A**. Stem Cell Treatment in Retinal Diseases: Recent Developments. *Turk J Ophthalmol* 2018; **48**: 33-38 [PMID: 29576896 DOI: 10.4274/tjo.89972]
- 4 **Rüster B**, Göttig S, Ludwig RJ, Bistrrian R, Müller S, Seifried E, Gille J, Henschler R. Mesenchymal stem cells display coordinated rolling and adhesion behavior on endothelial cells. *Blood* 2006; **108**: 3938-3944 [PMID: 16896152 DOI: 10.1182/blood-2006-05-025098]
- 5 **Baddour JA**, Sousounis K, Tsonis PA. Organ repair and regeneration: an overview. *Birth Defects Res C Embryo Today* 2012; **96**: 1-29 [PMID: 22457174 DOI: 10.1002/bdrc.21006]
- 6 **Lo Furno D**, Mannino G, Giuffrida R, Gili E, Vancheri C, Tarico MS, Perrotta RE, Pellitteri R. Neural differentiation of human adipose-derived mesenchymal stem cells induced by glial cell conditioned media. *J Cell Physiol* 2018; **233**: 7091-7100 [PMID: 29737535 DOI: 10.1002/jcp.26632]
- 7 **Luo S**, Hao L, Li X, Yu D, Diao Z, Ren L, Xu H. Adipose tissue-derived stem cells treated with estradiol enhance survival of autologous fat transplants. *Tohoku J Exp Med* 2013; **231**: 101-110 [PMID: 24107653 DOI: 10.1620/tjem.231.101]
- 8 **Hu ZL**, Li N, Wei X, Tang L, Wang TH, Chen XM. Neuroprotective effects of BDNF and GDNF in intravitreally transplanted mesenchymal stem cells after optic nerve crush in mice. *Int J Ophthalmol* 2017; **10**: 35-42 [PMID: 28149774 DOI: 10.18240/ijo.2017.01.06]
- 9 **Vancheri C**, Mastruzzo C, Trovato-Salinaro E, Gili E, Lo Furno D, Pistorio MP, Caruso M, La Rosa C, Crimi C, Failla M, Crimi N. Interaction between human lung fibroblasts and T-lymphocytes prevents activation of CD4+ cells. *Respir Res* 2005; **6**: 103 [PMID: 16159396 DOI: 10.1186/1465-9921-6-103]
- 10 **Gao F**, Chiu SM, Motan DA, Zhang Z, Chen L, Ji HL, Tse HF, Fu QL, Lian Q. Mesenchymal stem cells and immunomodulation: current status and future prospects. *Cell Death Dis* 2016; **7**: e2062 [PMID: 26794657 DOI: 10.1038/cddis.2015.327]
- 11 **Lo Furno D**, Graziano AC, Avola R, Giuffrida R, Perciavalle V, Bonina F, Mannino G, Cardile V. A Citrus bergamia Extract Decreases Adipogenesis and Increases Lipolysis by Modulating PPAR Levels in Mesenchymal Stem Cells from Human Adipose Tissue. *PPAR Res* 2016; **2016**: 4563815 [PMID: 27403151 DOI: 10.1155/2016/4563815]
- 12 **Harrell CR**, Fellabaum C, Arsenijevic A, Markovic BS, Djonov V, Volarevic V. Therapeutic Potential of Mesenchymal Stem Cells and Their Secretome in the Treatment of Glaucoma. *Stem Cells Int* 2019; **2019**: 7869130 [PMID: 31949441 DOI: 10.1155/2019/7869130]
- 13 **Kim JM**, Hong KS, Song WK, Bae D, Hwang IK, Kim JS, Chung HM. Perivascular Progenitor Cells Derived From Human Embryonic Stem Cells Exhibit Functional Characteristics of Pericytes and Improve the Retinal Vasculature in a Rodent Model of Diabetic Retinopathy. *Stem Cells Transl Med* 2016; **5**: 1268-1276 [PMID: 27388242 DOI: 10.5966/sctm.2015-0342]
- 14 **Santos GSP**, Prazeres PHDM, Mintz A, Birbrair A. Role of pericytes in the retina. *Eye (Lond)* 2018; **32**: 483-486 [PMID: 29125148 DOI: 10.1038/eye.2017.220]
- 15 **Alikhani M**, Roy S, Graves DT. FOXO1 plays an essential role in apoptosis of retinal pericytes. *Mol Vis* 2010; **16**: 408-415 [PMID: 20300563]
- 16 **Beltramo E**, Porta M. Pericyte loss in diabetic retinopathy: mechanisms and consequences. *Curr Med Chem* 2013; **20**: 3218-3225 [PMID: 23745544 DOI: 10.2174/09298673113209990022]
- 17 **Elshaer SL**, Evans W, Pentecost M, Lenin R, Periasamy R, Jha KA, Alli S, Gentry J, Thomas SM, Sohl N, Gangaraju R. Adipose stem cells and their paracrine factors are therapeutic for early retinal complications of diabetes in the Ins2^{Akita} mouse. *Stem Cell Res Ther* 2018; **9**: 322 [PMID: 30463601 DOI: 10.1186/s13287-018-1059-y]
- 18 **Valle MS**, Russo C, Malaguarnera L. Protective role of vitamin D against oxidative stress in diabetic retinopathy. *Diabetes Metab Res Rev* 2021 [PMID: 33760363 DOI: 10.1002/dmrr.3447]
- 19 **Mendel TA**, Clabough EB, Kao DS, Demidova-Rice TN, Durham JT, Zotter BC, Seaman SA, Cronk SM, Rakoczy EP, Katz AJ, Herman IM, Peirce SM, Yates PA. Pericytes derived from adipose-derived stem cells protect against retinal vasculopathy. *PLoS One* 2013; **8**: e65691 [PMID: 23741506 DOI: 10.1371/journal.pone.0065691]
- 20 **Mannino G**, Gennuso F, Giurdanella G, Conti F, Drago F, Salomone S, Furno DL, Bucolo C, Giuffrida R. Pericyte-like differentiation of human adipose-derived mesenchymal stem cells: An *in vitro* study. *World J Stem Cells* 2020; **12**: 1152-1170 [PMID: 33178398 DOI: 10.4252/wjsc.v12.i10.1152]
- 21 **Hajmoussa G**, Elorza AA, Nies VJ, Jensen EL, Nagy RA, Harmsen MC. Hyperglycemia Induces Bioenergetic Changes in Adipose-Derived Stromal Cells While Their Pericytic Function Is Retained.

- Stem Cells Dev* 2016; **25**: 1444-1453 [PMID: 27473785 DOI: 10.1089/scd.2016.0025]
- 22 **Fiori A**, Hammes HP, Bieback K. Adipose-derived mesenchymal stromal cells reverse high glucose-induced reduction of angiogenesis in human retinal microvascular endothelial cells. *Cytotherapy* 2020; **22**: 261-275 [PMID: 32247542 DOI: 10.1016/j.jcyt.2020.02.005]
 - 23 **Hajmoussa G**, Przybyl E, Pfister F, Paredes-Juarez GA, Moganti K, Busch S, Kuipers J, Klaassen I, van Luyn MJA, Krenning G, Hammes HP, Harmsen MC. Human adipose tissue-derived stromal cells act as functional pericytes in mice and suppress high-glucose-induced proinflammatory activation of bovine retinal endothelial cells. *Diabetologia* 2018; **61**: 2371-2385 [PMID: 30151615 DOI: 10.1007/s00125-018-4713-0]
 - 24 **Terlizzi V**, Kolibabka M, Burgess JK, Hammes HP, Harmsen MC. The Pericytic Phenotype of Adipose Tissue-Derived Stromal Cells Is Promoted by NOTCH2. *Stem Cells* 2018; **36**: 240-251 [PMID: 29067740 DOI: 10.1002/stem.2726]
 - 25 **Kicic A**, Shen WY, Wilson AS, Constable IJ, Robertson T, Rakoczy PE. Differentiation of marrow stromal cells into photoreceptors in the rat eye. *J Neurosci* 2003; **23**: 7742-7749 [PMID: 12944502 DOI: 10.1523/JNEUROSCI.23-21-07742.2003]
 - 26 **Ezquer M**, Urzua CA, Montecino S, Leal K, Conget P, Ezquer F. Intravitreal administration of multipotent mesenchymal stromal cells triggers a cytoprotective microenvironment in the retina of diabetic mice. *Stem Cell Res Ther* 2016; **7**: 42 [PMID: 26983784 DOI: 10.1186/s13287-016-0299-y]
 - 27 **Rajashekhhar G**, Ramadan A, Abburi C, Callaghan B, Traktuev DO, Evans-Molina C, Maturi R, Harris A, Kern TS, March KL. Regenerative therapeutic potential of adipose stromal cells in early stage diabetic retinopathy. *PLoS One* 2014; **9**: e84671 [PMID: 24416262 DOI: 10.1371/journal.pone.0084671]
 - 28 **Cronk SM**, Kelly-Goss MR, Ray HC, Mendel TA, Hoehn KL, Bruce AC, Dey BK, Guendel AM, Tavakol DN, Herman IM, Peirce SM, Yates PA. Adipose-derived stem cells from diabetic mice show impaired vascular stabilization in a murine model of diabetic retinopathy. *Stem Cells Transl Med* 2015; **4**: 459-467 [PMID: 25769654 DOI: 10.5966/sctm.2014-0108]
 - 29 **Harrell CR**, Jovicic N, Djonov V, Arsenijevic N, Volarevic V. Mesenchymal Stem Cell-Derived Exosomes and Other Extracellular Vesicles as New Remedies in the Therapy of Inflammatory Diseases. *Cells* 2019; **8** [PMID: 31835680 DOI: 10.3390/cells8121605]
 - 30 **Li W**, Jin L, Cui Y, Nie A, Xie N, Liang G. Bone marrow mesenchymal stem cells-induced exosomal microRNA-486-3p protects against diabetic retinopathy through TLR4/NF- κ B axis repression. *J Endocrinol Invest* 2021; **44**: 1193-1207 [PMID: 32979189 DOI: 10.1007/s40618-020-01405-3]
 - 31 **Wang J**, Zhang W, He GH, Wu B, Chen S. Transfection with CXCR4 potentiates homing of mesenchymal stem cells *in vitro* and therapy of diabetic retinopathy *in vivo*. *Int J Ophthalmol* 2018; **11**: 766-772 [PMID: 29862173 DOI: 10.18240/ijo.2018.05.08]
 - 32 **Lin W**, Xu G. Over-expression of CNTF in bone marrow mesenchymal stem cells protects RPE cells from short-wavelength, blue-light injury. *In Vitro Cell Dev Biol Anim* 2018; **54**: 355-365 [PMID: 29564604 DOI: 10.1007/s11626-018-0243-9]
 - 33 **Kim JY**, Park S, Park SH, Lee D, Kim GH, Noh JE, Lee KJ, Kim GJ. Overexpression of pigment epithelium-derived factor in placenta-derived mesenchymal stem cells promotes mitochondrial biogenesis in retinal cells. *Lab Invest* 2021; **101**: 51-69 [PMID: 32724163 DOI: 10.1038/s41374-020-0470-z]
 - 34 **Gu X**, Yu X, Zhao C, Duan P, Zhao T, Liu Y, Li S, Yang Z, Li Y, Qian C, Yin Z, Wang Y. Efficacy and Safety of Autologous Bone Marrow Mesenchymal Stem Cell Transplantation in Patients with Diabetic Retinopathy. *Cell Physiol Biochem* 2018; **49**: 40-52 [PMID: 30134223 DOI: 10.1159/000492838]
 - 35 **Yücel Y**. Central nervous system changes in glaucoma. *J Glaucoma* 2013; **22** Suppl 5: S24-S25 [PMID: 23733121 DOI: 10.1097/IJG.0b013e3182934a55]
 - 36 **Almasieh M**, Wilson AM, Morquette B, Cueva Vargas JL, Di Polo A. The molecular basis of retinal ganglion cell death in glaucoma. *Prog Retin Eye Res* 2012; **31**: 152-181 [PMID: 22155051 DOI: 10.1016/j.preteyeres.2011.11.002]
 - 37 **Hu Y**, Tan HB, Wang XM, Rong H, Cui HP, Cui H. Bone marrow mesenchymal stem cells protect against retinal ganglion cell loss in aged rats with glaucoma. *Clin Interv Aging* 2013; **8**: 1467-1470 [PMID: 24204132 DOI: 10.2147/CIA.S47350]
 - 38 **Mead B**, Amaral J, Tomarev S. Mesenchymal Stem Cell-Derived Small Extracellular Vesicles Promote Neuroprotection in Rodent Models of Glaucoma. *Invest Ophthalmol Vis Sci* 2018; **59**: 702-714 [PMID: 29392316 DOI: 10.1167/iovs.17-22855]
 - 39 **Ji S**, Xiao J, Liu J, Tang S. Human Umbilical Cord Mesenchymal Stem Cells Attenuate Ocular Hypertension-Induced Retinal Neuroinflammation via Toll-Like Receptor 4 Pathway. *Stem Cells Int* 2019; **2019**: 9274585 [PMID: 31737079 DOI: 10.1155/2019/9274585]
 - 40 **Downs JC**, Girkin CA. Lamina cribrosa in glaucoma. *Curr Opin Ophthalmol* 2017; **28**: 113-119 [PMID: 27898470 DOI: 10.1097/ICU.0000000000000354]
 - 41 **Yun H**, Zhou Y, Wills A, Du Y. Stem Cells in the Trabecular Meshwork for Regulating Intraocular Pressure. *J Ocul Pharmacol Ther* 2016; **32**: 253-260 [PMID: 27183473 DOI: 10.1089/jop.2016.0005]
 - 42 **Osborne A**, Sanderson J, Martin KR. Neuroprotective Effects of Human Mesenchymal Stem Cells and Platelet-Derived Growth Factor on Human Retinal Ganglion Cells. *Stem Cells* 2018; **36**: 65-78 [PMID: 29044808 DOI: 10.1002/stem.2722]
 - 43 **Li Y**, He X, Li J, Ni F, Sun Q, Zhou Y. Proliferation and differentiation of direct coculture of bone marrow mesenchymal stem cells and pigmented cells from the ciliary margin. *Mol Med Rep* 2017; **15**:

- 3529-3534 [PMID: [28440470](#) DOI: [10.3892/mmr.2017.6481](#)]
- 44 **Manuguerra-Gagné R**, Boulos PR, Ammar A, Leblond FA, Krosi G, Pichette V, Lesk MR, Roy DC. Transplantation of mesenchymal stem cells promotes tissue regeneration in a glaucoma model through laser-induced paracrine factor secretion and progenitor cell recruitment. *Stem Cells* 2013; **31**: 1136-1148 [PMID: [23495088](#) DOI: [10.1002/stem.1364](#)]
 - 45 **Roubeix C**, Godefroy D, Mias C, Sapienza A, Riancho L, Degardin J, Fradot V, Ivkovic I, Picaud S, Sennlaub F, Denoyer A, Rostene W, Sahel JA, Parsadaniantz SM, Brignole-Baudouin F, Baudouin C. Intraocular pressure reduction and neuroprotection conferred by bone marrow-derived mesenchymal stem cells in an animal model of glaucoma. *Stem Cell Res Ther* 2015; **6**: 177 [PMID: [26377305](#) DOI: [10.1186/s13287-015-0168-0](#)]
 - 46 **Lo Furno D**, Graziano AC, Caggia S, Perrotta RE, Tarico MS, Giuffrida R, Cardile V. Decrease of apoptosis markers during adipogenic differentiation of mesenchymal stem cells from human adipose tissue. *Apoptosis* 2013; **18**: 578-588 [PMID: [23479126](#) DOI: [10.1007/s10495-013-0830-x](#)]
 - 47 **Johnson TV**, Bull ND, Hunt DP, Marina N, Tomarev SI, Martin KR. Neuroprotective effects of intravitreal mesenchymal stem cell transplantation in experimental glaucoma. *Invest Ophthalmol Vis Sci* 2010; **51**: 2051-2059 [PMID: [19933193](#) DOI: [10.1167/iovs.09-4509](#)]
 - 48 **Johnson TV**, DeKorver NW, Levasseur VA, Osborne A, Tassoni A, Lorber B, Heller JP, Villasmil R, Bull ND, Martin KR, Tomarev SI. Identification of retinal ganglion cell neuroprotection conferred by platelet-derived growth factor through analysis of the mesenchymal stem cell secretome. *Brain* 2014; **137**: 503-519 [PMID: [24176979](#) DOI: [10.1093/brain/awt292](#)]
 - 49 **Su W**, Li Z, Jia Y, Zhu Y, Cai W, Wan P, Zhang Y, Zheng SG, Zhuo Y. microRNA-21a-5p/PDCD4 axis regulates mesenchymal stem cell-induced neuroprotection in acute glaucoma. *J Mol Cell Biol* 2017; **9**: 289-301 [PMID: [28655163](#) DOI: [10.1093/jmcb/mjx022](#)]
 - 50 **Vilela CAP**, Messias A, Calado RT, Siqueira RC, Silva MJL, Covas DT, Paula JS. Retinal function after intravitreal injection of autologous bone marrow-derived mesenchymal stromal cells in advanced glaucoma. *Doc Ophthalmol* 2021 [PMID: [33469852](#) DOI: [10.1007/s10633-021-09817-z](#)]
 - 51 **Pagon RA**. Retinitis pigmentosa. *Surv Ophthalmol* 1988; **33**: 137-177 [PMID: [3068820](#) DOI: [10.1016/0039-6257\(88\)90085-9](#)]
 - 52 **Hamel C**. Retinitis pigmentosa. *Orphanet J Rare Dis* 2006; **1**: 40 [PMID: [17032466](#) DOI: [10.1186/1750-1172-1-40](#)]
 - 53 **Hartong DT**, Berson EL, Dryja TP. Retinitis pigmentosa. *Lancet* 2006; **368**: 1795-1809 [PMID: [17113430](#) DOI: [10.1016/S0140-6736\(06\)69740-7](#)]
 - 54 **Aït-Ali N**, Fridlich R, Millet-Puel G, Clérin E, Delalande F, Jaillard C, Blond F, Perrocheau L, Reichman S, Byrne LC, Olivier-Bandini A, Bellalou J, Moysse E, Bouillaud F, Nicol X, Dalkara D, van Dorsselaer A, Sahel JA, Léveillard T. Rod-derived cone viability factor promotes cone survival by stimulating aerobic glycolysis. *Cell* 2015; **161**: 817-832 [PMID: [25957687](#) DOI: [10.1016/j.cell.2015.03.023](#)]
 - 55 **Murakami Y**, Ikeda Y, Nakatake S, Miller JW, Vavvas DG, Sonoda KH, Ishibashi T. Necrotic cone photoreceptor cell death in retinitis pigmentosa. *Cell Death Dis* 2015; **6**: e2038 [PMID: [26720347](#) DOI: [10.1038/cddis.2015.385](#)]
 - 56 **Campochiaro PA**, Mir TA. The mechanism of cone cell death in Retinitis Pigmentosa. *Prog Retin Eye Res* 2018; **62**: 24-37 [PMID: [28962928](#) DOI: [10.1016/j.preteyeres.2017.08.004](#)]
 - 57 **Moreno ML**, Mérida S, Bosch-Morell F, Miranda M, Villar VM. Autophagy Dysfunction and Oxidative Stress, Two Related Mechanisms Implicated in Retinitis Pigmentosa. *Front Physiol* 2018; **9**: 1008 [PMID: [30093867](#) DOI: [10.3389/fphys.2018.01008](#)]
 - 58 **Nowak JZ**. Oxidative stress, polyunsaturated fatty acids-derived oxidation products and bisretinoids as potential inducers of CNS diseases: focus on age-related macular degeneration. *Pharmacol Rep* 2013; **65**: 288-304 [PMID: [23744414](#) DOI: [10.1016/s1734-1140\(13\)71005-3](#)]
 - 59 **He Y**, Zhang Y, Liu X, Ghazaryan E, Li Y, Xie J, Su G. Recent advances of stem cell therapy for retinitis pigmentosa. *Int J Mol Sci* 2014; **15**: 14456-14474 [PMID: [25141102](#) DOI: [10.3390/ijms150814456](#)]
 - 60 **Tucker BA**, Mullins RF, Stone EM. Stem cells for investigation and treatment of inherited retinal disease. *Hum Mol Genet* 2014; **23**: R9-R16 [PMID: [24647603](#) DOI: [10.1093/hmg/ddu124](#)]
 - 61 **Oner A**, Gonen ZB, Sinim N, Cetin M, Ozkul Y. Subretinal adipose tissue-derived mesenchymal stem cell implantation in advanced stage retinitis pigmentosa: a phase I clinical safety study. *Stem Cell Res Ther* 2016; **7**: 178 [PMID: [27906070](#) DOI: [10.1186/s13287-016-0432-y](#)]
 - 62 **Limoli PG**, Limoli CSS, Morales MU, Vingolo EM. Mesenchymal stem cell surgery, rescue and regeneration in retinitis pigmentosa: clinical and rehabilitative prognostic aspects. *Restor Neurol Neurosci* 2020; **38**: 223-237 [PMID: [32310198](#) DOI: [10.3233/RNN-190970](#)]
 - 63 **Zhao PT**, Zhang LJ, Shao H, Bai LL, Yu B, Su C, Dong LJ, Liu X, Li XR, Zhang XM. Therapeutic effects of mesenchymal stem cells administered at later phase of recurrent experimental autoimmune uveitis. *Int J Ophthalmol* 2016; **9**: 1381-1389 [PMID: [27803852](#) DOI: [10.18240/ijo.2016.10.03](#)]
 - 64 **Limoli PG**, Vingolo EM, Limoli C, Scalinci SZ, Nebbioso M. Regenerative Therapy by Suprachoroidal Cell Autograft in Dry Age-related Macular Degeneration: Preliminary In Vivo Report. *J Vis Exp* 2018 [PMID: [29553543](#) DOI: [10.3791/56469](#)]
 - 65 **Huo DM**, Dong FT, Yu WH, Gao F. Differentiation of mesenchymal stem cell in the microenvironment of retinitis pigmentosa. *Int J Ophthalmol* 2010; **3**: 216-219 [PMID: [22553557](#) DOI: [10.3980/j.issn.2222-3959.2010.03.08](#)]
 - 66 **Zarbin M**. Cell-Based Therapy for Degenerative Retinal Disease. *Trends Mol Med* 2016; **22**: 115-134

- [PMID: 26791247 DOI: 10.1016/j.molmed.2015.12.007]
- 67 **Salehi H**, Amirpour N, Razavi S, Esfandiari E, Zavar R. Overview of retinal differentiation potential of mesenchymal stem cells: A promising approach for retinal cell therapy. *Ann Anat* 2017; **210**: 52-63 [PMID: 27986614 DOI: 10.1016/j.aanat.2016.11.010]
- 68 **Soleimannejad M**, Ebrahimi-Barough S, Nadri S, Riazi-Esfahani M, Soleimani M, Tavangar SM, Ai J. Retina tissue engineering by conjunctiva mesenchymal stem cells encapsulated in fibrin gel: Hypotheses on novel approach to retinal diseases treatment. *Med Hypotheses* 2017; **101**: 75-77 [PMID: 28351499 DOI: 10.1016/j.mehy.2017.02.019]
- 69 **Arnhold S**, Absenger Y, Klein H, Addicks K, Schraermeyer U. Transplantation of bone marrow-derived mesenchymal stem cells rescue photoreceptor cells in the dystrophic retina of the rhodopsin knockout mouse. *Graefes Arch Clin Exp Ophthalmol* 2007; **245**: 414-422 [PMID: 16896916 DOI: 10.1007/s00417-006-0382-7]
- 70 **Wang S**, Lu B, Girman S, Duan J, McFarland T, Zhang QS, Grompe M, Adamus G, Appukuttan B, Lund R. Non-invasive stem cell therapy in a rat model for retinal degeneration and vascular pathology. *PLoS One* 2010; **5**: e9200 [PMID: 20169166 DOI: 10.1371/journal.pone.0009200]
- 71 **Yu B**, Shao H, Su C, Jiang Y, Chen X, Bai L, Zhang Y, Li Q, Zhang X, Li X. Exosomes derived from MSCs ameliorate retinal laser injury partially by inhibition of MCP-1. *Sci Rep* 2016; **6**: 34562 [PMID: 27686625 DOI: 10.1038/srep34562]
- 72 **Usategui-Martín R**, Puertas-Neyra K, García-Gutiérrez MT, Fuentes M, Pastor JC, Fernandez-Bueno I. Human Mesenchymal Stem Cell Secretome Exhibits a Neuroprotective Effect over *In Vitro* Retinal Photoreceptor Degeneration. *Mol Ther Methods Clin Dev* 2020; **17**: 1155-1166 [PMID: 32514411 DOI: 10.1016/j.omtm.2020.05.003]
- 73 **Ding SLS**, Koh AE, Kumar S, Ali Khan MS, Alzahrani B, Mok PL. Genetically-modified human mesenchymal stem cells to express erythropoietin enhances differentiation into retinal photoreceptors: An in-vitro study. *J Photochem Photobiol B* 2019; **195**: 33-38 [PMID: 31060031 DOI: 10.1016/j.jphotobiol.2019.04.008]
- 74 **Mahmoudian-Sani MR**, Forouzanfar F, Asgharzade S, Ghorbani N. Overexpression of MiR-183/96/182 Triggers Retina-Like Fate in Human Bone Marrow-Derived Mesenchymal Stem Cells (hBMSCs) in Culture. *J Ophthalmol* 2019; **2019**: 2454362 [PMID: 31885884 DOI: 10.1155/2019/2454362]
- 75 **Rahmani A**, Naderi M, Barati G, Arefian E, Jedari B, Nadri S. The potency of hsa-miR-9-1 overexpression in photoreceptor differentiation of conjunctiva mesenchymal stem cells on a 3D nanofibrous scaffold. *Biochem Biophys Res Commun* 2020; **529**: 526-532 [PMID: 32736669 DOI: 10.1016/j.bbrc.2020.06.006]
- 76 **Machalińska A**, Kawa M, Pius-Sadowska E, Stępniewski J, Nowak W, Rogińska D, Kaczyńska K, Baumert B, Wiszniewska B, Józkowicz A, Dulak J, Machaliński B. Long-term neuroprotective effects of NT-4-engineered mesenchymal stem cells injected intravitreally in a mouse model of acute retinal injury. *Invest Ophthalmol Vis Sci* 2013; **54**: 8292-8305 [PMID: 24265016 DOI: 10.1167/iovs.13-12221]
- 77 **Tuekprakhon A**, Sangkitporn S, Trinavarat A, Pawestri AR, Vamvanij V, Ruangchainikom M, Luksanaprukpa P, Pongpaksupasin P, Khorchai A, Dambua A, Boonchu P, Yodtup C, Uiprasertkul M, Atchaneeyasakul LO. Intravitreal autologous mesenchymal stem cell transplantation: a non-randomized phase I clinical trial in patients with retinitis pigmentosa. *Stem Cell Res Ther* 2021; **12**: 52 [PMID: 33422139 DOI: 10.1186/s13287-020-02122-7]
- 78 **Kahraman NS**, Oner A. Umbilical cord derived mesenchymal stem cell implantation in retinitis pigmentosa: a 6-month follow-up results of a phase 3 trial. *Int J Ophthalmol* 2020; **13**: 1423-1429 [PMID: 32953582 DOI: 10.18240/ijo.2020.09.14]
- 79 **Zhao T**, Liang Q, Meng X, Duan P, Wang F, Li S, Liu Y, Yin ZQ. Intravenous Infusion of Umbilical Cord Mesenchymal Stem Cells Maintains and Partially Improves Visual Function in Patients with Advanced Retinitis Pigmentosa. *Stem Cells Dev* 2020; **29**: 1029-1037 [PMID: 32679004 DOI: 10.1089/scd.2020.0037]



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